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Effects of housing systems on behaviour and welfare of autochthonous laying hens and a commercial hybrid

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Abstract

The welfare of laying hens is crucial for a sustainable and ethical food industry. Free-range systems offer a more natural and stimulating environment, while dual-purpose autochthonous breeds and crossbreeding could address ethical concerns and improve productivity. In this study, behaviour, plumage status, and leg health of five genotypes (two local breeds, two crossbreeds with a medium growth hybrid, and one commercial hybrid) in two different housing systems (enriched cage/free-range) were evaluated. The animals were filmed with professional cameras for three consecutive days at 30 and 64 weeks of age. Each day the recording session lasted 20 minutes, resulting in a total of 120 minutes per pen (20 min X 6 days recorded). A total of 16 behaviours divided into six categories (feeding, movement, social, resting, comfort, stretching) were scored. In addition, the tonic immobility test was performed, and the plumage status and footpad dermatitis of the birds were assessed. The results revealed a significant influence of the housing system on the behaviour, plumage status, and foot health of laying hens. While the housing system affected all

behaviours, mainly feeding and comfort behaviours were influenced by the genotype of the hens and the interaction between the housing system and genotype. All genotypes displayed a similar behavioural pattern within the two housing systems, except for the commercial hybrid, which differed in its behaviour from the others adapting better to the enriched cage environment. In particular, the free-range system provided hens with more opportunities to engage in movement and comfort behaviour, and social interactions. Furthermore, hens in the free-range system exhibited better plumage conditions and lower rates of footpad dermatitis, indicating a positive impact on their overall welfare.

1. Introduction

Studying chicken behaviour is important for several reasons. First and foremost, chickens are an important source of food for humans, and understanding their behaviour can enhance their welfare and productivity. Studying how these animals interact with each other and with their environment enables the development of more effective animal management strategies, promoting their health and mitigating stress. The welfare of laying hens is a critical factor for a modern, sustainable, and ethical food industry. Proper management practices to promote welfare can prevent the spread of diseases, reduce stress, and improve egg quality traits (Janczak and Riber, 2015). By providing adequate space, feed, water, and environmental enrichment, laying hens can perform natural behaviours and maintain their physical and welfare status. This can contribute to a reduction in the mortality rate and even to an increase in productivity (Rodenburg et al., 2008). Furthermore, guaranteeing the welfare of laying hens is also important from an ethical and human point of view as well as to satisfy the requests from consumers that are increasingly interested in buying products with both high ethical standards and nutritional value (Vale et al., 2019). The European Union's Directive 1999/74/EC,

enforced in 2012, banning conventional cages for egg-laying hens, underscores the imperative for alternative non-cage systems in poultry farming to comply with welfare standards.

Free-range systems for laying hens offer an alternative to intensive cage-systems, providing hens with a more natural and stimulating environment. These systems typically include open grass, shade, and shelter, allowing hens to roam freely, engage in natural behaviours like foraging and dust bathing, develop social interactions, and reduce stress (Campbell et al., 2021). The primary challenges of free-range systems are the greater space requirements and the need for more complex management compared to intensive systems (Campbell et al., 2020).

In egg production industry, the culling of male chicks poses a significant ethical and sustainability issue. The potential use of local breeds as dual-purpose lines could help solve this problem. Indeed, while females from dual purpose breeds can be used for egg production, males can be raised for producing meat, thus providing an alternative to the killing of day-old male chicks (Bruijnis et al., 2015). The local breeds have been developed and adapted over generations to the local climate and production conditions, making them suitable for small-scale farming systems (Stefanetti et al., 2023). Furthermore, the use of indigenous breeds can also help diversify local food systems, reducing the dependence on imported breeds and genetic resources and improving food security (Fiorilla et al., 2023). Crossbreeding chickens involves mating different breeds to create offspring with desirable traits, such as improved egg production, growth rate, and disease resistance. This enhances adaptability to various conditions, making them cost-effective for producers. While hybridization can diversify the gene pool and boost resilience, careful consideration of potential drawbacks, like reduced heterozygosity and loss of genetic resources, is crucial. Responsible and sustainable breeding programs are essential for reaping the benefits without adverse effects (Besbes and Gibson, 1999).

Understanding complex behavioural datasets often requires advanced statistical techniques. The t-Stochastic Neighbor Embedding (t-SNE) method efficiently reduces data dimensionality while preserving its structure, facilitating visualization of intricate patterns. By mapping high-dimensional data to lower dimensions, t-SNE unveils relationships and clusters, especially beneficial for nonlinear data. When coupled with two-way analysis of variance (ANOVA) to detect group differences, t-SNE enhances analysis by visually representing data, aiding in identifying distinct behavioural patterns or clusters, thus enabling a comprehensive understanding of the dataset's structure and relationships.

In this study, the behaviours, plumage conditions, and leg health of five different genotypes of laying hens were assessed. These genotypes included a commercial hybrid, two local Italian dual-purpose breeds, and their crossbreeds with a medium-growing genotype (Sasso, (Hendrix Genetics)). Local chicken breeds have proven to be highly adaptable to low-input and alternative farming systems. Their resilience and efficiency minimize the need for artificial inputs, offering a concrete alternative to reduce the environmental footprint of poultry farming while preserving biodiversity (Fiorilla et al., 2024; Mancinelli et al., 2023). Our primary aim was to evaluate how housing systems, genotype, and animal age influence the health and behaviours of laying hens. Laying hens display diverse behaviours depending on the type of housing system they inhabit. Hens kept in cage systems often exhibit a higher frequency of discomfort indicators (Lay et al., 2011). We hypothesize distinct differences between the two housing systems among genotypes, particularly between the local breeds and the commercial hybrid. Moreover, an interaction effect between genotype and housing system is expected.

