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Medial anterior prefrontal cortex stimulation down-regulates implicit reactions to threats and prevents the return of fear

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Information about the manuscript

Medial anterior prefrontal cortex stimulation down-regulates implicit reactions to threats and prevents the return of fear

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14 Abstract

15 Down-regulating emotional overreactions toward threats is fundamental for developing treatments for anxiety and post-traumatic disorders. The prefrontal cortex (PFC) is critical for top-down 16 modulatory processes, and despite previous studies adopting repetitive Transcranial Magnetic 17 Stimulation (rTMS) over this region provided encouraging results in enhancing extinction, no 18 studies have hitherto explored the effects of stimulating the medial anterior PFC (aPFC, 19 20 encompassing the Brodmann area 10) on threat memory and generalization. Here we showed that rTMS over the aPFC applied before threat memory retrieval immediately decreases implicit 21 reactions to learned and novel stimuli in humans. These effects enduringly persisted one week later 22 23 in the absence of rTMS. No effects were detected on explicit recognition. Critically, rTMS over the aPFC resulted in a more pronounced reduction of defensive responses compared to rTMS targeting 24 the dorsolateral PFC. These findings reveal a previously unexplored prefrontal region, the 25 26 modulation of which can efficiently and durably inhibit implicit reactions to learned threats. This represents a significant advancement towards the long-term deactivation of exaggerated responses 27 28 to threats.

29 Introduction

30 Emotional memories related to past threat experiences allow humans to predict future dangers and trigger adaptive defensive reactions when encountering learned threat-signaling cues¹. However, 31 extremely dangerous situations may lead to psychological disorders². Furthermore, the ability to 32 generalize defensive reactions to new stimuli enables organisms to anticipate potential threats and 33 respond to them based on similar perilous experiences lived in the past. On the other hand, 34 evaluation mechanisms excessively biased toward threat generalization (i.e. overgeneralization) 35 may underlie anxiety disorders and trauma³. At the base of these processes, in a previous work⁴ we 36 observed that autonomic-implicit and cognitive-explicit tunings may diverge when humans are 37 38 exposed to the same new stimuli, where cognitive generalization may enable a flexible evaluation of incoming cues to develop adaptive predictions of potential dangers. The crosswise presence of 39 overgeneralization in anxiety diseases, and the dissociation between autonomic and cognitive 40 defensive response patterns, highlight the importance of including both implicit and explicit 41 generalization tasks to characterize fear-related processes in humans. 42

Attempting to down-regulate the emotional overreactions toward threat-predictive and new stimuli is one of the main routes for developing effective treatments for anxiety and post-traumatic disorders. Common approaches such as pharmacological treatments and cognitive-behavioral therapy (CBT) have demonstrated partial efficacy⁵, and recent evidence suggests that the functional outcome of behavioral methods may depend on the extent to which the prefrontal cortex is recruited during these processes⁶. Hence, new intervention strategies influencing the prefrontal dynamics would represent an important advance in the field⁷.

Previous studies adopted transcranial direct current stimulation (tDCS) or transcranial electrical stimulation (tES) to disrupt the consolidation of these memories^{8–10}, potentiate extinction processes^{11,12}, and narrow threat generalization patterns¹³, leading to contradictory results. According to one work⁸, cathodal stimulation over the dorsolateral prefrontal cortex (dlPFC) disrupted threat memory consolidation, with no enhancing effect of anodal stimulation. In contrast, other studies found an increase in implicit responses with anodal stimulation⁹ and no effect of cathodal stimulation¹⁰ over the same site. Moreover, one study employing anodal stimulation over the dlPFC¹² revealed an improvement in extinction learning but no delayed effects on the recall of the extinction memory. A further investigation¹¹ reported that low-frequency alternating-current (AC) stimulation of the medial prefrontal cortex (mPFC) augmented the defensive responses, whereas direct-current (DC) stimulation widened threat generalization profiles.

An alternative neurostimulation approach is repetitive transcranial magnetic stimulation 61 (rTMS), which ensures greater focality^{14,15}. Some rTMS studies targeted the mPFC¹⁶ and the 62 posterior PFC¹⁷ to obtain a successful enhancement of extinction learning, while others^{18,19} targeted 63 64 the dIPFC to disrupt threat-memory reconsolidation. Indeed, most rTMS-based research targeting the PFC has pursued an improvement of fear extinction, which may be followed by a return of fear 65 with a change of context (i.e. renewal)²⁰ where prevention of relapse over time is the main 66 67 challenge for therapies dedicated to post-traumatic and anxiety disorders. No previous studies reported significant effects in down-modulating the defensive responses triggered by a learned 68 69 threatening stimulus without adopting fear extinction.

