



#### AperTO - Archivio Istituzionale Open Access dell'Università di Torino

#### **Fate of regulated, masked, emerging mycotoxins and secondary fungal metabolites during different large-scale maize dry-milling processes**

#### **This is a pre print version of the following article:**

Original Citation:

Availability:

This version is available http://hdl.handle.net/2318/1796900 since 2021-08-13T19:04:41Z

Published version:

DOI:10.1016/j.foodres.2020.109861

Terms of use:

Open Access

Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)

# **FOOD RESEARCH INTERNATIONAL**

# **Fate of Regulated, Masked, Emerging Mycotoxins and Secondary Fungal Metabolites during different large-scale maize dry-milling processes**

- 
- Authors:

7 Valentina Scarpino<sup>a</sup>, Francesca Vanara<sup>a</sup>, Micheal Sulyok<sup>b</sup>, Rudolf Krska<sup>b</sup>, Massimo Blandino<sup>a\*</sup> 

Affiliation:

<sup>a</sup> University of Turin, Department of Agricultural, Forest and Food Sciences, Largo Paolo Braccini

- 2, 10095 Grugliasco (TO), Italy.
- <sup>b</sup> University of Natural Resources and Life Sciences, Vienna (BOKU), Center for Analytical
- Chemistry, Department of Agrobiotechnology (IFA-Tulln), Konrad-Lorenz-Str. 20, Tulln 3430,
- Austria.
- 
- \*Corresponding author: massimo.blandino@unito.it
- Phone: 00390116708895; Fax: 00390116708798
- 

#### **Abstract**

 The worldwide consumption of maize for food is increasing, since it is used as an ingredient for several foods and in particular for gluten-free products, whose consumption is rising.

 In temperate areas, the main limitation to the use of maize in the food chain is its contamination by mycotoxins. Limited information is available on the fate of masked, modified and emerging mycotoxins or of other secondary fungal metabolites in maize products and by-products. For this reason, 3 maize lots, obtained in different growing seasons, were processed using two different degermination processes, a dry-degermination system or a tempering-degermination one, in order to compare the interaction between mycotoxins and the dry-milling management system. Whole grain before and after cleaning, and all the products and the by-products were sampled twice for each lot and were subjected to a multi-mycotoxin LC-MS/MS analysis. More than 30 mycotoxins and other fungal metabolites, including masked or modified forms, co-occurred in all the maize milling fractions. Grain cleaning reduced all the detected fungal metabolites by 1.2-2 times, compared to the grain before cleaning. Animal feed flour showed the highest content of almost all the mycotoxins and fungal metabolites, with a consequent negative impact on animal health. Overall, the sum of the 3 food-grade endosperm fractions from tempering-degermination (flaking grits, medium and small hominy grits) resulted in a lower contamination than those obtained from the dry-degermination (pearl meal, break meal and maize flour). Moreover, considering that for all the mycotoxins and fungal metabolites an inverse relationship with particle size was observed, flaking grits represented the healthiest maize products with the least contamination level, while the abatement was always lower for maize flour.

 Furthermore, the metabolites were variably redistributed in the maize fractions. The total aflatoxins, kojic acid, deoxynivalenol and its modified form, culmorin, and its associated forms, butenolide, fusaproliferin, fusaric acid, fusarinolic acid and, in some cases, zearalenone and its modified forms, and fusarin C were found to be concentrated significantly in the germ. Moreover, the total

 aflatoxins, deoxynivalenol-3-glucoside, fusarinolic acid, fusarin C, moniliformin and butenolide had a greater permanence in the maize food fractions and a weaker decontamination, both of which point to a higher risk of exposure for the end consumers.

 The co-occurrence of a such a high number of mycotoxins and fungal metabolites and their different fates during the dry-milling process have never been described before and could be useful for future risk assessment studies to correctly assess the risk of exposure to such substances. Moreover, the continuous exposure to these mycotoxins and fungal metabolites should be considered in particular for consumers in the many parts of the world where maize is a staple food, and where it is used for the baby food supply chain and for the celiac population in developed countries, due to the high consumption of maize gluten-free products.

- 
- 
- 
- 
- 

 **KEYWORDS:** aurofusarin; beauvericin; culmorin; deoxynivalenol-3-glucoside; fusaric acid; fusarin C; moniliformin; zearalenone-sulphate.

#### **ABBREVIATIONS**

 3-ADON, 3-acetyldeoxynivalenol; 15-ADON, 15-acetyldeoxynivalenol; 5-OH-CULM, 5-hydroxy- culmorin; 15-OH-CULM, 15-hydroxy-culmorin; 15-OH-culmoron, 15-hydroxy-culmoron; α-ZEA- ol, alpha-zearalenol; β-ZEA-ol, beta-zearalenol; AFs, Aflatoxins; AFTOT, Total aflatoxins, sum of AFB1, AFB2, AFG1 and AFG2; ALS, altersetin; AME, alternariol methyl ether; ANOVA, Analysis of variance; AOH, alternariol; AUR, aurofusarin; BEA, beauvericin; BIK, bikaverin; BUT, 67 butenolide; CAC, Codex Alimentarius Commission; CULM, culmorin; CULM<sub>TOT</sub>, Total culmorin forms, sum of CULM, 5-OH-CULM, 15-OH-CULM, 15-OH-culmuron; DAS, diacetoxyscirpenol; DD, Dry-Degermination; DON, Deoxynivalenol; DON-3-G, deoxynivalenol-3-glucoside; DONTOT, Total deoxynivalenol forms, sum of DON, DON-3-G, 3-ADON and 15-ADON; EC, European Commission; EFSA, European Food Safety Authority; ENNs, enniatins; ENNTOT, Total enniatins, 72 sum of ENN A,  $A_1$ , B and B<sub>1</sub>; EQU, equisetin; ESI, Electrospray Ionization; FA<sub>TOT</sub>, Total 73 fumonisins A, sum of FA<sub>1</sub>, FA<sub>2</sub>; FBs, Fumonisins B; FB<sub>TOT</sub>, Total fumonisins B, sum of FB<sub>1</sub>, FB<sub>2</sub>, FB3 and FB4; FnA, fusarinolic acid; FSA, fusaric acid; FUS, fusaproliferin; HFB1, hydorlized 75 fumonin B<sub>1</sub>; JECFA, Joint FAO/WHO Expert Committee on Food Additives; LD<sub>50</sub>, Lethal Dose 50%; LOD, limit of detection; LOQ, limit of quantification; MON, moniliformin; NIV, nivalenol; OTA, ochratoxin A; REGWF, Ryan-Einot-Gabriel-Welsh F post-hoc test; SD, Standard Deviation; TD, Tempering-Degermination; TeA, tenuazonic acid; TEN, tentoxin; ZEA, Zearalenone; ZEA-S, zearalenone-sulphate; ZEATOT, total zearalenone forms sum of ZEA, ZEA-S, α-ZEA-ol and β-ZEA-ol.

### *1. Introduction*

 Maize is the main cereal grain produced worldwide, although it ranks third as a staple food, after wheat and rice. The consumption of this crop has recently increased in developed countries, as it is used as an ingredient for breakfast cereals, snacks, dietetic products, and in particular for baby food and gluten-free food formulations, whose consumption is rising (Rai et al., 2018).

 Unfortunately, maize can be colonised competitively by several spoilage fungi of the *Fusarium*, *Aspergillus*, *Alternaria* and *Penicillium* species, which are capable of producing a large variety of mycotoxins and other secondary fungal metabolites as a result of fungal ear rot on maize ears (Marin et al., 2012), which in turn lead to a negative impact on the safety and quality of this agricultural commodity. In this regard, a recent worldwide study on the contamination of food- crops with mycotoxins has pointed out that 60-80% of food crops are contaminated with mycotoxins (Eskola et al., 2019).