2. Materials and Methods

2.1 Animals and housing

The present study was conducted at the poultry facility of the Department of Agricultural and Food Sciences of the University of Bologna (Italy) and was approved by the Ethical Committe (Prot. ID: 1102/2019). Three hundred chicks of five different genotypes, Robusta Maculata (RM, 60 hens), Bionda Piemontese (BP, 60 hens), RM x Sasso (RMxS, 60 hens), BP x Sasso (BPxS, 60 hens) and a commercial hybrid (Lohmann Brown, LB, 60 hens) were used for the present study. The chicks were immunized against Marek and Newcastle diseases and sexing was determined through visual examination of the cloaca at hatching. As dual-purpose breeds, the males from the two local breeds and their crossbreed were involved in a fattening trial conducted at the University of Turin (Prot. ID: 251833), while the females were utilized in this trial, ensuring that no chicks were culled at hatching. At 18 weeks of age, the hens were randomly allocated to two different housing systems: enriched cage and free-range system. Each one of the five genotypes was divided in three replicates, for a total of 15 pens in the enriched cage farming system and 15 in the free-range farming system. The enriched cages had a total of 75 hens, 5 in each pen with 0.144 m² available floorspace each hen while the free-range farming system had a total of 225 hens, 15 in each pen with 1.067 m² available floorspace each hen as shown in table 1 and figure S1. The dimension of the enriched cages was 120 X 60 cm, each cage had a 30 X 30 cm nest and three 60 cm perches. A feeder measuring 120 cm was positioned in front of each cage (supplementary material, Figure S2). Lighting was regulated to provide 16 hours of light and 8 hours of darkness daily, while maintaining an average temperature of 23°C.

In the free-range system, the birds were housed in fenced pens with an insulated nest for egg-laying. The pens measured 4 X 4 meters, and the nests measured 140 X 120 cm (supplementary material, Figure S2). The entire facility, including the outdoor areas, was safeguarded from wild birds and predators using fences. The trial took place from September 2022 to June 2023 in Ozzano dell'Emilia (Bologna, Italy). Throughout the entire experimental period, the birds were subjected to natural temperature, lighting, and humidity

conditions. Details regarding the average temperature, rainfall in millimeters, and hours of daylight recorded in Ozzano dell'Emilia (Bologna, Italy) during the experimental trial can be found in Table S1 in the supplementary material. Water was provided *ad libitum* through a single large drinker that allowed for the simultaneous drinking of four birds.

The feed provided was a standard commercial feed designed for layers and utilized in both the enriched cage and free-range systems. It was provided in four different formulations tailored to the hen's age: Pre-Layer (18-25 weeks), Layer 1 (26-51 weeks), Layer 2 (52-62 weeks) and Layer 3 (63-66 weeks) (Table S2 supplementary material). Throughout the entire experimental trial, the birds' health status and mortality were checked daily.

2.2 Behavioural observations

The animals were recorded using professional cameras (Sony FDRAX43A 4K) for three consecutive days, for a total of 60 minutes per pen at 30 weeks (20 minutes each day) of age and 60 minutes per pen at 64 weeks of age (20 minutes each day). The recordings were performed every day at the same time at 11 a.m. The camera was placed outside the pen allowing to monitor all the animals' behaviours. The recorded videos were analyzed using the Behavioural Observation Research Interactive Software (BORIS, v 7.9.7) (Friard and Gamba, 2016). The behaviours frequencies were evaluated as the number of times that a specific behaviour occurred in the pen during the daily 20 minutes periods of observations. The behaviours were divided according to a typical laying hen behaviour ethogram (Table 2). The ethogram was elaborated starting from existing literature and adapted to the specific rearing conditions of this trial (Biasato et al., 2022; Hurnik and Webster, 1990; Kenny et al., 2017; Liu et al., 2020; McCowan et al., 2006).

2.3 Tonic immobility test

The tonic immobility (TI) test was performed on 3 laying hens each pen at 30 and 64 weeks of age after video recording. The hens were captured from their home pen placed on their backs in a plastic cradle and restrained for 5 seconds by a trained experimenter, who placed one hand on the bird's chest and another over its head with the head hanging down. The experimenter then removed its hand and stepped aside with their eyes averted downwards (Campbell et al., 2019). A maximum of three attempts were made to induce immobility. The animals were not allowed to remain in the immobility condition for more than 3 minutes (Ferrante et al., 2005). The duration of TI was subsequently recorded, with 3 minutes representing the maximum duration observed.

2.4 Feather score and feet health assessment

At 30 and 64 weeks of age, after the TI test, the feather condition score and feet health were assessed on all the animals present in each pen.

Feather score was assessed based on the protocol by Tauson et al. (2006) on six body parts, including the neck, breast, cloaca, back, wing, and tail. Feather scores ranging from 1 to 5 were used to indicate the extent of coverage, with the following criteria: score 1, minimal coverage (<25% coverage); score 2, 25%–50% coverage; score 3, 50%–75% coverage; score 4, >75% coverage; and score 5, complete coverage (Lai et al., 2010).

The footpad dermatitis (FPD) was assessed and scored using the methods of Rushen, Butterworth, and Swanson: score 0 = no lesion or slight discoloration of the skin, or healed lesion; score 1 = mild lesion, characterized by superficial discoloration of the skin and hyperkeratosis; score 2: severe lesion, involving the affected epidermis, presence of blood scabs, hemorrhages, and significant swelling of the skin (Rushen et al., 2011).

2.5 Data Analysis

The average percentage of each behaviour and the average duration (in minutes) of TI test, expressed as mean ± standard deviation, was calculated for each genotype (BP, BPxS, RM, RMxS, and LB), housing system (free-range and enriched cage), and age (30 and 64 weeks). The data were assessed for normality using the Shapiro-Wilk test. To assess the effects of genotype, housing system, age, and their interactions on behaviours and TI test, a two-way analysis of variance (ANOVA) was employed. Multiple comparisons of the means were conducted using the Duncan test, and the least significant difference was calculated.

The presence of behavioural patterns in the enriched cage and free-range systems was analyzed using an unsupervised technique for dimensionality reduction: the t-Stochastic Neighbor Embedding (t- SNE) technique. t-SNE is an unsupervised, non-linear machine learning technique primarily used for data exploration and visualizing high-dimensional data (van der Maaten and Hinton, 2008). This method allows to visualize the resulting similar data and thus, to identify behavioural patterns in the two housing systems. In particular, the focus was set on the link between the 16 analyzed behaviours and the genotype, the age, and the two housing systems with t-SNE technique. The behavioural data set was converted into a matrix of pairwise similarities, and as an input dataset was used a set of vectors where each vector described the behavioural features of each behavioural observation.