So far, human brain stimulation studies have been mainly focused on the dorsolateral region 70 of the PFC⁷, partly because other prefrontal areas involved in the top-down regulation of subcortical 71 72 threat-detection systems –such as the ventromedial PFC (vmPFC), are too deep to be reached with TMS¹⁷. However, within the PFC, a brain structure that is emerging to be engaged in downstream 73 emotional regulation is the anterior prefrontal cortex (aPFC), also known as the frontopolar cortex 74 or rostral frontal cortex. The aPFC encompasses the most anterior portion of the prefrontal cortex 75 (Brodmann area 10, BA 10)²¹ and extends over a wider cortical space in humans than in other 76 species²². Even if it has not been included in fear network models so far, many studies^{23–25} 77 highlighted its role in emotional down-regulation. Anatomical projections have been found between 78 the lateral^{25,26} and the medial aPFC²⁷ and the amygdala, and functional connectivity has been 79 detected between the aPFC and the vmPFC during fear down-regulation²⁸. Notably, hypoactivation, 80

reduced connectivity, and altered thickness of aPFC were reported in PTSD patients^{29–32}, whereas a longitudinal study³³ showed that strong activation of the aPFC resulted in a higher resilience against PTSD onset. Accordingly, enhanced aPFC activity and potentiated aPFC-vmPFC connectivity were detected after an effective therapy in PTSD patients³⁴. Crucially, the aPFC is a surface area easily accessible with rTMS. However, to our knowledge, no study has been so far conducted to explore the effects of aPFC stimulation on the expression of a threat memory without extinction learning in humans.

In our current study, we posited that applying rTMS to the aPFC could influence implicit defensive responses to a learned threat-predictive stimulus and/or the conscious recognition of it. Subsequently, we explored additional hypotheses. The second hypothesis centered on the potential extension of rTMS dampening effects to new stimuli, thereby reducing threat generalization. The third hypothesis focused on the enduring persistence of rTMS effects on defensive responses over time. The final hypothesis proposed that the dampening effects achieved by stimulating the aPFC might surpass those observed when targeting the dorsolateral PFC. 95 **Results**

96 aPFC-focused rTMS effects on implicit defensive reactions toward threat-predictive and new 97 cues

98 To explore the effects of an aPFC-centered rTMS on the implicit responses to a learned threat, we99 designed a three-session experiment starting with a threat learning session followed by an implicit100 retention test and a follow-up implicit re-test (Figure 1).

During the learning session, participants learned to associate an auditory cue (conditioned 101 stimulus, CS, 800Hz) with a mild electric stimulation (unconditioned stimulus, US, individually 102 calibrated intensity) in a given environment (context A). We adopted a single-cue learning 103 paradigm because it more ecologically reflects real-life traumatic experiences^{35–39}. To validate the 104 between-groups homogeneity in the painful stimuli perception, we compared the post-conditioning 105 US ratings, and we observed no significant differences between groups (Student's unpaired t test, 106 $t_{(58)} = 0.799$, P = 0.428, $\eta_p^2 = 0.011$) (Table 1). We also did not observe significant differences 107 between groups in SCRs to the CS during the preconditioning phase ($t_{(58)} = 0.418$, P = 0.677, $\eta_p^2 =$ 108 0.003), to the CS during the conditioning phase (2×15 mixed ANOVA; main effect of group: $F_{(1,52)}$ 109 = 2.367, P = 0.130, $\eta_p^2 = 0.044$; main effect of trial: $F_{(8,762,455,600)} = 13.366$, P < 0.001, $\eta_p^2 = 0.204$; 110 group × trial interaction: $F_{(8,762,455,600)} = 1.619$, P = 0.109, $\eta_p^2 = 0.030$; Student's unpaired *t* test on 111 the averaged response, $t_{(58)} = 1.290$, P = 0.202, $\eta_p^2 = 0.028$, nor to the US during the conditioning 112 phase ($t_{(58)} = 1.011$, P = 0.316, $\eta_p^2 = 0.017$) (Figure 2-figure supplement 1). 113

One week later, we tested the implicit memory of the learned association in control shamstimulated subjects and in those who received rTMS over the aPFC shortly before the memory test. To locate this brain region, which corresponds to the BA 10^{40} , we positioned the coil over the frontopolar midline electrode (Fpz) adopting the international 10–20 electroencephalogram (EEG) coordinate system⁴¹ since previous rTMS studies^{16,42,43} ensured this placement reached the aPFC. An offline 10-min session of 1Hz-rTMS targeting this neural site (aPFC, n = 30) was applied immediately before memory retrieval (Figure 2A). Control subjects underwent a 10-min sham stimulation procedure over the same cortical area (sham, n = 30).