 Approximately 400 mycotoxins or potential risky fungal metabolites are known to date throughout the world (Berthiller et al., 2007), but aflatoxins (AFs), fumonisins B (FBs), deoxynivalenol (DON), zearalenone (ZEA) and ochratoxin A (OTA) are the only mycotoxins that are generally regulated and monitored (Binder, 2007). The other mycotoxins, which are less known from a scientific point of view and which may co-occur with the regulated mycotoxins, have become part of the so-called "masked", "modified" and "emerging" mycotoxins or other secondary fungal metabolites (Streit et al., 2013). Masked mycotoxins are plant metabolites of mycotoxins, or according to Rychlik et al.'s (2014) systematic definition "biologically modified" mycotoxins, whose chemical modifications, introduced by the plant's metabolism, have the potential to affect both their toxicity and analytical detectability. Among the group of masked mycotoxins, deoxynivalenol-3-glucoside (DON-3-G) and zearalenone-sulphate (ZEA-S) and are the most commonly found in food and feeds. Their toxicological properties are currently being investigated, and mainly involve the conversion of DON-3-G to DON and ZEA-S to ZEA by microbiota of the

 intestinal tract (Dall'Erta et al., 2013). Emerging mycotoxins are a group of chemically diverse mycotoxins, for which, to date, no regulations exist, and ongoing risk assessment studies are still in progress. Aflatoxin precursors, ergot alkaloids, enniatins (ENNs), beauvericin (BEA) and moniliformin (MON) are those that are more commonly mentioned in this group (Jestoi, 2008). Moreover, there is no clear indication of the toxicity of the other secondary fungal metabolites that are frequently found in cereals, such as aurofusarin (AUR) and culmorin (CULM), and they are still 112 the subject of detailed studies.

 Since little is known about the toxicological effects of these compounds and limited information is available about the synergistic or additive toxic effects related to their co-presence with the regulated mycotoxins, a higher risk of exposure for the end consumers and health issues could emerge.

 Dry-milling is the main industrial process adopted in the maize food chain to obtain hominy grits, maize flours and meals for human consumption. This technology consists of a mechanical kernel processing that creates whole or fractionated products, separated according to their anatomical features, such as bran, germ and endosperm (Gwirtz & Garcia-Casal, 2014). Because of the important role of dry milling in re-distributing contaminants in the different milling products and by-products, several scientific contributions have focused on the fate of the main regulated mycotoxins, such as fumonisins, aflatoxins, deoxynivalenol and zearalenone (Scudamore & Patel, 2000; Brera et al., 2004, 2006; Bullerman & Bianchini, 2007; Castells et al., 2008; Schollenberger et al., 2008; Pietri et al., 2009; Vanara et al., 2009; Burger et al., 2013; Aprodu & Banu, 2015; Bordini et al., 2017; Vanara et al., 2018). Furthermore, there is a lack of information on the fate of masked, modified and emerging mycotoxins and on other secondary fungal metabolites in maize products and by-products (Schollenberger et al., 2008; Scarpino et al., 2020).

 To the best of the authors' knowledge, the simultaneous fate and re-distribution of such a high number of mycotoxins, including the regulated, masked, modified, emerging mycotoxins and other secondary fungal metabolites, in maize destined for human consumption, through the application of

 the dry-milling process, has not yet been considered in the scientific literature. Moreover, the European Food Safety Authority (EFSA) is continuously engaged in collecting the occurrence data of masked, modified and emerging mycotoxins in food and feeds, in order to establish scientific opinions on their risks for human and animal health. Information on the fate of these contaminants, throughout the supply chain, is an essential information to carry out future risk assessments based on the real exposure of humans and animals, from the raw materials to the final food and feed products.

 For this purpose, 3 maize lots, obtained in different growing seasons, were processed using two different degermination processes, a dry-degermination (DD) system and a tempering- degermination (TD) one, in order to compare the interaction between mycotoxins and the adopted dry-milling management process.

### *2. Material and methods*

### *2.1 Maize milling processes and sampling*

 The occurrence and distribution of regulated, masked, emerging mycotoxins and secondary fungal metabolites have been investigated by sampling and analysing in 3 different growing seasons (2012, 2013 and 2014), in the same growing area (North West Italy, the province of Turin), a single maize hybrid each year from 3 commercial lots (Pioneer P1547 in 2012 and 2013, Pioneer P0722 in 2014), for food dry milling purposes.

 The maize from each lot was milled in two separate dry-milling industrial lines, which were based on different degermination processes. The first line consisted of a dry-milling technology, coupled to a dry-degermination (DD) system, while the dry-milling technology in the second line was based on a tempering-degermination (TD) process. The two processes have been described in detail by Blandino et al. (2017a).

 Germs and animal feed flour were the main by-products of both processes, and they have expected yields of 10% and 35%, respectively. The maize products of the 3 lots recorded mean yields of 5%, 20% and 30% for maize flour, break meal and pearl meal during the DD process and of 7%, 19% and 29% for small, medium and flaking grits, whose different particle sizes are shown in Figure 1, during the TD process.

 The sampled products of each process represented a lot of origin of about 200 t and were collected during the milling process according to European Commission Regulation (EC) No 401/2006. An aggregate sample was obtained for each milling fraction by carefully blending 40 incremental samples, of 100 g each, which were collected, by means of a dynamic sampling procedure, from opening slits of the plant for a period of 1 hour at regular intervals. All the maize products and by- products of each lot were sampled twice, before and after cleaning, and were collected from both processes (DD and TD), for a total of 72 samples.

167 The samples were stored at -18°C until the multi-mycotoxin analysis was performed.

#### *2.2 Multi-mycotoxin LC-MS/MS analysis*

 The samples were prepared according to Sulyok et al. (2006). The chromatographic and mass spectrometric parameters of the investigated analytes were described by Malachova et al. in 2014. Quantification was performed on the basis of an external calibration, and the results were corrected for apparent recoveries, as determined in the maize. Fumonisins A were semi-quantified using the response of FB2. The accuracy of the method was verified by participating in proficiency testing schemes organised by BIPEA (Gennevilliers, France), with 160 out of the 168 results submitted for maize and maize-based feeds exhibiting a z-score of between -2 and 2.

#### *2.3 Statistical analysis*

 An analysis of variance (ANOVA) was run for each maize lot to compare the mycotoxin contaminations. The raw kernel and the milling fractions of the two dry milling processes (TD and

- DD) were considered as the independent variables. The mycotoxin concentrations were transformed 180 using the y'=ln(x+1) equation to normalise the residuals. Multiple comparison tests were carried out, according to the Ryan-Einot-Gabriel-Welsh F (REGWF) post-hoc test, on the mycotoxin contamination means of the different dry-milling fractions.
- SPSS Version 24.0 of the Windows statistical package, (SPSS Inc., 2017) was used for the
- statistical analysis.
- 

### *3. Results and Discussion*

 As reported in Table 1, the following main regulated, masked, modified, emerging mycotoxins and other secondary fungal metabolites were simultaneously detected in the pre-cleaned whole grain from the maize from the 3 lots processed in the industrial mill during the 2012-2014 period: 190 fumonisins B (total fumonisins  $B = FB_{TOT} =$  the sum of  $FB_1$ ,  $FB_2$ ,  $FB_3$  and  $FB_4$ ); fumonisins A 191 (total fumonisins  $A = FA_{TOT}$  = the sum of  $FA_1$ ,  $FA_2$ ); hydorlyzed fumonin  $B_1$  (HFB<sub>1</sub>); fusaric acid (FSA); fusarinolic acid (FnA); fusarin C; bikaverin (BIK); moniliformin (MON); beauvericin 193 (BEA); fusaproliferin (FUS); enniatins (total enniatins =  $ENN<sub>TOT</sub>$  = the sum of ENN A, A<sub>1</sub>, B and 194 B<sub>1</sub>); total deoxynivalenol forms ( $DON<sub>TOT</sub>$  = the sum of deoxynivalenol ( $DON$ ), deoxynivalenol-3- glucoside (DON-3-G), 3-acetyldeoxynivalenol (3-ADON) and 15-acetyldeoxynivalenol (15- 196 ADON)); total zearalenone forms ( $ZEA<sub>TOT</sub>$  = the sum of zearalenone ( $ZEA$ ), zearalenone-sulphate 197 (ZEA-S), alpha-zearalenol (α-ZEA-ol) and beta-zearalenol (β-ZEA-ol)); total culmorin (CULM<sub>TOT</sub>) = the sum of culmorin (CULM), 5-hydroxy-culmorin (5-OH-CULM), 15-hydroxy-culmorin (15- OH-CULM) and 15-hydroxy-culmoron (15-OH-culmoron)); aurofusarin (AUR); butenolide (BUT); diacetoxyscirpenol (DAS); nivalenol (NIV); equisetin (EQU); T-2 toxin; HT-2 toxin; aflatoxins 201 (total aflatoxins =  $AF_{TOT}$  = the sum of  $AFB_1$ ,  $AFB_2$ ,  $AFG_1$  and  $AFG_2$ ); kojic acid; alternariol (AOH); alternariol methyl ether (AME); tentoxin (TEN); tenuazonic acid (TeA); altersetin (ALS). As reported in Table 1, the contamination levels of the different mycotoxins and fungal metabolites may vary significantly from year to year in maize and mainly depend on the environmental conditions of each year, which have an impact on the production of these co-occurring compounds by the main fugal causal agents of diseases on maize (Blandino et al. 2017b).