The absolute and relative frequencies for the feather and FPD scores were calculated. To assess differences based on genotype, housing system, and age, a chi-squared test was conducted.

Statistical analyses were performed using R software, Version 3.1.2, with a significance level of $p \le 0.05$ considered statistically significant. The t-SNE technique was performed using Python Software.

3. Results

3.1 Behavioural observations

The effects of genotype and housing system on the behaviours of laying hens are reported in Table 3. The housing system had a significant impact on the behaviour of laying hens, affecting all the behaviours considered in this study (p < 0.001), whereas the influence of the hens' genotype and the interaction between genotype and housing system was evident only in specific behaviours.

Specifically, laying hens in the free-range system displayed higher frequencies of behaviours like 'pecking', 'fighting', 'crouch', 'self-preening', 'leg stretching', except for 'feeding at feeders' and 'wing stretching', which were performed more frequently in the enriched cage system. Significant differences were observed in various behaviours among different genotypes, including 'scratching', 'feeding at feeders', 'grass feeding and object pecking', 'running', 'allo-preening', 'fighting', 'stand', 'sand bath', and 'self-preening'. All the behaviours related to feeding category exhibited variations across genotypes. In particular, the LB demonstrated a higher frequency of 'feeding at feeders' behaviour compared to the other genotypes. Conversely, LB showed a relatively lower occurrence of 'grass feeding and object pecking'. Specifically, our analysis revealed significant differences in the 'scratching' behaviour among genotypes (p < 0.001), as well as in their interaction with the housing system (p < 0.001). RM and BP exhibited the highest percentages of 'scratching', while LB displayed the lowest percentage of 'scratching'.

Under the category movement behaviours, a significant effect of genotype and a significant interaction between genotype and housing system was only found for 'running' (p = 0.003 and p < 0.001, respectively).

When considering differences among genotypes, our analysis indicated that LB exhibited a reduced frequency of 'running' behaviour, whereas RM and RMxS demonstrated higher percentages of 'running'. Social behaviours, i.e., 'allo-preening' and 'fighting,' were significantly affected by genotype (p = 0.03 and p < 0.001, respectively). LB spent less time in social behaviours compared to the other genotypes. Additionally, the interaction effect between genotype and housing system significantly influenced these behaviours (p < 0.001). This suggests that the influence of genotype on these social behaviours may vary based on the specific housing environment, indicating a potential interplay between the environmental context and the genetic traits of the hens. In terms of resting category, the 'crouch' behaviour was primarily influenced by the housing system, while the 'stand' behaviour was affected by both genotype and housing system (p < 0.001). Specifically, LB displayed the highest percentage of the 'stand' behaviour compared to the other genotypes. Both the comfort behaviours (i.e., 'sand bath' and 'self-preening') were influenced by the genotype (p = 0.01 and p <0.001, respectively) and its interaction with the housing system (p < 0.001), with LB that showed the lowest percentage of these behaviours. No significant differences were found between the age of the animals in terms of the percentage of behaviours. For a detailed analysis of the percentages of behaviour including the age and their interactions, refer to the Table S1 reported in the supplementary material.

3.2 t-SNE behavioural pattern

The influence of the housing system on behavioural patterns was also demonstrated using the t-SNE technique. This method processed each behavioural observation into a feature vector containing the frequency of the 16 observed behaviours. These 16-dimensional vectors were then visually represented using t-SNE, a dimensionality reduction technique that transforms the 16-dimensional data into a more easily interpretable 2-dimensional space (X1 and X2 axes; see Figure 1). This visualization aimed to provide a clearer

understanding of how the housing system influenced the overall behavioural characteristics of the hens. Two defined clusters of observations with similar behavioural patterns are visible. The top-right cluster contains all the observations registered for the enriched cage system, and the bottom-left cluster contains all the observations registered for the free-range system (Figure 1, right panel). The clear separation into distinct clusters suggests a notable divergence in behavioural patterns between the enriched cage and free-range systems. On color coding by genotype, both clusters contain the five genotypes, and LB, in both clusters is the genotype that shows more similar behaviours (Figure 1, left panel, yellow points). Each cluster exhibits closely grouped yellow dots representing LB, evident in both the free-range and enriched cage clusters. This indicates that LB shows the highest similarity in behaviours across both systems.

On color coding by age, similar behaviour patterns are not identified (Figure 1, middle panel). In Figure 2, the frequency and the type of every single behaviour displayed by the animals is highlighted (darker color shows higher frequency than light color). In the free-range system, the behaviours belonging to the movement category (i.e., 'walking', 'running' and 'wing flap') are performed more frequently. On the contrary, the behaviours belonging to the stretching category (i.e., 'leg stretching' and 'wing stretching') and social behaviours as 'pecking' and 'fighting', are performed more frequently in the enriched cage system.

3.2 Tonic immobility test

Table 4 presents the results of the statistical analysis for the TI test. The average duration of TI, measured in minutes and expressed as mean \pm standard deviation, is provided for each attempt. No significant differences were observed among the five genotypes in the duration of the TI test for each attempt. However, when considering the age of the laying hens, a significant effect was found on the second and third attempts. At 64 weeks, the birds displayed a longer duration of TI during the second attempt (p = 0.028), whereas during the

third attempt, the youngest birds (30 weeks old) exhibited an increased duration of TI (p < 0.001). Additionally, we observed an interaction between age and the housing system on the second and third attempts (p = 0.003 and p = 0.010 respectively). This implies that the effects of age may exhibit variations in different housing environments or conditions, indicating a potential interplay or relationship between age and housing system factors. For a detailed analysis of the TI test attempts, including genotypes and their interactions, refer to the Supplementary material (Table S4).