Memory retention was tested in a different environment from that where the learning had 122 occurred (context B) to avoid any contextual influence on retrieval^{4,44–47}. Indeed, the context shift 123 for this session mirrors a real-life treatment setting –which unlikely takes place in the threatening 124 location. To test implicit threat memory, we performed an implicit recognition task in which 125 subjects were exposed to the CS while being recorded in their evoked autonomic reactions (i.e., 126 electrodermal skin conductance responses, SCRs). No US shocks were delivered during this phase. 127 Besides the CS, participants were presented with two novel but perceptually similar tones (NS₁, 128 1000Hz; NS₂, 600Hz) to study threat generalization. Auditory frequencies of NSs were selected to 129 obtain a slowly decaying gradient of defensive tunings^{4,48,49}. To test the effects of rTMS on memory 130 retention, we compared the between-group differences as well as the within-group differences from 131 the acquisition phase to the testing phase through a 2×2 mixed ANOVA. This analysis yielded a 132 not significant main effect of group ($F_{(1,58)} = 2.015$, P = 0.161, $\eta_p^2 = 0.034$), a not significant main 133 effect of phase (F_(1,58) = 0.053, P = 0.818, $\eta_p^2 = 0.001$) and a significant group × phase interaction 134 $(F_{(1.58)} = 13.445, P = 0.001, \eta_p^2 = 0.188)$. Simple main effects analysis revealed no significantly 135 different mean CS-evoked SCRs between groups during the conditioning phase (P = 0.506; 136 137 Bonferroni corrected). On the contrary, during the test phase subjects that received rTMS over the aPFC exhibited weakened CS-related SCRs than those observed in the sham group (P = 0.006; 138 Bonferroni corrected). Moreover, the aPFC group showed reduced autonomic responses to the CS 139 from conditioning to test (P = 0.008; Bonferroni corrected) whereas the sham group displayed 140 increased mean SCRs to the CS from conditioning to test (P = 0.018; Bonferroni corrected) (Figure 141 2B,C). This data indicates that the rTMS procedure affected SCRs triggered by memory retrieval 142 performed shortly after rTMS. To the best of our knowledge, this is the first evidence that brain 143 stimulation may promptly attenuate implicit defensive reactions during memory retrieval. 144

In the test session, we also analyzed threat generalization to the NSs through a 2 × 3 mixed ANOVA, which showed a significant main effect of group ($F_{(1,58)} = 5.310$, P = 0.025, $\eta_p^2 = 0.084$), a not significant main effect of tone ($F_{(2,116)} = 0.690$, P = 0.504, $\eta_p^2 = 0.012$) and a not significant group × tone interaction ($F_{(2,116)} = 1.301$, P = 0.276, $\eta_p^2 = 0.022$), revealing that the aPFC group displayed overall attenuated responses to tones relative to the sham condition (Figure 2D,E).

We next sought to disambiguate whether the rTMS effects were due to a general downregulation of electrodermal responsivity, or whether they specifically targeted the threat memory. To this end, subjects were presented with an unconditioned threatening stimulus consisting of a female scream sample (unconditioned stimulus 2, US₂) while being recorded in their SCRs. No significant differences emerged between conditions ($t_{(58)} = 0.334$, P = 0.739, $\eta_p^2 = 0.002$), indicating that the rTMS did not cause an overall inhibition of electrodermal reactivity (Figure 2F).

To test whether and to what extent rTMS-related outcomes endured beyond the after-effect window and persisted over a long-term period, we planned a follow-up session. One week after the threat memory retrieval test, all participants returned to the conditioning room (context A) and underwent a re-testing phase, identical to the testing one except for the absence of rTMS administration. This phase also allowed us to test a possible renewal effect²⁰ since subjects were reexposed to the original threatening environment.

Concerning the implicit responses to the CS, a 2×2 mixed ANOVA showed a not 162 significant main effect of group ($F_{(1,58)} = 1.952$, P = 0.168, $\eta_p^2 = 0.033$), a significant main effect of 163 phase (F_(1.58) = 7.690, P = 0.007, $\eta_p^2 = 0.117$) and a significant group × phase interaction (F_(1.58) = 164 9.966, P = 0.003, $\eta_p^2 = 0.147$). Simple main effects analysis revealed that participants of the aPFC 165 group persisted in displaying weaker SCRs than those observed in the sham group (P = 0.006; 166 Bonferroni corrected). Moreover, the aPFC group persisted in showing a decrease of defensive 167 reactions to the CS from conditioning to follow-up (P < 0.001; Bonferroni corrected), while the 168 sham group did not display significantly different SCRs in the two phases (P = 0.787; Bonferroni 169 corrected) (Figure 2G,H). 170

These findings support an enduring effect of the aPFC-rTMS in attenuating the long-term implicit defensive responses to the learned threat-predictive cue, even with the re-exposition to the environment where threat learning had occurred. We next analyzed the autonomous response patterns to the female scream sample (unconditioned stimulus 2, US₂) and again we found that reactions did not differ between groups ($t_{(58)} = 0.057$, P = 0.955, $\eta_p^2 < 0.001$) (Figure 2I). Thus, the persistent effect was expressed notwithstanding an unaffected electrodermal overall reactivity.