207 The fate of  $FB_{TOT}$  in the milling fractions of the different dry milling processes and maize lots (2012, 2013 and 2014) is reported in Figure 2. On average, in all of the lots and for all the fractions, 209 FB<sub>1</sub> was about the 68% of the FB<sub>TOT</sub>, FB<sub>2</sub> was 16%, FB<sub>3</sub> was 9% and FB<sub>4</sub> was 7%. The cleaning 210 step on average reduced the FB<sub>TOT</sub> content by -47%. Overall, the animal feed flour represented the 211 fraction with the highest  $FB_{TOT}$  content for all the lots, with a significant increase, that is, of 3.0 and 212 2.8 times, respectively, in the DD and TD processes, compared to the corresponding pre-cleaned whole grain. The germ significantly differed from the pre-cleaned whole grain for all the lots, with the exception of the germ from the DD process of the 2012 lot, and showed mean reductions of - 58% (DD) and -45% (TD), with no significant differences between the processes. Within endosperm products, the maize flour, the break meal and the pearl meal (DD process) showed an FBTOT decrease, compared to the pre-cleaned whole grain, of -24%, -78% and -80%, respectively, while the small, medium hominy grits and flaking grits (TD process) showed decreases of -81% and -89% and -95%, respectively. Decontamination was greater in TD process than in the DD one and an inverse correlation with the milling fraction particle size was observed for both processes.

 Several studies have been conducted on the distribution of fumonisins in dry-milled maize fractions (Katta et al., 1997; Scudamore & Patel, 2000; Broggi et al., 2002; Brera et al., 2004; Bullerman & Bianchini, 2007; Castells et al., 2008; Pietri et al., 2009; Vanara et al., 2009; Burger et al., 2013; Aprodu & Banu, 2015; Generotti et al., 2015; Bordini et al., 2017; Vanara et al., 2018; Scarpino et al., 2020). Although the approach of each study was different, the cited studies reported a similar trend of the FB distribution in the various maize-milled fractions, in particular with respect to those that considered the 2 different dry-milling processes (DD and TD) at the same time and separately (Vanara et al., 2018; Scarpino et al., 2020).

 The fumonisin A-series are N-acetyl analogs of FBs and, in 1998, Van der Westhuizen et al. reported that these series of fumonisins, have the ability to inhibit sphingosine N-acyltransferase, 231 just like FBs. FA<sub>1</sub> on average represented about 60% of FA<sub>TOT</sub>, while FA<sub>2</sub> represented the 232 remaining 40%. Although the  $FA_{TOT}$  concentration was about 30 times lower than that of  $FB_{TOT}$ , its 233 distribution was almost the same as that of  $FB_{TOT}$ . The cleaning phase on average led to a reduction 234 of -54%, compared to the pre-cleaned whole grain. The  $FA_{TOT}$  content in the animal feed flours from the DD and TD processes increased by 2.4 and 1.9 times, respectively, compared to the pre-236 cleaned whole grain, while the  $FA_{TOT}$  content in the germ from the DD (-53%) and TD (-57%)  processes instead reduced. The maize flour, break meal and pearl meal (DD process) showed an 238 average  $FA_{TOT}$  decrease of  $-61\%$ ,  $-87\%$  and  $-90\%$ , respectively, compared to the pre-cleaned whole grain, while the small and medium hominy grits and the flaking grits (TD process) showed a decrease of -91%, -95% and -97%, respectively.

 The redistribution of the other *Fusarium* mycotoxins and fungal metabolites, produced by species belonging to the *Liseola* section together with the FB and FA, in the different maize dry-milling fractions is reported in Table 2 and Table 3. The cleaning phase always led to a similar reduction of the MON (-45%), BEA (-45%), FUS (-45%), FSA (-53%), FnA (-37%), fusarin C (-60%) and BIK (-27%) contents, in comparison to the pre-cleaned whole grain. The feed flour always showed increases in the MON, BEA, FUS, FSA, FnA, fusarin C and BIK contents of 1.6, 3.4, 2.5, 3.4, 2.3, 247 5.3 and 2.8 times, respectively, compared to the pre-cleaned whole grain. The germ from the DD and TD processes instead presented reduced MON (-41%) and BIK (-54%) contents, an unchanged BEA content, but also increases in the FUS, FSA, FnA and fusarin C contents of 1.6, 3.8, 1.3 and 1.9 times, respectively, compared to the pre-cleaned whole grain. An inverse correlation between the level of contamination of the food grade milling fraction and the particle size was also observed for these other *Fusarium* mycotoxins and metabolites. The maize flour of the DD process was the fraction with the lowest particle size and the smallest reduction, which on average was equal to - 41% for MON, -62% for BEA, -71% for FUS, -69% for FSA, -39% for FnA, -54% for fusarin C and -60% for BIK, compared to the pre-cleaned whole grain. The flaking grits, the fraction with the highest particle size and greatest reduction, on average showed decreases of -87% for MON, -98% for BEA, -94% for FUS, -92% for FSA, -88% for FnA, -60% for fusarin C and -95% for BIK.

258 As far as the toxicological relevance of these other mycotoxins co-produced with  $FB_{TOT}$  by the *Fusarium* spp. of the *Liseola* section is concerned, particular attention should be paid to fusarin C. Although IARC classified it as part of the 2B group in 1993, due to its carcinogenic potential for 261 humans, together with  $FB_1$  and  $FB_2$ , it has not yet been taken into consideration in any legislation. To date, no regulatory limits have also been established concerning the presence of MON. Jonsson

263 et al. (2015) reported a high acute toxicity of MON in rats, with the  $LD_{50}$  value being at the same level as that of T-2 and HT-2 toxins, the most toxic of the *Fusarium* mycotoxins. Moreover, a recent review (Fremy et al., 2019) has underlined an interactive toxicity of MON and FB1. For these reasons, EFSA has recently requested the collection of further data on the presence of MON in food and feeds to allow a comprehensive human risk assessment to be made (EFSA, 2018).

 Toxic effects have also been documented for FSA (Dhani et al., 2017; Mamur et al., 2018), BEA (Ojcius et al., 1991; Logrieco et al., 2002) and for FUS (Logrieco et al., 1996; Ritieni et al., 1997) in humans and animals. FnA is closely related to FSA and is enzymatically derived from it (Fumero et al., 2020), but its toxicity towards humans and animals has not been evaluated extensively. Similarly, there is also a lack of toxicological data for BIK and further support studies are certainly needed (Santos et al., 2020).

 DON was the main regulated mycotoxin among the fungal metabolites produced by *Fusarium* spp. of the Discolor section. However, together with DON, its plant metabolites, that is, DON-3-G, 3- ADON and 15-DON, were always detected in all the maize fractions of both the dry-milling 277 processes. The fate of  $DOM<sub>TOT</sub>$  in the milling fractions is reported in Figure 4. The relative 278 abundance, compared to  $DOM_{TOT}$ , was 56% for DON, 29% for DON-3-G, 14% for 3-ADON and 279 1% for 15-ADON. Interestingly, the DON and the DON-3-G percentages in  $DON<sub>TOT</sub>$  varied as a function of the different milling fractions, as highlighted by the DON-3-G/DON molar ratio (Figure 5). This ratio increased significantly, compared to the pre-cleaned whole grain, in both the DD (+44%) and TD (+33%) germs and, albeit to a lesser extent, in the break meal (+40%) and pearl 283 meal  $(+36%)$  from the DD process and in the small hominy grits  $(+21%)$ , medium hominy grits (+16%) and flaking grits (+30%) from the TD process. On the other hand, the DON-3-G/DON molar ratio decreased in the animal feed flour (-23% and -37% for DD and TD, respectively) and maize flour (-12%). The higher content of this masked mycotoxin, which is not usually monitored, in certain products and by-products, highlights an even greater risk of the consumption of the derived food. This important aspect for consumer health has never been reported before.