3.3 Feather score and feet health assessment

The results of the feather score and the assessment of feet health are reported in Table 5. The housing system had a significant influence on both feather and FPD scores. Specifically, the laying hens housed in the enriched cage system exhibited poorer plumage and feet condition across all body parts compared to those in the free-range housing system (neck: p < 0.001; breast: p < 0.001; cloaca: p < 0.001; back: p < 0.001; wing: p < 0.001; tail: p < 0.001). Additionally, age was found to have an impact on the feather score, specifically in the back and cloaca regions (back: p = 0.012; cloaca: p = 0.03). No differences were found among the five genotypes in terms of feather and FPD scores.

4. Discussion

In summary, our findings underscore the profound impact of the housing system on the behaviour, plumage condition, and foot health of laying hens. The free-range system allows the hens to perform a broader range of behaviours, like movement, comfort, and social interactions. Additionally, hens in the free-range system exhibit improved plumage conditions and lower rates of footpad dermatitis, signifying a positive influence on their overall welfare as compared to the hens in the enriched cages. Furthermore, all genotypes

demonstrate a similar behavioural pattern within the two housing systems. Notably, the age of the animals had minimal impact on the ethogram, plumage condition, or foot health, except for the results of the TI test. In our study, we employed both a two-way analysis of variance (ANOVA) and the t-Stochastic Neighbor Embedding (t-SNE) technique. Employing these diverse analytical methods aims to provide a more comprehensive understanding of the complex behavioural patterns exhibited by the hens, especially when dealing with a larger set of behavioural variables such as the 16 behaviours scored in the present study. Our results demonstrate that the behavioural patterns exhibited by the hens are influenced by the housing system, with age and genotype having minimal effects. Despite expectations that the LB genotype might adapt more to cage systems and others to free-range systems, the observed behavioural responses across genotypes are quite similar. Indeed, despite behavioural differences among the various genotypes, all genotypes appear to demonstrate adaptability to both housing systems. However, for a thorough understanding of hen behaviour, a detailed study of individual behaviours, emphasizing the frequency and specific types, is essential.

The higher frequency of movement behaviours observed in the free-range system, including 'walking,' 'running,' and 'wing flapping,' can be attributed to the increased space and freedom available to the hens (Appleby, 2004). On the other hand, the lower frequency of 'running' behaviour exhibited by the LB genotype suggests that certain genotypes may be less inclined to engage in this specific behaviour. These results support the theory of energy conservation, suggesting that the higher kinetic activity of slow-growing animals reduces available body energy for productive performance(Castellini et al., 2012; Castellini and Bosco, 2017; Fanatico et al., 2007). Additionally, as highlighted by Schütz and Jensen (Schütz and Jensen, 2001), selection for high production rates can lead to modified behaviour. The free-range system provides a more spacious and natural environment, allowing the hens to engage in these movement activities more frequently. These

results emphasize the impact of housing conditions, particularly the lack of space in the enriched cage system, on the expression of movement behaviours in laying hens. Providing hens with adequate space and environmental enrichment in enriched cages could potentially encourage more natural and active behaviours, contributing to their overall welfare (Xu et al., 2022).

In terms of the feeding category, it was observed that in the enriched cage system, laying hens spent more time engaging in feeding behaviour. This aligns with expectations, as the restricted environment of the enriched cage system encourages the birds to utilize the provided feeders for their feeding activities. On the other hand, in the free-range system, laying hens displayed a higher frequency of 'grass feeding and object pecking' behaviour. The presence of natural elements and objects in the free-range environment allows the hens to engage in foraging behaviours, such as pecking at grass and objects, aligning more closely with their natural feeding behaviour. This highlights the importance of providing hens with an extensive system rather than confining them to cages, significantly contributing to their welfare. Moreover, the extent to which animals can manifest their natural behaviours is deemed a cornerstone of animal welfare. This underscores the significance of creating environments conducive to such behaviours. Allowing animals to engage in behaviours typical of their natural habitats enhances their physical and psychological health, thus elevating overall standards of animal welfare (Colditz, 2023). This perspective underscores the necessity of comprehending and encouraging natural behaviours in assessing and implementing animal welfare protocols, encompassing the principles of the Five Freedoms and the Five Domains of Welfare (Brambell, 1965; Mellor et al., 2020).

The housing system significantly impacted resting behaviours, especially the 'crouching' behaviour, as indicated by our findings. Laying hens within the enriched cage system tended to display a higher frequency of 'crouching' behaviour, along with restricted movement. This behaviour may be associated with increased

instances of footpad dermatitis and poorer feather quality, particularly in the breast area, as hens tend to rest more in that position (Göransson et al., 2023; Meluzzi and Sirri, 2009).

The higher occurrence of social behaviours such as 'pecking' and 'fighting' in the enriched cage system can be attributed to several factors. Firstly, the lack of environmental stimuli and limited space in enriched cages may contribute to increased aggression among the confined animals (Nicol et al., 1999). This aggressive behaviour may have implications for the plumage status of the hens, particularly in areas such as the neck, tail, and wings. The higher stocking density in cage systems compared to free-range systems can further escalate social interactions and competition for resources. This is also confirmed by a greater amount of time lying down in the enriched cage system, where the animals seem to be less active and interested in their surroundings, reducing object pecking behaviour (Koelkebeck et al., 1987). Additionally, the cage flooring itself can contribute to the development of footpad dermatitis.

Regarding genotype differences, our analysis revealed that the LB genotype exhibited less engagement in social behaviours, specifically 'allo-preening' and 'fighting', compared to the other genotypes. This suggests a potential genetic influence on social behaviour tendencies in laying hens. The LB genotype may have inherent traits or characteristics that contribute to a decreased inclination for social interactions, resulting in less time spent on allo-preening and engaging in fights (Wurtz et al., 2022). Furthermore, it is conceivable that social behaviour has been influenced by selection in LB, potentially aimed at preventing injurious pecking behaviour within larger flocks.

The higher frequency of stretching observed in the enriched cage system can be interpreted as a compensatory behaviour for the lack of movement. Chickens often engage in stretching activities involving their wings and legs when they experience muscle stiffness or joint pain, potentially resulting from inadequate physical exercise (Albentosa and Cooper, 2004; Engel et al., 2019; Hughes and Duncan, 1988;

Nicol, 1987). This behaviour may be interpreted as a response to discomfort, indicating an attempt to alleviate potential muscle and joint tension or discomfort experienced by the chickens.