An important aspect to consider is that rTMS application over the forehead can be 177 subjectively perceived as unpleasant. We, therefore, investigated whether an rTMS-related 178 discomfort before memory retrieval might have provoked habituation to unpleasant stimulations, 179 180 leading to a reduction in SCR levels during CS presentations. We repeated the entire experiment in one further group (ctrl discomfort, n = 10) by replacing the rTMS procedure with a 10-min 181 182 discomfort-inducing procedure over the same site of the forehead to mimic the rTMS-evoked 183 unpleasant sensations in the absence of neural stimulation effects. This group showed no significantly different CS-evoked SCR levels to those of the sham group during the test session as 184 well as during the follow-up session (Figure 2-figure supplement 2). Thus, the discomfort 185 experienced during the rTMS procedure did not contribute to the reduction of electrodermal 186 responses observed in the aPFC-stimulated group. 187

188

aPFC-focused rTMS effects on the explicit memory recognition and perceptual discrimination 189 We then investigated the effect of rTMS over the aPFC on the retention of explicit-declarative 190 threat memories. A further group of subjects that received the identical 1Hz-rTMS procedure over 191 the aPFC (aPFC-E, n = 21) and a further control group (sham-E, n = 21) underwent an explicit two-192 alternative forced-choice (2AFC) recognition task, in which they were presented with a random 193 sequence of tone pairs, each composed of the CS and one of the two NSs. Subjects were asked to 194 consciously identify which stimulus of each pair was the one previously paired with the US (i.e., the 195 CS), and to provide a subjective confidence level for each choice using a scale ranging from 0 196

(completely unsure) to 10 (completely sure)^{4,50}. Both groups reported not significantly different post-conditioning US ratings ($t_{(40)} = 0.339$, P = 0.737, $\eta_p^2 = 0.003$) and successfully identified the CS amongst the NSs with an accuracy level above the 50% chance level (aPFC-E: $t_{(20)} = 9.226$, P < 0.001, $\eta_p^2 = 0.810$; sham-E: $t_{(20)} = 14.240$, P < 0.001, $\eta_p^2 = 0.910$). A between-groups comparison ($t_{(40)} = 1.114$, P = 0.272, $\eta_p^2 = 0.030$) showed no differences in the explicit recognition accuracy (Figure 3A). The two groups were not differently confident when making their choices ($t_{(40)} = 0.842$, P = 0.405, $\eta_p^2 = 0.017$) (Figure 3B), thereby supporting the lack of rTMS-related effects.

Next, since a previous study¹³ targeting the vmPFC modulated perceptual discrimination 204 processes, we implemented a 2AFC perceptual task in which we investigated the ability of 205 206 participants to sensory discriminate between the CS and the two NSs by collecting binary 'same or different' judgments as well as confidence ratings. The perceptual discrimination test yielded no 207 significant between-groups differences in accuracy ($t_{(40)} = 1.362$, P = 0.181, $\eta_p^2 = 0.044$) as well as 208 confidence levels ($t_{(40)} = 0.917$, P = 0.365, $\eta_p^2 = 0.021$). Indeed, both groups discriminated the CS 209 from the NSs with high precision (aPFC-E: 0.980 ± 0.015 SEM; sham-E: 1.000 ± 0.000 SEM) and 210 with no different confidence levels (aPFC-E: 9.409 ± 0.153 SEM; sham-E: 9.586 ± 0.117 SEM), 211 thereby showing no rTMS effects on sensory abilities. 212

These data suggest that the pre-retrieval rTMS procedure over the aPFC did not affect the explicit recognition nor the perceptual discrimination of a learned threat.

During the follow-up session, explicit recognition patterns demonstrated an over-chance accuracy level for each group (aPFC-E: $t_{(20)} = 13.780$, P < 0.001, $\eta_p^2 = 0.905$; sham-E: $t_{(20)} = 7.162$, P < 0.001, $\eta_p^2 = 0.720$). Again, here there were no between-group differences ($t_{(40)} = 1.024$, P = 0.312, $\eta_p^2 = 0.026$) since both groups achieved a high recognition accuracy (Figure 3C). Groups did also not report different confidence levels ($t_{(40)} = 0.084$, P = 0.934, $\eta_p^2 < 0.001$) (Figure 3D).

As in the case of the previous session, we did not observe significant between-group differences in the perceptual discrimination ($t_{(40)} = 1.000$, P = 0.323, $\eta_p^2 = 0.024$) and the respective confidence ratings ($t_{(40)} = 0.149$, P = 0.882, $\eta_p^2 < 0.001$). Indeed, the discrimination accuracy (aPFC-E: 1.000 ± 0.000 SEM; sham-E: 0.993 ± 0.007 SEM) and the self-assessed confidence
(aPFC-E: 9.598 ± 0.147 SEM; sham-E: 9.633 ± 0.182 SEM) were high in each condition.

225

Topographical selectivity of rTMS effects on implicit defensive responses to threat-predictive and new cues

To ascertain the topographical selectivity, in one further condition (OC, n = 30) we applied the rTMS over the left occipital cortex as an active control site (Figure 4A) and we contrasted its implicit reactions with those of the group stimulated over the aPFC.