289 The cleaning step on average reduced the  $DOM_{TOT}$  content, in comparison to that of the pre-cleaned whole grain content, by -35%. Overall, the germ and the animal feed flour from both processes represented the fractions with the highest DONTOT content. The animal feed flour on average 292 increased  $DOM_{TOT}$  by 2.1 times, in comparison to the pre-cleaned whole grain. On the other hand, contrary to what has been recorded for most metabolites produced by *Fusarium* spp. of the *Liseola* 294 section, the DON<sub>TOT</sub> content always significantly increased in the germ, for both the DD and TD, in comparison to the post-cleaned wholegrain, by 2.8 times. As for the endosperm products, the maize 296 flour, break meal and pearl meal (DD process) showed  $DON<sub>TOT</sub>$  decreases, in comparison to the pre-cleaned whole grain, of -61%, -71% and -78%, respectively, while the small and medium hominy grits and the flaking grits (TD process) showed decreases of -76%, -83% and -93%, respectively, thus confirming an inverse relationship with the particle size.

 ZEA, another regulated mycotoxin produced by *Fusarium* spp. of the *Discolor* section, co-occurred in all the maize fractions with the masked or modified forms ZEA-S, α-ZEA-ol and β-ZEA-ol. ZEA accounted for about 27% of ZEATOT, ZEA-S for 60%, α-ZEA-ol for 5% and β-ZEA-ol for 8%. The 303 redistribution of  $ZEA<sub>TOT</sub>$  in the dry-milling fractions is shown in Figure 6. The cleaning phase on 304 average led to a reduction of  $-60\%$ , compared to the ZEA<sub>TOT</sub> content of the pre-cleaned whole grain. The animal feed flour from both the DD and TD processes on average presented a 2.8 times 306 increase of the  $ZEA_{TOT}$  content, compared to the pre-cleaned whole grain. As for the germ, the ZEA<sub>TOT</sub> content of both DD and TD showed a variable redistribution over the years and on average increased 1.6 times, compared to the pre-cleaned whole grain, and 5.2 times, compared to the post- cleaned whole grain. The endosperm fractions for human consumption, that is, the maize flour, break meal and pearl meal (DD process), showed ZEATOT decreases, compared to the pre-cleaned whole grain, of 48%, 81% and 85%, respectively, while the small and medium hominy grits and the flaking grits (TD process) showed decreases of 89%, 89% and 94%, respectively.

 The distribution of DON and ZEA in the maize dry-milled fractions has only been reported in a few studies (Schaafsma et al., 2004; Brera et al., 2006; Schollenberger et al., 2008; Burger et al., 2013),  and some of these only considered fractions purchased in local markets (Yang et al., 2019) and which were not derived from the same milling process. Moreover, most of the scientific literature has focused on wheat milling and its derived fractions (Scudamore et al., 2009; Kostelanska et al., 2011; Schwake-Anduschus et al., 2015; Edwards et al., 2018; Khaneghah et al., 2018; Guo et al., 2020).

 Like us, Brera et al. (2006) reported that the ZEA level was higher in bran and high fat fractions, such as germs. The present data are also in accordance with the redistribution described by Schaafsma et al. (2004), Schollenberger et al. (2008) and Burger et al. (2013). As for DON, the effects of the process may vary according to the degree of fungal penetration of the endosperm: if the fungal penetration is limited, a notable reduction in the DON level in maize fractions intended for human consumption can be achieved (Brera et al 2006; Khaneghah et al., 2018).

 The present data have pointed out the presence, together with DON and ZEA, of their associated metabolites (masked or modified). DON-3-G and ZEA-S are phase II plant metabolites of the *Fusarium* mycotoxins DON and ZEA, respectively (Berthiller et al., 2013). These associated forms could be hydrolysed in the digestive tract of mammals, thereby contributing to the total dietary exposure of individuals to DON (Berthiller et al., 2011). On the other hand, the acetylated derivatives of DON, that is, 3-ADON and 15-ADON, are usually considered as derived metabolites of phase I (Pinton et al., 2012). Moreover, 3-ADON has been found to be less toxic than DON, while 15-ADON presents a higher toxicity than its precursor DON, while α-ZEAol and β-ZEAol are phase I plant metabolites of ZEA, with a higher toxicity level and greater hyperestrogenic effects, especially for α-ZEAol (Berthiller et al., 2013). Thus, all these modified forms should be considered as additional contributing factors of the total dietary exposure to DON and ZEA and should also be taken into account for correct risk assessments and food safety (JECFA, 2010; CAC, 2011; Lorenz et al., 2019).

 The fate of the CULMTOT, fungal metabolites produced by *Fusarium* spp. of the *Discolor* section, is 340 shown in Figure 7. CULM accounted for about the 38% of CULM<sub>TOT</sub>, 5-OH-CULM for the 30%,  15-OH-CULM for the 24% and 15-OH-culmoron for the 9%. The cleaning phase led to an average 342 reduction of -34%, compared to the CULM<sub>TOT</sub> content of the pre-cleaned whole grain. The animal feed flour from both the DD and TD processes on average increased 2.5 times, compared to the 344 pre-cleaned whole grain. Like the  $DOM_{TOT}$  redistribution, the CULM<sub>TOT</sub> content always significantly increased in the germ, for both the DD and TD processes, compared to the content in the post-cleaned wholegrain, that is, on average by 3.2 times. When considering the maize fractions destined for human consumption with the smallest and largest particle sizes, the maize flour and the 348 flaking grits on average showed CULM<sub>TOT</sub> decreases, compared to the pre-cleaned whole grain, of -64% and -90%, respectively.

 The fate of other fungal metabolites produced by *Fusarium* spp. of the *Discolor* and *Roseum*  sections, including AUR, BUT and EQU, is summarised in Table 4. The cleaning phase generally led to a notable reduction of the AUR content (-60%), compared to the pre-cleaned whole grain, but a slight increase was recorded for BUT and EQU of +2% and +7%, respectively. Overall, the animal feed flour from both the DD and TD processes always showed increases of the AUR, BUT and EQU contents of 2.2, 2.5 and 4.1 times, respectively, compared to the pre-cleaned whole grain. On the other hand, the germ only presented a reduction of the AUR (-46%) and EQU (-54%) contents, but a 1.4 times increase in the BUT content, compared to the pre-cleaned whole grain. Like the other fungal metabolites, among the endosperm fraction intended for human consumption, maize flour on average showed a decrease for the AUR (-85) and BUT (-20%) contents, while the EQU content increased (+37%), compared to the pre-cleaned whole grain. The flaking grits always showed a reduction of the AUR (-99%), BUT (-90%) and EQU (-96%) contents.

 Although CULM was previously reported to have a limited toxic potential in mammals (Dowd et al., 1989; Miller & MacKenzie, 2000), Woelfingseder et al. (2019) have recently reported that CULM could partially inhibit the glucuronidation activity of human liver microsomes. The study carried out by Woelfingseder et al. (2019) underlined the necessity of further studies on the relevance of CULM as a potentially co-occurring modulator of DON toxicokinetics in vivo, and it

 led to the discussion about the possibility of classifying CULM not only as a secondary fungal metabolite but also as an "emerging mycotoxin".

 AUR is a golden yellow *F. graminearum* polyketide bioactive pigment produced under plant stress conditions (Medentsev et al., 2005). It is considered a neglected mycotoxin (Streit et al., 2013; Jarolim et al., 2018), since it is known to induce oxidative stress, cytotoxicity and genotoxicity in human colon cells (Jarolim et al., 2018) and also shows toxicity for differentiated intestinal porcine epithelial cells (IPEC-J2) when combined with DON (Springler et al., 2016). BUT possesses the potential to induce myocardial toxicity (Liu et al., 2007), while EQU has recently been reported to be toxic for chicks (Tayo et al., 2017).