The influence of the housing system, genotype, and their interaction on comfort behaviours, specifically 'sand bath' and 'self-preening', is evident from our analysis. The LB genotype exhibited the lowest percentage of these behaviours, indicating potential differences in their expression across genotypes. However, it is important to interpret the results of self-preening behaviour cautiously. While self-preening is a natural behaviour for chickens to maintain feather condition and cleanliness, excessive self-preening can be indicative of underlying anxiety or stress. In environments where animals are unable to express freely or are subjected to confined spaces, they may engage in excessive comfort behaviours as a coping mechanism, which can be described as stereotypical behaviour (Broom, 1983).

Moreover, the evident disparity between the two housing systems, as evidenced by the analysis of FPD and feather condition scores, underscores how the free-range system better fulfills the birds' requirements. For instance, it offers a superior flooring that mitigates the occurrence of FPD and enables chickens to express their natural behaviours more freely, thereby reducing the likelihood of feather pecking. In fact, all the animals housed in the free-range system didn't present any FPD nor feather damage contrary to what was found in the enriched cages system, as previously reported also by other authors (Erensoy et al., 2021; Rørvang et al., 2019; Wei et al., 2020). The lack of differences in TI results showed how the examined genotypes did not exhibit distinct variations in their responses to the test. Contrary, a consistent trend towards greater adrenocortical activation was reported in high-fear hens, which showed long TI reactions when compared to low-fear counterparts (Beuving et al., 1989). Interestingly, hen's age did not affect general behaviours, but it did influence the response in the TI test. TI in chickens serves as a fear indicator, with susceptibility changing during development and being influenced by various factors like exposure to

threatening stimuli (Tiemann et al., 2023). However, the effectiveness of TI as a fear indicator, especially concerning age, is debated. Studies suggest that TI duration in chickens can be modulated by environmental and individual factors, with age impacting susceptibility to induction procedures. Additionally, genetic predispositions inherited in different chicken breeds lead to substantial variation in fear responses, indicating that TI might not be universally effective across all ages and breeds (Carli and Farabollini, 2022).

These results highlight the complex nature of stress response in laying hens, indicating that age can play a role in physiological reactions to the environment. Our findings underscore the importance of continually evaluating and refining housing systems to ensure the welfare of laying hens. Efforts should be made to strike a balance between productivity and animal welfare, considering the natural behavioural needs and health requirements of the birds.

5. Conclusions

In conclusion, our research has revealed differences in the adaptability of different chicken genotypes to various farming systems. While it may initially seem that all genotypes adapt well, a deeper analysis revealed that the enriched cage system can restrict natural behaviours and have adverse effects on feather and leg health. These findings underscore the importance of carefully assessing the specific impacts of farming systems on hens to develop more sustainable poultry practices that prioritize animal welfare. Analyzing chicken behaviour enables early problem detection and implementation of preventive measures. Prioritizing animal welfare by farmers, the poultry industry, and consumers is essential for advocating ethical practices and alternative farming systems, ensuring humane treatment of birds and the industry's long-term sustainability.

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Institutional Review Board Statement

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Xu, D., Shu, G., Liu, Y., Qin, P., Zheng, Y., Tian, Y., Zhao, X., Du, X., 2022. Farm Environmental Enrichments Improve the Welfare of Layer Chicks and Pullets: A Comprehensive Review. Animals 12. https://doi.org/10.3390/ani12192610 Table 1. Experimental design of trial and animal distribution in the farming systems.

Construits	Enriched cage	Free-Range
Genotype	(0.144 m²/hen)	(1.066 m²/hen)
Lohman Brown	3 pens with 15 hens/pen	3 pens with 5 hens/pen
Bionda Piemontese	3 pens with 15 hens/pen	3 pens with 5 hens/pen
Robusta Maculata	3 pens with 15 hens/pen	3 pens with 5 hens/pen
Bionda Piemontese × Sasso	3 pens with 15 hens/pen	3 pens with 5 hens/pen
Robusta Maculata × Sasso	3 pens with 15 hens/pen	3 pens with 5 hens/pen

Table 2. Laying hen ethogram for behavioural classification. The ethogram is divided into six macro-categories containing specific behaviours.

Category	Behaviour	Description								
Feeding	Scratching	Move the litter/dirt with claws (Biasato et al., 2022)								
	Feeding at feeders	Eating feed from feeders (Jhetam et al., 2022)								
	Grass feeding/	Eating grass in outdoor space/pecking object indoor								
	object pecking	(Veldkamp and Niekerk, 2019)								
	Walking	Walking at slow speed (Rieke et al., 2021)								
Movomont	Running	Running (Veldkamp and Niekerk, 2019)								
WOVEINEII	Wing flapping	Open wings wide and flapping up and down while walking or								
		running (Liu et al., 2020)								
	Menacing	Initiate fight with another chicken (Kayla et al., 2023)								
Social	Fighting	Fighting between two or more chickens(Veldkamp and								
	i igritirig	Niekerk, 2019)								

		Journal Pre-proof
	Pecking	Pecking movements directed at a pen mate (McCowan et al.,
	recking	2006)
	Allo-preening	Social preening (Kenny et al., 2017)
Resting	Crouch	Sitting down (Webster, 2000)
Resting	Stand	Holding standing position (Veldkamp and Niekerk, 2019)
	Sand bath	Rolling or moving around in dust, dry earth, or sand (Grebey
Comfort	Sand bath	et al., 2020)
connort	Colf prooping	Self-feathers grooming by means of beak (McCowan et al.,
	Sen-preening	2006)
	Log stratabing	Putting the leg in a certain position to lengthen and elongate
	Leg stretching	the muscle (Biasato et al., 2022; Carvalho et al., 2022)
Stretching		Putting the wing in a certain position to lengthen and elongate
	wing stretching	the muscle (Biasato et al., 2022; Carvalho et al., 2022)

Table 3. Effect of genotype (G) and housing system (HS) and their interaction on percentage of behaviours. Mean \pm standard deviation of the behaviours is indicated. Means with letters (a, b, ab) denote significant differences (p \leq 0.05).