No differences emerged between the two conditions in terms of post-conditioning US 231 ratings $(t_{(58)} = 0.000, P = 1.000, \eta_p^2 = 0.000)$ (Table 1), SCR responses to the CS during the 232 preconditioning phase ($t_{(58)} = 1.037$, P = 0.304, $\eta_p^2 = 0.018$), to the CS during the conditioning phase 233 $(2 \times 15 \text{ mixed ANOVA}; \text{ main effect of group: } F_{(1.54)} = 0.124, P = 0.726, \eta_p^2 = 0.002; \text{ main effect of}$ 234 trial: $F_{(9,368,505,856)} = 13.341$, P < 0.001, $\eta_p^2 = 0.198$; group × trial interaction: $F_{(9,368,505,856)} = 0.994$, P 235 = 0.445, $\eta_p^2 = 0.018$; Student's unpaired t test on the averaged response, $t_{(58)} = 0.162$, P = 0.872, η_p^2 236 < 0.001), and to the US during the conditioning phase ($t_{(58)} = 1.210$, P = 0.231, $\eta_p^2 = 0.025$) (Figure 237 4-figure supplement 1). 238

Next, we analyzed implicit reactions toward the CS in both conditioning and test sessions. A 239 2×2 mixed ANOVA revealed a not significant group main effect (F_(1.58) = 2.952, P = 0.091, η_p^2 = 240 0.048), a not significant phase main effect ($F_{(1,58)} = 2.027$, P = 0.160, $\eta_p^2 = 0.034$), and a significant 241 group × phase interaction ($F_{(1.58)} = 4.705$, P = 0.034, $\eta_p^2 = 0.075$). CS-related SCRs did not differ 242 between groups during conditioning (P = 0.798; Bonferroni corrected) but, during the test, the aPFC 243 group exhibited weaker defensive responses than the OC group (P = 0.019; Bonferroni corrected). 244 Unlike the aPFC group, whose implicit reactions to the CS diminished from conditioning to test (P 245 = 0.014; Bonferroni corrected), the OC group's responses did not differ in the two phases (P =246 0.600; Bonferroni corrected) (Figure 4B,C). 247

No significant between-group differences were observed in implicit responses to new tones 248 $(2 \times 3 \text{ mixed ANOVA}; \text{ main effect of group: } F_{(1.58)} = 2.775, P = 0.101, \eta_p^2 = 0.046; \text{ main effect of}$ 249 tone: $F_{(2,116)} = 5.857$, P = 0.004, $\eta_p^2 = 0.092$; group × tone interaction: $F_{(2,116)} = 3.739$, P = 0.027, η_p^2 250 = 0.061). While the CS triggered weaker reactions in the aPFC group (P = 0.019; Bonferroni 251 corrected), both the NS₁ (P = 0.203; Bonferroni corrected) and the NS₂ (P = 0.323; Bonferroni 252 corrected) elicited not significantly different responses in the two conditions. These findings 253 254 underscored the selectivity of divergent rTMS effects in the aPFC and OC groups specifically for the CS. Fear tuning analysis of the aPFC group's implicit reactions unveiled no differences in SCR 255 amplitudes elicited by the CS and the NS₁ (P = 0.378; Bonferroni corrected), by the CS and the NS₂ 256 (P = 1.000; Bonferroni corrected), and by the NSs (P = 0.552; Bonferroni corrected). In the case of 257 the OC group, implicit reactions were not different for the CS and the NS₁ (P = 0.876; Bonferroni 258 corrected) but the NS₂ evoked lower SCRs than the CS (P < 0.001; Bonferroni corrected) and the 259 NS_1 (P = 0.041; Bonferroni corrected) (Figure 4D,E). Furthermore, no significant group differences 260 were detected in SCRs elicited by US₂ during the test session ($t_{(58)} = 0.175$, P = 0.862, $\eta_p^2 < 0.001$) 261 (Figure 4F). 262

The distinctive pattern toward the learned threatening cue persisted during the follow-up 263 session (2 × 2 mixed ANOVA; main effect of group: $F_{(1.58)} = 2.141$, P = 0.149, $\eta_p^2 = 0.036$; main 264 effect of phase: $F_{(1.58)} = 26.023$, P < 0.001, $\eta_p^2 = 0.310$; group × phase interaction: $F_{(1.58)} = 3.167$, P 265 = 0.080, η_p^2 = 0.052). The aPFC group continued to react more dimly to the CS compared to the OC 266 group (P = 0.026; Bonferroni corrected). Both the aPFC (P < 0.001; Bonferroni corrected) and the 267 OC (P = 0.022; Bonferroni corrected) groups showed decreased responses relative to conditioning 268 (Figure 4G,H). Conversely, no significant differences were observed in SCRs evoked by US₂ 269 during the follow-up session ($t_{(58)} = 0.574$, P = 0.568, $\eta_p^2 = 0.006$) (Figure 4I). 270