 Among all the previous described emerging *Fusarium* mycotoxins and fungal metabolites, only the fate of MON has been considered in the scientific literature, through the dry-milling of maize (Scarpino et al., 2020), while the other ones have never been reported before in maize dry-milled fractions. Moreover, to the best of the authors' knowledge, this is the first time that the presence and distribution of DON and ZEA have been reported in dry-milled fractions together with their main masked or modified metabolites. Schollenberger et al. (2008) only reported 3-ADON and 15- ADON for DON, and α-ZEAol and β-ZEAol for ZEA, but did not consider DON-3-G or ZEA-S, which are the most commonly and abundantly modified forms of DON and ZEA in food and feeds. Considering the mycotoxins produced from fungal species that do not belong to the *Fusarium* 385 genus, the highest  $AF_{TOT}$  contamination levels were present in the milling fractions during the year 2012 (Figure 8), followed by the year 2014, while the levels were between the limit of detection (LOD) and the limit of quantification (LOQ) for 2013. AFB1 was the form that was present the 388 most, and on average represented about the 70% of the AFTOT content. The fraction with the highest contamination level was the germ of the DD process, in both 2012 and 2014, with a significant increase of 13.3 times in 2012 and a lower increase, that is, of 2.3 times, in 2014, compared to the pre-cleaned whole grain. Moreover, the germ from the TD process presented a significantly lower AFTOT contamination in 2012 than the DD germ. The maize dry-milling products with a

 significantly lower content in 2012 were the pearl meal of the DD process and the small hominy 394 grits of the TD process, with an average  $AF_{TOT}$  content reduction of 60% for both fractions, compared to the pre-cleaned whole grain. On the other hand, no significant differences were recorded for any of the fractions in any of the lots for 2013 and 2014. However, it is important to highlight that since  $AF_{TOT}$  was present at low contamination levels and since fungal growth often occurs in localised hot spots, the mycotoxin distribution in contaminated lots tends to be very heterogeneous and the sampling has even more effect on these mycotoxins (Streit et al., 2012).

 The redistribution of aflatoxins in dry-milled maize fractions was previously considered by Brera et al. (2006), Castells et al. (2008) and Pietri et al. (2009). According to these studies, aflatoxin contamination was uniformly distributed and was more superficial and concentrated in the germ than fumonisin contamination, which conversely affected the inner layers of the kernels and was mainly concentrated in the finer size fractions. However, to the best of the authors' knowledge, among the regulated mycotoxins, the AFs, as well as DON and ZEA distribution in maize-milled fractions, have never been treated before at the same time and separately on the same maize lots through the comparison of 2 different dry-milling processes (DD and TD).

 Some metabolites, such as ENNs, T-2 and HT-2 Toxin, NIV, DAS and *Alternaria* metabolites were present at detectable levels in only a few samples of the pre-cleaned grain. For this reason, their distribution was not evaluated.

 Table 5 summarises the decontamination of the different detected mycotoxins and fungal metabolites in the endosperm fractions (the sum of the maize flour, break meal and pearl meal from DD and the sum of small and medium hominy grits and flaking grits from TD) obtained from different milling processes. Overall, the endosperm fractions from the TD process resulted in less contamination than DD. Thus, considering the inverse relationship with the particle size, flaking grits represented the healthiest maize product for all the metabolites, while the abatement was 417 always lower for maize flour. Taking  $FB_{TOT}$  and  $DOM_{TOT}$  as references, FnA, fusarin C, MON, 418 BUT and AFTOT resulted in an overall higher contamination of the endosperm fractions.

 Nevertheless, it should be considered that very variable behavior was recorded for the fusarin C and 420 AF<sub>TOT</sub> (data not shown), due to their low levels of contamination. On the other hand, FA<sub>TOT</sub>, BIK, BEA, FUS, ZEATOT and AUR showed a higher decontamination in both processes, while FSA, 422 CULM and EQU resulted in a similar behaviour to  $FB_{TOT}$  and  $DOM_{TOT}$ .

 The greater permanence of some mycotoxins and fungal metabolites in the maize food-grade products from the TD and DD dry-milling processes points out a higher risk of exposure for the end consumers. The different fate of the contaminants observed in the present work could allow regulation limits to be defined considering the health impact of the aforementioned mycotoxins.

### *4. Conclusions*

 This is the first time that the redistribution and co-occurrence of a broad spectrum of mycotoxins and fungal metabolites have been considered and reported in an industrial dry-milling study, through the application of different degermination processes.

 The obtained data confirm that a cleaning process is essential to reduce the risk of contamination of almost all the mycotoxins and fungal metabolites. Moreover, the endosperm fractions from the TD process generally showed a lower contamination than DD, for all the metabolites, and an inverse relationship with particle size was always detected.

436 However, the weaker decontamination of some mycotoxins and fungal metabolites  $(AF<sub>TOT</sub>, DON-3-$  G, FnA, fusarin C, MON and BUT) in the food-grade milling fractions points to a higher risk of exposure for the end consumers, particularly when environmental conditions favour their simultaneous increase in whole grain at harvesting. It is also of great importance to point out the 440 concentrations of some mycotoxins and fungal metabolites that were found in the germ (AFTOT and 441 kojic acid, DONTOT, CULMTOT, BUT, FUS, FSA, FnA and in some cases ZEATOT and fusarin C), as well as the significant increase in the content of almost all the mycotoxins and fungal metabolites in the animal feed flour, with a consequent negative impact on animal health.



# *Acknowledgements*

 The authors would like to thank Alessandro Peila, Ugo Peila (Molino Peila Spa, Valperga, To, Italy), Carlo Ferrero and Andrea Pilati (CAPAC Consorzio Agricolo Piemontese per Agroforniture e Cereali Soc. Coop. Agr., Torino, Italy) for their precious help and cooperation in the laboratory and field work. The research has been conducted thanks to the financial support of the Regione Piemonte (Rural

 Development Programme F.E.A.R.S. 2007/2013), as a part of the ALIMAIS and WHITEGRITS projects.

### *References*

- Aprodu, I., & Banu, I. (2015). Co-occurrence of fumonisins and T-2 toxins in milling fractions under industrial conditions. *CyTA - Journal of Food*, *13 (1)*, 102-106. https://doi.org/10.1080/194763372014917702.
- Berthiller, F., Sulyok, M., Krska, R., & Schuhmacher, R. (2007). Chromatographic methods for the simultaneous determination of mycotoxins and their conjugates in cereals. International Journal of Food Microbiology. *119*, 33-37. https://doi.org/10.1016/j.ijfoodmicro.2007.07.022.
- Berthiller, F., Krska, R., Domig, K. J., Kneifel, W., Juge, N., Schuhmacher, R., & Adam, G. (2011). Hydrolytic fate of deoxynivalenol-3-glucoside during digestion. *Toxicology Letters*, *206(3)*, 264-267. https://doi.org/10.1016/j.toxlet.2011.08.006.
- Berthiller, F., Crews, C., Dall'Asta, C., De Saeger, S., Haesaert, G., Karlovsky, P., Oswald, I.P., Walburga, S., Gerrit, S., & Stroka, J. (2013). Masked mycotoxins: a review. *Molecular Nutrition and Food Research*, *57(1)*, 165-186. https://doi.org/10.1002/mnfr.201100764.
- Binder, E. M. (2007). Managing the risk of mycotoxins in modern feed production. *Animal Feed Science and Technology*. *133*, 149-166. https://doi.org/10.1016/j.anifeedsci.2006.08.008.
- Blandino, M., Alfieri, M., Giordano, D., Vanara, F., & Redaelli, R. (2017a). Distribution of bioactive compounds in maize fractions obtained in two different types of large scale milling processes. *Journal of Cereal Science*, *77*, 251-258. https://doi.org/10.1016/j.jcs.2017.08.006.
- Blandino, M., Scarpino, V., Giordano, D., Sulyok, M., Krska, R., Vanara, F., & Reyneri, A. (2017b). Impact of sowing time, hybrid and environmental conditions on the contamination of maize by emerging mycotoxins and fungal metabolites. *Italian Journal of Agronomy*, *12(928)*, 215-224. https://doi.org/10.4081/ija.2017.928.
- Bordini, J. G., Ono, M. A., Garcia, G. T., Fazani, V. H. M., Vizoni, E., Rodrigues, K. C. B., Hirooka, E. Y., & Ono, E. Y. H. (2017). Impact of industrial dry-milling on fumonisin
- redistribution in non-transgenic corn in Brazil. *Food Chemistry*, *220*, 438-443. https://doi.org/10.1016/j.foodchem.2016.10.028.
- Brera, C., Debegnach, F., Grossi, S., & Miraglia, M. (2004). Effect of the industrial processing on the distribution of fumonisin B1 in dry milling corn fractions. *Journal of Food Protection*, *67*, 1261-1266. https://doi.org/10.4315/0362-028X-67.6.1261.
- Brera, C., Catano, C., De Santis, B., Debegnach, F., De Giacomo, M., Pannunzi, E., & Miraglia, M. (2006). Effect of Industrial Processing on the Distribution of Aflatoxins and Zearalenone in Corn-Milling Fractions. *Journal of Agricultural and Food Chemistry*, *54*, 5014-5019. https://doi.org/10.1021/jf060370s.
- Broggi, L. E., Resnik, S. L., Pacin, A. M., Gonzalez, H. H. L., Cano, G., & Taglieri, D. (2002). Distribution of fumonisins in dry-milled corn fractions in Argentina. Food Additives & Contaminants, *19(5)*, 465-469. https://doi.org/10.1080/02652030110103484.
- Bullerman, L. B., & Bianchini, A. (2007). Stability of mycotoxins during food processing. *International Journal of Food Microbiology*, *119*, 140-146. https://doi.org/10.1016/j.ijfoodmicro.2007.07.035.
- Burger, H-m., Shephard, G. S., Louw, W., Rheeder, J. P., & Gelderblom, W. C. A. (2013). The mycotoxin distribution in maize milling fractions under experimental conditions. *International Journal of Food Microbiology*, *165*, 57-64. http://dx.doi.org/10.1016/j.ijfoodmicro.2013.03.028.
- Castells, M., Marin, S., Sanchis, V., & Ramos, A. J. (2008). Distribution of fumonisins and aflatoxins in corn fractions during industrial cornflake processing. *International Journal of Food Microbiology*, *123*, 81-87. https://doi.org/10.1016/j.ijfoodmicro.2007.12.001.
- Codex Alimentarius Commission. (2011). *Report of the Fifth Session of the Codex Committee on*
- *Contaminants. In Foods, The Hague, The Netherlands.* http://www.codexalimentarius.net/download/report/758/REP11 CFe.pdf.