		G	Syster	n (HS)	p-value					
Behaviour	LB	ВР	BPxS	RM	RMxS	Free- range	Enriched cage	G	HS	GxHS
Scratching	2.51±3.72 (b)	5.92±6.77 (a)	4.61±5.07 (ab)	6.23 <u>±</u> 6.88 (a)	4.38±5.16 (ab)	9.47±4.64 (a)	0 (b)	<0.002	1<0.001	L<0.001
Feeding at feeders	19.08±10.08 (b)	13.27 <u>+</u> 10.31 (a)	13.41 <u>+</u> 11.37 (a)	713.29 <u>+</u> 10.95 (a)	12.66±10.25 (a)	5.21 <u>+</u> 4.16 (a)	23.48 <u>+</u> 7.02 (b)	0.001	<0.001	L 0.23
feeding/object pecking	7.25 <u>+</u> 4.58 (b)	8.42 <u>+</u> 6.47 (ab)	9.69 <u>+</u> 6.58 (ab)	9.46 <u>+</u> 6.26 (ab)	10.90 <u>+</u> 7.99 (a)	13.92 <u>+</u> 5.38 (a)	4.37 <u>+</u> 3.37 (b)	0.012	<0.001	L<0.001
Walking	13.75±11.68	11.07 <u>+</u> 8.98	11.23 <u>+</u> 8.98	10.05±7.57	10.78 <u>+</u> 7.22	18.65 <u>+</u> 6.77 (a)	4.10 <u>+</u> 3.52 (b)	0.14	<0.001	L<0.001
Running	1.88±2.60 (b)	3.86 <u>+</u> 4.57 (ab)	3.83 <u>+</u> 4.64 (ab)	4.35 <u>+</u> 5.31(a)	4.72±5.72 (a)	7.47 <u>±</u> 4.19 (a)	0 (b)	0.003	<0.001	L<0.001
Wing flap	1.99 <u>+</u> 2.99	1.43 <u>+</u> 2.25	1.25±2.08	1.43 <u>+</u> 2.25	1.44 <u>+</u> 2.23	3.02 <u>+</u> 2.61 (a)	0 (b)	0.40	<0.001	L 0.14
Menacing	0	0.10±0.49	0.04±0.33	0.19 <u>+</u> 0.71	0.08±0.48	0.16 <u>±</u> 0.65 (a)	0 (b)	0.14	<0.001	L 0.13
Allo-preening	0.91 <u>+</u> 1.57 (b)	2.03 <u>+</u> 2.92 (ab)	1.85 <u>+</u> 2.79 (ab)	1.98 <u>+</u> 2.74 (ab)	2.14 <u>+</u> 2.86 (a)	3.57 <u>+</u> 2.76 (a)	0 (b)	0.03	<0.001	L<0.001

Pecking	4.41 <u>+</u> 4.97	3.20 <u>+</u> 3.60	4.48 <u>+</u> 4.39	3.80 <u>+</u> 3.78	3.48 <u>+</u> 3.89	1.41±1.54	6.34 <u>+</u> 4.49	0.26 <0	0.001 0.051
-						(a)	(d)		
Fighting	0 (b)	3.36+4.26 (a)	2.97 <u>+</u> 4.36	3.26 <u>+</u> 4.41	2.94 <u>+</u> 4.09	0 (a)	5.02 ± 4.43	<0.001<0	.001<0.001
	- (-)		(a)	(a)	(a)	- (-)	(b)		
Crouch	10 20+8 56	9 03+7 82	7 98+6 44	7 54+6 78	9 24+7 76	3.03±2.49	14.57 ± 6.38	0.22 <0	
crouch	10.20-0.50	5.05 17.02	7.50-0.44	7.54 <u>+</u> 0.70	J.24 <u>1</u> 7.70	(a)	(b)	0.22 <0	
Stand	20.23±10.22	15.60±7.54	15.82 ± 8.76	15.58 ± 6.76	14.59 ± 6.09	20.55±8.00	12.18±5.99	~0 001 ~0	001 < 0 001
Stanu	(b)	(a)	(a)	(a)	(a)	(a)	(b)	0.001<0	.001<0.001
Cand bath	1.72 <u>+</u> 2.54	3.28 <u>+</u> 3.96	3.48±4.15	3.86±4.62	3.52 <u>+</u> 4.30	6.35±3.62	0 (h)	0.01 -0	001 -0 001
Sand bath	(b)	(ab)	(ab)	(a)	(ab)	(a)	0 (0)	0.01 <0	0.001<0.001
Solf prooping	5.36±4.49	0.21+6.60 (2)	9.73±6.85	8.74±6.56	9.44 <u>+</u> 6.24	5.52±4.29	11.51±6.72	~0 001 ~0	001 < 0 001
Sell-bleening	(b)	9.51 <u>+</u> 0.00 (a)	(a)	(a)	(a)	(a)	(b)	<0.001<0	.001<0.001
	2 0 0 1 5 4 0	2 57 4 04	2 26 4 70	2 20 1 4 62	2.80±3.97	0 (1)	6 72 4 6 6	0.70.00	004 0 07
Leg stretcning	3.86±5.18	3.57 <u>+</u> 4.81	3.26 <u>+</u> 4.70	3.29 <u>+</u> 4.63	(a)	(d) U	6.72 <u>+</u> 4.58	0.72 <0	0.001 0.37
Wing stretching	6.76±5.74	6.46 <u>+</u> 6.32	6.29 <u>+</u> 6.24	6.89 <u>+</u> 6.74	6.81 <u>+</u> 6.47	1.61 <u>+</u> 1.61 (a)	11.67±5.07 (b)	0.97 <0	0.001 0.72
-									

LB: lohman brown; BP: bionda piemontese; BPxS: bionda piemontese x sasso; RM: robusta maculata; RMxS: robusta maculata x sasso.