271

272 Comparison between the effects of rTMS administered over the anterior versus the
 273 dorsolateral prefrontal cortex

Next, we asked whether the findings we obtained by targeting the aPFC were finely specific for this 274 275 site or, alternatively, they overlapped with those observed by targeting other prefrontal sub-regions. For this purpose, in one further group (dlPFC, n = 30) we applied the same rTMS procedure over 276 the left dorsolateral PFC (Figure 5A) and we then compared the implicit patterns of this group with 277 those displayed by the aPFC condition. We selected the left dlPFC since previous studies^{e.g. 17} 278 targeted the left hemisphere for testing the rTMS effects on the PFC, and some evidence^{see 7} 279 suggested that inhibitory tDCS and rTMS over the left dlPFC may disrupt threat memory 280 consolidation. 281

We found no significant differences between the two conditions in the post-conditioning US 282 ratings $(t_{(58)} = 0.908, P = 0.368, \eta_p^2 = 0.014)$ (Table 1), in SCRs to the CS during the 283 preconditioning phase ($t_{(58)} = 0.967$, P = 0.337, $\eta_p^2 = 0.016$), to the CS during the conditioning phase 284 $(2 \times 15 \text{ mixed ANOVA}; \text{ main effect of group: } F_{(1.51)} = 0.026, P = 0.873, \eta_p^2 = 0.001; \text{ main effect of}$ 285 trial: $F_{(8,026,409,333)} = 12.135$, P < 0.001, $\eta_p^2 = 0.192$; group × trial interaction: $F_{(8,026,409,333)} = 1.042$, P 286 = 0.403, $\eta_p^2 = 0.020$; Student's unpaired t test on the averaged response, $t_{(58)} = 0.378$, P = 0.707, η_p^2 287 = 0.002), and to the US during the conditioning phase ($t_{(58)} = 1.752$, P = 0.085, $\eta_p^2 = 0.050$) (Figure 288 5-figure supplement 1). 289

Then we compared the implicit reactions toward the CS during conditioning and test 290 sessions. A 2 \times 2 mixed ANOVA indicated a not significant main effect of group (F_(1.58) = 1.874, P 291 = 0.176, $\eta_p^2 = 0.031$), a not significant main effect of phase (F_(1.58) = 0.122, P = 0.729, $\eta_p^2 = 0.002$), 292 and a significant group × phase interaction ($F_{(1.58)} = 10.810$, P = 0.002, $\eta_p^2 = 0.157$). CS-evoked 293 SCRs did not differ between the two groups during conditioning (P = 0.647; Bonferroni corrected) 294 while during the test we found weaker defensive responses in the aPFC group relative to the dlPFC 295 group (P = 0.009; Bonferroni corrected). At odds with the aPFC group whose implicit reactions to 296 the CS were diminished from conditioning to test (P = 0.013; Bonferroni corrected), the dlPFC 297 group increasingly responded during the test relative to conditioning (P = 0.042; Bonferroni 298

corrected) (Figure 5B,C). This incremental trend is in line with a previous study that delivered a
 1Hz-rTMS protocol over the left dlPFC¹⁸.

We found no between-groups differences in the implicit responses to the new tones (2×3 301 mixed ANOVA; main effect of group: $F_{(1,58)} = 3.967$, P = 0.051, $\eta_p^2 = 0.064$; main effect of tone: 302 $F_{(2,116)} = 2.819, P = 0.064, \eta_p^2 = 0.046; \text{ group} \times \text{tone interaction: } F_{(2,116)} = 3.286, P = 0.041, \eta_p^2 = 0.0$ 303 0.054) since to both the NS₁ (P = 0.188; Bonferroni corrected) and the NS₂ (P = 0.110; Bonferroni 304 corrected) were not significantly different. These data showed that the divergent rTMS effects in the 305 aPFC and the dlPFC groups were selective for the CS. Fear tuning analysis of the dlPFC group's 306 implicit reactions revealed no different SCR amplitudes elicited by the CS and the NS₁ (P = 0.158; 307 Bonferroni corrected) and by the NSs (P = 0.721; Bonferroni corrected), but the NS₂ evoked lower 308 SCRs than the CS (P = 0.014; Bonferroni corrected) (Figure 5D,E). We also detected no significant 309 differences between groups in the SCRs elicited by the US₂ during the test session ($t_{(58)} = 1.762$, P =310 0.083, $\eta_p^2 = 0.051$) (Figure 5F). 311