- Dall'Erta, A., Cirlini, M., Dall'Asta, M., Del Rio, D., Galaverna, G., & Dall'Asta C. (2013). Masked Mycotoxins Are Efficiently Hydrolyzed by Human Colonic Microbiota Releasing Their Aglycones. *Chemical Research in Toxicolology*, *26*, 305-312. https://doi.org/10.1021/tx300438c.
- Dhani, S., Nagiah, S., Naidoo, D. B., & Chuturgoon, A. A. (2017). Fusaric acid immunotoxicity and MAPK activation in normal peripheral blood mononuclear cells andThp-1 cells. *Scientific Reports*, *7(1)*, 3051. https://doi.org/10.1038/s41598-017-03183-0.
- Dowd, P.F., Miller, J., & Greenhalgh, R. (1989). Toxicity and Interactions of some *Fusarium graminearum* metabolites to caterpillars. *Mycologia*, 81, 646-650. https://doi.org/10.2307/3760143.
- Edwards, S. G., Kharbikar, L. L., Dickin, E. T., MacDonald, S., & Scudamore, K. A. (2018). Impact of pre-harvest rainfall on the distribution of fusarium mycotoxins in wheat mill fractions. *Food Control*, *89*, 150-156. https://doi.org/10.1016/j.foodcont.2018.02.009.
- EFSA CONTAM Panel (EFSA Panel on Contaminants in the Food Chain), Knutsen, H. K.,
- Alexander, J., Barregård, L., Bignami, M., Brüuschweiler, B., Ceccatelli, S., Cottrill, B.,
- Dinovi, M., Grasl-Kraupp, B., Hogstrand, C., Hoogenboom, L. R., Nebbia, C. S., Oswald, I.
- P., Petersen, A., Rose, M., Roudot, A.-C., Schwerdtle, T., Vleminckx, C., Vollmer, G.,
- Wallace, H., De Saeger, S., Eriksen, G. S., Farmer, P., Fremy, J.-M., Gong, Y. Y., Meyer,
- K., Naegeli, H., Parent-Massin, D., van Egmond, H., Altieri, A., Colombo, P., Eskola, M.,
- van Manen, M., & Edler, L. (2018). Scientific Opinion on the risks to human and animal
- health related to the presence of moniliformin in food and feed. *EFSA Journal 2018*, *16(3)*, 5082, 95 pp. https://doi.org/10.2903/j.efsa.2018.5082.
- Eskola, M., Kos, G., Elliott, C. T., Hajšlová, J., Mayar, S., & Krska, R. (2019) Worldwide contamination of food-crops with mycotoxins: Validity of the widely cited 'FAO estimate' of 25%, *Critical Reviews in Food Science and Nutrition*. https://doi.org/10.1080/10408398.2019.1658570.
- European Commision. (2006). Commission Regulation (EC) No. 401/2006, of 23 February 2006 laying down the methods of sampling and analysis for the official control of the levels of mycotoxins in foodstuffs. *Off. J. Eur. Union., 70*, 12-34.
- Fremy, J.-M., Alassane-Kpembi, I., Oswald, I. P., Cottrill, B., & Van Egmond, H. P. (2019). A review on combined effects of moniliformin and co-occurring *Fusarium* toxins in farm
- animals. *World Mycotoxin Journal*, *12 (3),* 281-291. https://doi.org/10.3920/WMJ2018.2405.
- Fumero M. V., Sulyok M., Ramirez M. L., Leslie J. F. & Chulze S. N. (2020). Effects of water activity and temperature on fusaric and fusarinolic acid production by *Fusarium temperatum*. *Food Control*, *114*, 1072632. https://doi.org/10.1016/j.foodcont.2020.107263.
- Generotti, S., Cirlini, M., Dall'Asta, C., & Suman, M. (2015). Influence of the industrial process from caryopsis to cornmeal semolina on levels of fumonisins and their masked forms. *Food Control*, *48*, 170-174. https://doi.org/10.1016/j.foodcont.2014.06.003.
- Guo, H., Ji, J., Wang, J.-S., & Sun, X. (2020). Deoxynivalenol: Masked forms, fate during food processing, and potential biological remedies. *Comprehensive Reviews In Food Science And Food Safety*, *19*, 895-926. https://doi.org/10.1111/1541-4337.12545.
- Gwirtz, J. A., & Garcia-Casal, M. N. (2014). Processing maize flour and corn meal food products. *Annals of the New York Academy of Sciences*, *1312(1)*, 66-75. https://doi.org/10.1111/nyas.12299.
- International Agency for Research on Cancer (IARC). (1993). Some naturally occurring substances: food items and constituents, heterocyclic aromatic amines and mycotoxins. In IARC Monographs on the Evaluation of Carcinogenic Risks to Humans, (Vol. no. 56, pp. 1-599).
- Jarolim, K., Wolters, K., Woelflingseder, L., Pahlke, G., Beisl, J., Puntscher, H., Braun, D., Sulyok,
- M., Warth, B., & Marko, D. (2018). The secondary *fusarium* metabolite aurofusarin induces oxidative stress, cytotoxicity and genotoxicity in human colon cells. *Toxicology Letters*,
- 
- *284*, 170-183. https://doi.org/10.1016/j.toxlet.2017.12.008.
- JECFA. (2010). *Joint Food and Agriculture Organization/World health Organization Expert Committee on food Additives. In Joint FAO/WHO Expert Committee on food Additives Seventy-second Meeting, Rome, 16-25 February 2010: Summary and Conclusions*. http://www.who.int/foodsafety/chem/summary72\_rev.pdf.
- Jestoi, M. (2008). Emerging fusarium-mycotoxins fusaproliferin, beauvericin, enniatins, and moniliformin: a review. *Critical Review in Food Science and Nutrition*. *48(1)*, 21-49. https://doi.org/10.1080/10408390601062021.
- Jonsson, M., Atosuo, J., Jestoi, M., Nathanail, A. V., Kokkonen, U.-M., Anttila, M., Koivisto, P., Lilius, E.-M., & Peltonen, K. (2015). Repeated dose 28-day oral toxicity study of moniliformin in rats. *Toxicology Letters*, *233*, 38-44. https://doi.org/10.1016/j.toxlet.2014.11.006.
- Katta, S. K., Cagampang, A. E., Jackson, L. S., & Bullerman, L. B. (1997). Distribution of *Fusarium* molds and fumonisins in dry-milled corn fractions. *Cereal Chemistry*, *74(6)*, 858- 863. https://doi.org/10.1094/CCHEM.1997.74.6.858.
- Khaneghah, A. M., Martins, L. M., von Hertwig, A. M., Bertoldo, R., & Sant'Ana, A. S. (2018). Deoxynivalenol and its masked forms: Characteristics, incidence, control and fate during wheat and wheat based products processing-A review. *Trends in Food Science & Technology*, *71*, 13-24. https://doi.org/10.1016/j.tifs.2017.10.012.
- Kostelanska, M., Dzuman, Z., Malachova, A., Capouchova, I., Prokinova, E., Skerikova, A., & Hajslova, J. (2011). Effects of milling and baking technologies on levels of deoxynivalenol and its masked form deoxynivalenol-3-glucoside. *Journal of Agricultural and Food Chemistry*, *59(17)*, 9303-9312. https://doi.org/10.1021/jf202428f.
- Liu, J.-B., Wang, Y.-M., Peng, S.-Q., Han, G., Dong, Y.-S., Yang, H.-Y., Yan, C.-H., & Wang, G.-
- Q. (2007). Toxic effects of *Fusarium* mycotoxin butenolide on rat myocardium and primary culture of cardiac myocytes. *Toxicon*, *50*, 357-364. https://doi.org/10.1016/j.toxicon.2007.04.014.