Table 4. Effect of age and housing system (HS) and their interaction on tonic immobility test attempts. Mean and standard deviation of the minutes are indicated. Significant p-values are indicated in bold. Means with letters (a, b) denote significant differences ($p \le 0.05$).

Tonic immobility	Ag	ge	Housing Sys	p-value			
attempt	30 weeks	64 weeks	Free-range	Enriched cage	Age (A)	HS	AxHS
First	0.42 <u>+</u> 0.96	0.63±1.17	0.53 <u>±</u> 1.09	0.51 <u>+</u> 1.05	0.19	0.92	0.480
Second	0.89 <u>+</u> 1.30 (a)	1.32 <u>+</u> 1.36 (b)	1.27 <u>+</u> 1.36	0.94 <u>+</u> 1.32	0.028	0.09	0.003
Third	1.09±01.32 (a)	0.48 <u>±</u> 1.06 (b)	0.65±1.18	0.92 <u>+</u> 1.27	<0.001	0.14	0.010

Table 5. Absolute and relative frequencies of scores assigned to feather condition score and footpad

dermatitis (FDP) of laying hens. Significant chi-square test results are highlighted in bold.

Genotype (G)					System	(HS)	Age (A)	
	ВР	BPxS	LB	RM	RMxS	Free-range	Enriched cage	30 weeks 64 weeks

		Dear tama at	
618166	121		
JUUII.		110 proof	

					Neck				
1	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)
2	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)
3	18(50%)	20(55%)	20(55%)	19(53%)	19(53%)	6(7%)	90(100%)	48(53%)	48(53%)
4	18(50%)	16(45%)	16(45%)	17(47%)	17(47%)	84(93%)	0(0%)	42(47%)	42(47%)
		X ² = 0.	31; p-value =	= 0.98		X² = 153.77; p-v	value <0.001	X²=0; p-v	value = 1
					Breast				
1	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)
2	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)
3	18(50%)	19(53%)	18(50%)	18(50%)	18(50%)	1(2%)	90(100%)	46(51%)	45(50%)
4	18(50%)	17(47%)	18(50%)	18(50%)	18(50%)	89(98%)	0(0%)	44(49%)	45(50%)
		X ² = 0.	08; p-value =	= 0.99		X² = 172.11; p-v	value <0.001	X ² = 0; p-	value = 1
					Cloaca				
1	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)
2	2(6%)	2(6%)	1(3%)	0(0%)	1(3%)	0(0%)	6(7%)	0(0%)	6(7%)
2	16(44%)	17(47%)	18(50%)	19(53%)	18(50%)	17	84(93%)	47(52%)	40(44%)
4	18(50%)	17(47%)	17(47%)	17(47%)	18(50%)	18(50%)	0(0%)	43(48%)	44(49%)
•	10(0070)	$X^2 = 0.$	08: p-value =	= 0.99	10(00/0)	$X^2 = 168.41$: p-v	value <0.001	X ² =6.57: p-	value=0.03
		-			Back				
4	0(0)()	0(00()	0(00()	0(0)()	0(00()	0(00()	0(0)()	0(0)()	0(00()
1	0(0%)	0(0%)	0(0%)	0(0%)		0(0%)	0(0%)	0(0%)	0(0%)
2	U(U%)	3(8%)	1(3%)	2(6%)	2(6%)	0(0%)	8(9%)	8(9%)	
3	19(53%)	15(44%)	17(47%)	10(44%)	10(44%)	2(2%)	82(91%)	38(42%)	40(51%)
4	17(47%)	1/(48%) $x^2 = 2$	18(50%) 72: n voluo -	18(50%)	18(50%)	88(98%)	0(0%)	44(49%) V ² -0 76, p.	44(49%)
		∧ = J.	72, p-value -	- 0.88		Λ = 1/2.19, p-	Value <0.001	Λ -0.70, p-v	alue-0.012
					Wing				
1	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)
2	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)
3	18(50%)	19(53%)	18(50%)	19(53%)	19(53%)	3(3%)	90(100%)	47(52%)	46(51%)
4	18(50%)	17(47%)	18(50%)	17(47%)	17(47%)	87(97%)	0(0%)	43(48%)	44(49%)
		$X^2 = 0.$	133; p-value	= 0.9		X ² = 164.53; p-v	value <0.001	X ² = 0; p-	value = 1
					Tail				
1	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)	0(0%)
2	5(14%)	3(8%)	2(6%)	5(14%)	6(17%)	0(0%)	21(23%)	7(8%)	14(16%)
3	14(39%)	15(42%)	16(44%)	14(39%)	12(33%)	2(2%)	69(77%)	39(53%)	32(35%)
4	17(47%)	18(50%)	18(50%)	17(47%)	18(50%)	88(98%)	0(0%)	44(49%)	44(49%)
		X ² = 3.	26; p-value =	= 0.91		X ² = 172.22; p-v	value <0.001	X ² =3.02; p-v	value = 0.22
				Footpa	ad dermat	titis			
0	20(55%)	19(53%)	19(53%)	20(55%)	16(44%)	85(94%)	9(10%)	48(53%)	46(51%)
1	15(42%)	15(42%)	17(47%)	15(42%)	19(53%)	5(6%)	76(84%)	42(47%)	39(43%)
2	1(3%)	2(5%)	0(0%)	1(3%)	1(3%)	0(0%)	5(6%)	0(0%)	5(6%)
		X ² = 3.	36; p-value =	= 0.91		X² =128.68; p-v	alue <0.001/	X ² =5.15; p-	value = 0.07

LB: lohman brown; BP: bionda piemontese; BPxS: bionda piemontese x sasso; RM: robusta maculata; RMxS: robusta maculata x

Figure 1. Behavioural patterns of laying hens using t-Distributed Stochastic Neighbor Embedding (t-SNE) to map high-dimensional behavioural vectors to a 2-dimensional feature space X1 and X2 axes). Each point corresponds to a behavioural observation. Observations with similar behavioural vectors are mapped to neighboring points in the plane. Points (observations) are color coded according to genotypes (left panel), age of animals (middle panel), and housing systems (right panel). LB: lohman brown; BP: bionda piemontese; BPxS: bionda piemontese x sasso; RM: robusta maculata; RMxS: robusta maculata x sasso.