The different pattern toward the learned threatening cue was replicated during the follow-up 312 session (2 × 2 mixed ANOVA; main effect of group: $F_{(1,58)} = 3.751$, P = 0.058, $\eta_p^2 = 0.061$; main 313 effect of phase: $F_{(1,58)} = 3.114$, P = 0.083, $\eta_p^2 = 0.051$; group × phase interaction: $F_{(1,58)} = 15.248$, P 314 < 0.001, $\eta_p^2 = 0.208$) since the aPFC group persisted in more dimly reacting to the CS with respect 315 to the dlPFC group (P = 0.001; Bonferroni corrected), and the aPFC group endured in displaying 316 attenuated responses relative to conditioning (P < 0.001; Bonferroni corrected) while the dlPFC 317 group did not (P = 0.136; Bonferroni corrected) (Figure 5G,H). No significant differences were 318 instead observed in SCRs evoked by the US₂ during the follow-up session ($t_{(58)} = 1.927$, P = 0.059, 319 $\eta_{\rm p}^2 = 0.060$) (Figure 5I). 320

These findings demonstrated that rTMS over the left dorsolateral PFC did not diminish implicit defensive reactions in the absence of an extinction paradigm, as in other previous studies^{16–} Meanwhile, rTMS targeting the aPFC proved to be effective in achieving this outcome.

324 Discussion

In this study, we found that implicit reactions to both learned and novel stimuli were significantly down-regulated following a 1Hz-rTMS procedure over the aPFC.

So far, most rTMS studies targeting the prefrontal cortex have been conducted to enhance 327 fear extinction processes. A study¹⁶ administering one session of 10Hz-rTMS over the mPFC 328 observed enhancement of extinction learning. These behavioral results were mirrored by the 329 functional near-infrared spectroscopy (fNIRS) findings, which revealed increased mPFC activity in 330 the stimulated group relative to the sham group¹⁶. Subsequently, Raij *et al.* (2018) delivered brief 331 20Hz-rTMS trains over the left posterior PFC –a region that showed robust functional connectivity 332 333 with the vmPFC- during extinction learning and found a reduction of defensive responses during 334 extinction recall.

Our study differs from the previous ones because we tested rTMS effects over the medial 335 anterior prefrontal cortex (medial BA 10), and we did not include extinction training before 336 retrieval. We observed a significant decrease in defensive reactions shortly after rTMS, and this 337 effect was maintained until the follow-up session. Thus, we identified a previously unexplored 338 prefrontal region, the modulation of which can efficiently and durably inhibit implicit reactions to 339 learned threats. These dampening effects may be due to the fact that rTMS over the aPFC have 340 directly modulated the defensive responses activated by the implicit threat memory trace. 341 Alternatively, the rTMS procedure over the aPFC may have inhibited the recall of the CS-US 342 association, preventing the defensive responses from being activated by the CS. This possibility 343 would be in line with a large body of literature on humans^{see 51} which demonstrates the importance 344 of the medial PFC for value-based processing. 345

Autonomic reactions to the new tones in the aPFC group relative to the sham control group did not support the conclusion that rTMS targeted threat generalization, leaving open the question of the specificity of rTMS effects. However, the lack of between-group differences in the autonomic responses to the US_2 seems to suggest that the observed effect may be memory-related and not due to a general dampening of autonomic reactivity. Interestingly, defensive responses toward the NSs were decreased following the stimulation of the left occipital cortex (OC group, BA18/19). This effect might be explained by the fact that anatomical and functional reciprocal projections between the medial BA10 and visual association cortices (including BA17/18/19) have been traced via the fronto-occipital fasciculus (FOF) of the human brain^{52–54 but see 27}.

Regarding the persistence of inhibitory effects during the follow-up session, different factors 355 356 may have contributed to this result. Firstly, the inhibition of SCR responses induced by rTMS during the mnemonic retention test could have persistently reduced such conditioned responses 357 even at a distance from the treatment. Moreover, the inhibition of these responses during the test 358 359 might have boosted the extinction of these responses, contributing to keeping them low over time. On this possibility, it should be pointed out that one core knowledge about extinction is that under 360 certain circumstances -such as a simple passage of time (i.e., spontaneous recovery) or a change in 361 362 surrounding context (i.e., renewal)- extinguished reactions triggered by the CS may reoccur, giving rise to the phenomenon known as return of fear^{20,55,56}. To test potential renewal phenomena, which 363 have not been investigated in the aforementioned studies^{16,17}, we opted for a context-shift amongst 364 the learning (context A), the test (context B), and the follow-up phase (context A), and we found 365 down-regulated defensive reactions in both the test and the follow-up phases. These data 366 367 demonstrated that the aPFC-rTMS protocol long-term reduced threat memory expression in a different context as well as in the context in which the threatening experience had occurred, thus 368 preventing the return of fear. Finally, we cannot exclude that the rTMS applied immediately before 369 370 the mnemonic retention test interfered with the reconsolidation process that is known to occur after this test¹⁹, resulting in a persistent impairment in the retention of this mnemonic trace. 371

To potentiate the neural activity of the PFC, both the aforementioned studies^{16,17} adopted high-frequency rTMS protocols –which are conventionally considered excitatory of proximal brain activity⁵⁷. In our study, we adopted a low-frequency rTMS protocol –which is conventionally considered inhibitory⁵⁷. Recent evidence, however, challenged this common frequency-dependent rule⁵⁸. Resting-state functional magnetic resonance imaging (fMRI) studies demonstrated that 1HzrTMS protocols may also induce downstream distal effects and enhance functional connectivity amongst the brain regions located underneath the coil and remote brain areas of the stimulated neural network⁵⁹. Additionally, some studies^{60,61} reported that 1Hz-rTMS procedures delivered over the PFC may paradoxically increase regional cerebral blood flow (rCBF).