- Logrieco, A., Mulè, G., Moretti, A., & Bottalico, A. (2002). Toxigenic *Fusarium* Species and Mycotoxins Associated with Maize Ear Rot in Europe. *European Journal of Plant Pathology*, *108*, 597-609. https://doi.org/10.1023/A:1020679029993.
- Lorenz, N., Dänicke, S., Edler, L., Gottschalk, C., Lassek, E., Marko, D., Rychlik, M., & Mally, A.
- (2019). A critical evaluation of health risk assessment of modified mycotoxins with a special focus on zearalenone. *Mycotoxin Research*, *35*, 27-46. https://doi.org/10.1007/s12550-018- 0328-z.
- Malachova, A., Sulyok, M., Beltran, E., Berthiller, F., & Krska, R. (2014). Optimization and validation of a quantitative liquid chromatography - tandem mass spectrometric method covering 295 bacterial and fungal metabolites including all relevant mycotoxins in four model food matrices. *Journal of Chromatography A*, *1362*, 145-156. https://doi.org/10.1016/j.chroma.2014.08.037.
- Mamur, S., Unal, F., Yilmaz, S., Erikel, E., & Yuzbasioglu, D. (2018). Evaluation of the cytotoxic and genotoxic effects of mycotoxin fusaric acid. *Drug and ChemicalToxicology*, *43(2)*, 159- 157. https://doi.org/10.1080/01480545.2018.1499772.
- Marin, S., Ramos, A. J., Cano-Sancho, G., & Sanchis, V. (2012). Reduction of mycotoxins and toxigenic fungi in Mediterranean basin maize chain. *Phytopathologia Mediterranea*, *51(1)*, 93-118. https://doi.org/10.14601/Phytopathol\_Mediterr-9411.
- Medentsev, A. G., Arinbasarova, A., & Akimenko, V. K. (2005). Biosynthesis of naphthoquinone pigments by fungi of the genus *fusarium*. *Applied Biochemistry and Microbiology*, *41*, 503-
- 507. https://doi.org/10.1007/s10438-005-0091-8.
- Miller, J., & MacKenzie, S. (2000). Secondary metabolites of *Fusarium venenatum* strains with deletions in the Tri5 gene encoding trichodiene synthetase. *Mycologia*, *92*, 764-771. https://doi.org/10.2307/3761433.
- Ojcius, D. M., Zychlinsky, A., Zheng, L. M., & Young, J. D.-E. (1991). Ionophore-induced apoptosis: role of DNA fragmentation and calcium fluxes. *Experimental Cell Research*, *197*, 43-49. https://doi.org/10.1016/0014-4827(91)90477-C.
- Pietri, A., Zanetti, M., & Bertuzzi, T. (2009). Distribution of aflatoxins and fumonisins in dry milled maize fractions. *Food Additives & Contaminants - Part A Chemistry, Analysis, Control, Exposure and Risk Assessment*, *26(3)*, 372-380. https://doi.org/10.1080/02652030802441513.
- Pinton, P., Tsybulskyy, D., Lucioli, J., Callu, P., Lyazhri, F., Grosjean, F., Bracarense, A. P., Kolf- Clauwand, M., & Oswald, I. P. (2012). Toxicity of deoxynivalenol and its acetylated derivatives on the intestine: differential effects on morphology, barrier function, tight junctions' proteins and mitogen-activated protein kinases. *Toxicological Sciences*, *130*, 180- 190. https://doi.org/10.1093/toxsci/kfs239.
- Rai, S., Kaur, A., & Chopra, C. S. (2018). Gluten-Free Products for Celiac Susceptible People. *Frontiers in Nutrition*, *5*, 116. https://doi.org/10.3389/fnut.2018.00116.
- Ritieni, A., Monti, S. M., Randazzo, G., Logrieco, A., Moretti, A., Peluso, G., Ferracane, R., & Fogliano, V. (1997). Teratogenic effects of fusaproliferin on chicken embryos. *Journal of Agricultural and Food Chemistry*, *45*, 3039-3043. https://doi.org/10.1021/jf960890v.
- Rychlik, M., Humpf, H., Marko, D., Dänicke, S., Mally, A., Berthiller, F., Klaffke, H., & Lorenz, N. (2014). Proposal of a comprehensive definition of modified and other forms of mycotoxins including "masked" mycotoxins. *Mycotoxin Research*, *30*, 197-205. https://doi.org/10.1007/s12550-014-0203-5.
- Santos, M. C., de Lima Mendonça, M., & Bicas, J. L. (2020). Modeling bikaverin production by *Fusarium oxysporum* CCT7620 in shake flask cultures. *Bioresources and Bioprocessing*, *7*, 13. https://doi.org/10.1186/s40643-020-0301-5.
- Scarpino, V., Vanara, F., Reyneri, A., & Blandino, M. (2020). Fate of moniliformin during different large-scale maize dry-milling processes. *LWT - Food Science and Technology*, *123*, 1-7. https://doi.org/10.1016/j.lwt.2020.109098.
- Schaafsma, A. W., Frégeau-Reid, J., & Phibbs, T. (2004). Distribution of deoxynivalenol in *Gibberella*-infected food-grade maize kernels. *Canadian Journal Of Plant Science*, *84*, 909- 913. https://doi.org/10.4141/p03-049.
- Schollenberger, M., Müller, H.-M., Rüfle, M., Suchy, S., & Drochner., W. (2008). Redistribution of 16 Fusarium Toxins During Commercial Dry Milling of Maize. *Cereal Chemistry*, *85(4)*, 557-560. https://doi.org/10.1094/CCHEM-85-4-0557.
- Schwake-Anduschus, C., Proske, M., Sciurba, E., Muenzing, K., Koch, M., & Maul, R. (2015). Distribution of deoxynivalenol, zearalenone, and their respective modified analogues in milling fractions of naturally contaminated wheat grains. *World Mycotoxin Journal*, *8(4)*,
- 433-443. https://doi.org/10.3920/WMJ2014.1818.
- Scudamore, K. A., & Patel, S. (2000). Survey for aflatoxins, ochratoxin A, zearalenone and fumonisins in maize imported into the United Kingdom, *Food Additives & Contaminants*, *17(5)*, 407-416. https://doi.org/10.1080/026520300404824.
- Scudamore, K., Hazel, C. M., Patel, S., & Scriven, F. (2009). Deoxynivalenol and other *Fusarium* mycotoxins in bread, cake, and biscuits produced from UK-grown wheat under commercial and pilot scale conditions. *Food Additives & Contaminants*, *26(8)*, 1191–1198. https://doi.org/10.1080/02652030902919426.
- Springler, A., Vrubel, G. J., Mayer, E., Schatzmayr, G., & Novak, B. (2016). Effect of *fusarium*- derived metabolites on the barrier integrity of differentiated intestinal porcine epithelial cells (IPEC-J2). *Toxins*, *8*, 345. https://doi.org/10.3390/toxins8110345.