Figure 2. Behavioural patterns of laying hens using t-Distributed Stochastic Neighbor Embedding (t- SNE) to map high-dimensional behavioural vectors to a 2-dimensional feature space X1 and X2 axes). (Top cluster enriched cage system, bottom cluster free-range system) Each point corresponds to a behavioural observation. Observations with similar behavioural vectors are mapped to neighboring points in the plane. The frequency and type of each behaviour exhibited by the hens are depicted, with darker colors indicating higher frequencies and lighter colors indicating lower frequencies. Behaviours are sorted from most to least performed by the animals.



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Table 4. Effect of age and housing system (HS) and their interaction on tonic immobility test attempts. Mean and standard deviation of the minutes are indicated. Significant p-values are indicated in bold. Means with letters (a, b) denote significant differences ($p \le 0.05$).

Tonic immobility	А	ge	Housing Syst	tem (HS)	p-value		
attempt	30 weeks	64 weeks	Free-range	Enriched cage	Age (A)	HS	AxHS
First	0.42 <u>±</u> 0.96	0.63±1.17	0.53±1.09	0.51±1.05	0.19	0.92	0.480
Second	0.89±1.30 (a)	$1.32{\pm}1.36$ (b)	1.27±1.36	0.94±1.32	0.028	0.09	0.003
Third	1.09±01.32 (a)	$0.48 {\pm} 1.06$ (b)	0.65 ± 1.18	0.92±1.27	<0.001	0.14	0.010

Table 5. Absolute and relative frequencies of scores assigned to feather condition score and

Genotype (G) System (HS) Age (A) Enriched BP **BPxS** RM **RMxS** 30 weeks 64 weeks LB **Free-range** cage Neck 1 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 2 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 90(100%) 3 18(50%) 20(55%) 20(55%) 19(53%) 19(53%) 6(7%) 48(53%) 48(53%) 4 18(50%) 16(45%) 16(45%) 17(47%) 17(47%) 84(93%) 0(0%) 42(47%) 42(47%) X² = 0.31; p-value = 0.98 X² = 153.77; p-value < 0.001 X²=0; p-value = 1 Breast 1 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 2 0(0%) 0(0%) 90(100%) 3 18(50%) 19(53%) 18(50%) 18(50%) 18(50%) 1(2%) 46(51%) 45(50%) 4 18(50%) 89(98%) 0(0%) 44(49%) 18(50%) 17(47%) 18(50%) 18(50%) 45(50%) X² = 0.08; p-value = 0.99 X² = 172.11; p-value < 0.001 $X^{2} = 0$; p-value = 1 Cloaca 0(0%) 0(0%) 1 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 2 2(6%) 2(6%) 1(3%) 0(0%) 1(3%) 0(0%) 6(7%) 0(0%) 6(7%) 3 16(44%) 17(47%) 18(50%) 19(53%) 18(50%) 17 84(93%) 47(52%) 40(44%) 4 18(50%) 17(47%) 17(47%) 17(47%) 18(50%) 18(50%) 0(0%) 43(48%) 44(49%) X² = 0.08; p-value = 0.99 X² = 168.41; p-value < 0.001 X²=6.57; p-value=0.03 Back 0(0%) 1 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 2 8(9%) 0(0%) 3(8%) 1(3%) 2(6%) 2(6%) 0(0%) 8(9%) 0(0%) 3 19(53%) 16(44%) 17(47%) 16(44%) 16(44%) 2(2%) 82(91%) 38(42%) 46(51%) 17(47%) 17(48%) 4 18(50%) 18(50%) 18(50%) 88(98%) 0(0%) 44(49%) 44(49%) $X^2 = 3.72$; p-value = 0.88 X² = 172.19; p-value < 0.001 X²=8.76: p-value=0.012 Wing 0(0%) 0(0%) 1 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 2 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 3 18(50%) 19(53%) 18(50%) 19(53%) 19(53%) 3(3%) 90(100%) 47(52%) 46(51%) 4 18(50%) 17(47%) 18(50%) 17(47%) 17(47%) 87(97%) 0(0%) 43(48%) 44(49%) X² = 0.133; p-value = 0.9 X² = 164.53; p-value < 0.001 $X^{2} = 0$; p-value = 1 Tail 0(0%) 0(0%) 1 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 0(0%) 2 5(14%) 3(8%) 2(6%) 5(14%) 6(17%) 0(0%) 21(23%) 7(8%) 14(16%) 3 14(39%) 15(42%) 16(44%) 14(39%) 12(33%) 2(2%) 69(77%) 39(53%) 32(35%) 4 17(47%) 18(50%) 18(50%) 17(47%) 18(50%) 88(98%) 0(0%) 44(49%) 44(49%) X² = 3.26; p-value = 0.91 X² = 172.22; p-value < 0.001 X² =3.02; p-value = 0.22

footpad dermatitis (FDP) of laying hens. Significant chi-square test results are highlighted in bold.

	X ² = 3.36; p-value = 0.91						value <0.001	X ² =5.15; p-v	alue = 0.07/
2	1(3%)	2(5%)	0(0%)	1(3%)	1(3%)	0(0%)	5(6%)	0(0%)	5(6%)
1	15(42%)	15(42%)	17(47%)	15(42%)	19(53%)	5(6%)	76(84%)	42(47%)	39(43%)
0	20(55%)	19(53%)	19(53%)	20(55%)	16(44%)	85(94%)	9(10%)	48(53%)	46(51%)

LB: lohman brown; BP: bionda piemontese; BPxS: bionda piemontese x sasso; RM: robusta maculata; RMxS: robusta

maculata x sasso.

1. Declaration of interests

 \boxtimes The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as

potential competing interests:

Highlights

- Behaviour of 5 laying hen genotypes in enriched cage and free range housing systems washas been studied.
- Housing system affects behaviour, plumage status, and footpad dermatitis of laying hens.
- Free range system offers more opportunities to perform a wide range of behaviours.
- Significant behavioural differences were observed between the commercial hybrid.
- Behavioural patterns were identified using a machine learning technique