- Streit, E., Schwab, C., Sulyok, M., Naehrer, K., Krska, R., & Schatzmayr, G. (2013). Multi- mycotoxin screening reveals the occurrence of 139 different secondary metabolites in feed and feed ingredients. *Toxins*, *5(3)*, 504-523. https://doi.org/10.3390/toxins5030504.
- Sulyok, M., Berthiller, F., Krska, R., & Schuhmacher, R. (2006). Development and validation of a
- liquid chromatography/tandem mass spectrometric method for the determination of 39 mycotoxins in wheat and maize. *Rapid Communications in Mass Spectrometry*, *20*, 2649- 2659. https://doi.org/10.1002/rcm.2640.
- Tayo, G. O., Ajayi, B., Olarinmoye, A. O, Ezekiel, C., Taiwo, E. A., Babalola, O. O., Nwangburuka, C. C., Denton, L., Chioma, G. O., & Oyekale, K. O. (2017). Effect of levels of equisetin and fumonisin mycotoxins on blood parameters of broiler chicks. *Mycopath*, *15(2)*, 55-59.
- 681 Vanara, F., Reyneri, A., & Blandino, M. (2009). Fate of fumonisin  $B_1$  in the processing of whole maize kernels during dry-milling. *Food Control*, *20*, 235-238. https://doi.org/10.1016/j.foodcont.2008.05.014.
- Vanara, F., Scarpino, V., & Blandino, M. (2018). Fumonisin Distribution in Maize Dry-Milling Products and By-Products: Impact of Two Industrial Degermination Systems. *Toxins*, *10(9)*, 357. https://doi.org/10.3390/toxins10090357.
- van der Westhuizen, L., Shephard, G. S., Snyman, S. D., Abel, S., Swanevelder, S., & Gelderblom,
- W. C. A. (1998). Inhibition of sphingolipid biosynthesis in rat primary hepatocyte cultures
- by fumonisin B1 and other structurally related compounds. *Food and Chemical Toxicology*,
- *36*, 497-503. https://doi.org/10.1016/s0278-6915(98)00012-x.



# **Figure Captions**

**Figure 1.** Distribution of the particle sizes of the maize products after the milling process.

- 719 **Figure 2.** Total fumonisin B (FB<sub>TOT</sub>) distribution in the fractions of different dry milling processes and different maize lots.
- 721 **Figure 3.** Total fumonisin A (FA<sub>TOT</sub>) distribution in the fractions of different dry milling processes and different maize lots.
- 723 **Figure 4.** Total deoxynivalenol (DON<sub>TOT</sub>) distribution in the fractions of different dry milling processes and different maize lots.
- **Figure 5.** Averaged DON-3-G/DON molar ratio distribution in the fractions of different dry milling processes.
- 727 **Figure 6.** Total zearalenone (ZEA<sub>TOT</sub>) distribution in the fractions of different dry milling processes and different maize lots.
- 729 **Figure 7.** Total culmorin (CULM<sub>TOT</sub>) distribution in the fractions of different dry milling processes and different maize lots.
- 731 **Figure 8.** Total aflatoxin (AF<sub>TOT</sub>) distribution in the fractions of different dry milling processes and maize lots.





736 **Figure 2.**





738 DD = dry-degermination; TD = tempering-degermination; WG = whole grain; HG = hominy grits.<br>739 Different letters above the bars indicate significant differences in the fractions ( $p < 0.05$ ). Different letters above the bars indicate significant differences in the fractions  $(p < 0.05)$ .

740 **Figure 3.**







742 DD = dry-degermination; TD = tempering-degermination; WG = whole grain; HG = hominy grits.<br>743 Different letters above the bars indicate significant differences in the fractions ( $p < 0.05$ ). Different letters above the bars indicate significant differences in the fractions  $(p < 0.05)$ .





746 DD = dry-degermination; TD = tempering-degermination; WG = whole grain; HG = hominy grits. 747 Different letters above the bars indicate significant differences in the fractions  $(p < 0.05)$ .

#### **DON-3-G/DON Molar Ratio**





752

749

- 753
- 754

755

756

757

- 758
- 759
- 760
- 761

762

763

764

765





 $769$  DD = dry-degermination; TD = tempering-degermination; WG = whole grain; HG = hominy grits.<br>770 Different letters above the bars indicate significant differences in the fractions ( $p < 0.05$ ). Different letters above the bars indicate significant differences in the fractions ( $p < 0.05$ ).



**CULM 5-OH-CULM 15-OH-CULM 15-OH-culmoron**









 $DD = dry-degermination$ ;  $TD = temperature-degermination$ ;  $WG = whole grain$ ;  $HG = hominy$  grits. 778 Different letters above the bars indicate significant differences in the fractions  $(p < 0.05)$ .

# 779 **Tables**

780 **Table 1.** Maize lots processed in the industrial mill, ranked according to the maize production year 781 and the average values of the main regulated, masked, emerging mycotoxins and other secondary

782 fungal metabolites in pre-cleaned whole grain expressed as  $\mu$ g kg<sup>-1</sup>  $\pm$  standard deviation (SD).



783 The reported contamination means for each lot were based on 2 repetitions.

**784**  $a$   $a$ DO = limit of quantification = 1.6 µg kg<sup>-1</sup> for HFB1; 4.8 µg kg<sup>-1</sup> for fusarin C; 0.1 µg kg<sup>-1</sup> for ENN<sub>TOT</sub>; 0.4 µg kg<sup>-1</sup> for DAS; 1.2 µg 785 kg<sup>-1</sup> for NIV; 3.2 µg kg<sup>-1</sup> for HT-2 toxin; 0.4 µg kg<sup>-1</sup> for AOH; 0.032 µg kg<sup>-1</sup> for AME; 0.08 µg kg<sup>-1</sup> for TEN; 8.0 µg kg<sup>-1</sup> for TeA;

**786** 0.4  $\mu$ g kg<sup>-1</sup> for ALS.

787 **Table 2.** Moniliformin (MON), beauvericin (BEA) and fusaproliferin (FUS) distributions in the fractions of different dry milling processes and 788 different maize lots.



789 <sup>*a*</sup> dry milling process: DD, dry-degermination; TD, tempering-degermination.

790 *<sup>b</sup>*  $\mathbf{W}$ <sup>b</sup>WG = whole grain; HG = hominy grits.

**791** *c*LOQ = limit of quantification = 0.008  $\mu$ g kg<sup>-1</sup> for BEA; 40  $\mu$ g kg<sup>-1</sup> for FUS.

792 Means followed by different letters are significantly different (the significance level is shown in the table).<br>793 The reported mycotoxin contamination values for the fractions of each lot/year are based on 2 repetiti

The reported mycotoxin contamination values for the fractions of each lot/year are based on 2 repetitions.

794

795



797 **Table 3.** Fusaric acid (FSA), fusarinolic acid (FnA), Fusarin C and bikaverin (BIK) distributions in the fractions of different dry milling processes 798 and different maize lots.

799

<sup>*a*</sup> dry milling process: DD, dry-degermination; TD, tempering-degermination.

801 *<sup>b</sup>*  $W\ddot{G}$  = whole grain; H $G$  = hominy grits.

802 *c*LOQ = limit of quantification = 16 µg kg<sup>-1</sup> for FSA; 4.8 µg kg<sup>-1</sup> for fusarin C; 8 µg kg<sup>-1</sup> for BIK.

803 Means followed by different letters are significantly different (the significance level is shown in the table).<br>804 The reported mycotoxin contamination values for the fractions of each lot/year are based on 2 repetiti

The reported mycotoxin contamination values for the fractions of each lot/year are based on 2 repetitions.

805

806

807

808



810 **Table 4.** Aurofusarin (AUR), butenolide (BUT) and equisetin (EQU) distributions in the fractions of different dry milling processes and different maize lots.

812 <sup>d</sup>ry milling process: DD, dry-degermination; TD, tempering-degermination.

813 *<sup>b</sup>*  $b\text{WG}$  = whole grain; HG = hominy grits.

814  $CDOQ =$  limit of quantification = 2.4  $\mu$ g kg<sup>-1</sup> for AUR; 5.6  $\mu$ g kg<sup>-1</sup> for BUT; 0.24  $\mu$ g kg<sup>-1</sup> for EQU.

815 Means followed by different letters are significantly different (the significance level is shown in the table).<br>816 The reported mycotoxin contamination values for the fractions of each lot/year are based on 2 repetiti

The reported mycotoxin contamination values for the fractions of each lot/year are based on 2 repetitions.

## 817 **Table 5.** Contamination percentage of different mycotoxins in endosperm fractions obtained from

818 different milling processes.



820 *<sup>a</sup>* Data were calculated according to the balance mass criteria, considering the specific milling yield and the 821 contamination of each fraction.<br>822  $b$ The occurrence of each myce

<sup>*b*</sup> The occurrence of each mycotoxin is reported for each milling process (DD, dry degermination; TD, tempering degermination) as the percentage with respect to the raw material content (contamination of pre-cleaned who 823 degermination) as the percentage with respect to the raw material content (contamination of pre-cleaned whole grain =  $824$  100).

 $100$ ).