The dorsolateral PFC is another prefrontal region that is assumed to be critically involved in 381 threat learning^{62,63} and the down-regulation of the cortico-meso-limbic network⁶⁴. One 382 investigation¹⁸ probed the effects of a 1Hz-rTMS over the dlPFC after memory reactivation to 383 disrupt threat-memory reconsolidation. Stimulated groups failed to discriminate between 384 threatening and safe stimuli, with an increase in autonomic responses to these last ones. A more 385 recent study¹⁹ adopted the continuous theta-burst stimulation (cTBS) over the right dlPFC during 386 the reconsolidation window and successfully decreased the defensive responses for threat 387 388 memories. In our study, we found an immediate and long-term reduction of defensive responses to the CS only in subjects that were stimulated over the aPFC, while reactions to the NSs were 389 390 decreased in both conditions. This evidence suggests that targeting the aPFC might represent a more promising approach for therapeutic applications. The lack of any down-regulation of CS-evoked 391 reactions that we found in the dIPFC group, at odds with previous studies targeting the same 392 cortical area^{18,19}, might be due either to the fact that we did not adopt an extinction paradigm, or to 393 the different brain stimulation approach (rTMS vs cTBS). 394

The neural mechanisms by which rTMS over the aPFC decreases threat-conditioned responses can be manifold. Fear memories are formed and retrieved by an intricate neural network encompassing the amygdala⁶⁵, the cerebellum^{66–68}, and sensory cortices^{46,69–77}. Indeed, previous evidence showed both structural connections between the aPFC and the amygdala^{25–27} and a connectivity pathway of downstream modulation from the aPFC to the vmPFC³⁴. This projection is activated during fear regulation²⁸, possibly supporting the vmPFC in top-down modulating the amygdala⁷⁸. Through the direct or indirect connections of the aPFC with these areas, it might be that the effects of focal manipulations of aPFC activity reflect complex and dynamic changes in theoverall neural network state and/or influence the activity of some of these areas.

Although previous studies enlightened the role of the medial BA10 and BA10-posterior 404 hippocampus functional connectivity in episodic memory retrieval^{see 79}, we did not detect any 405 rTMS-driven effect on explicit recognition memory. The observed divergence between autonomous 406 and declarative patterns might have been due to a selective rTMS action upon the neural system 407 supporting implicit threat processing, which has been widely dissociated from the neural system 408 underlying explicit memory processes^{80–82}. Critically, an rTMS procedure that shapes implicit 409 overreactions to learned threats without affecting conscious knowledge of danger might represent a 410 411 strategic advantage for therapeutic applications.

Since prevention of relapse is the main challenge for therapies dedicated to post-traumatic and anxiety disorders, our findings may represent an advance in this direction by providing a potential strategy to deactivate emotional overreactions and, most of all, to prevent the return of fear. Future research perspectives might consist of exploring this rTMS application over the aPFC in clinical populations displaying high levels of anxiety or suffering from anxiety disorders and PTSDs.

418 Materials and Methods

419 **Participants**

All participants (n = 183) were healthy volunteers (mean age: 23.86 ± 2.90, 74 males and 109 420 females) with no history of psychiatric disorders, neurological illnesses, cardiovascular diseases, 421 illegal drug use, musical training, or any other exclusion criteria for rTMS administration⁸³. During 422 the pre-experimental screening phase, each volunteer was also administered the *State-Trait Anxiety* 423 Inventory Form $Y^{84,85}$, and those who showed a score >80 in the sum of the two subscales (State + 424 Trait anxiety) were not included in the sample (see Table 1 for all groups' mean State-Trait Anxiety 425 Inventory scores). Participants were then randomly assigned to each experimental condition, based 426 427 on sex and age (see Table 1 for all groups' mean age and sex distribution). We discarded eleven participants because of a complete absence of skin conductance responses (SCRs) during the test 428 session, leaving a total of 172 participants. Each participant provided written informed consent after 429 430 receiving a complete description of the experimental procedures. All experimental procedures were performed in accordance with the ethical standards of the Declaration of Helsinki and were 431 approved by the Bioethics Committee of the University of Turin (protocols N. 19961 and N. 432 161427). 433

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435 Auditory stimuli

Auditory stimuli were pure sine wave tones with oscillation frequencies of 800Hz (CS), 1000Hz
(NS₁), and 600Hz (NS₂), lasting 6s with onset/offset ramps of 5ms. Tones were digitally generated
using Audacity 2.1.2 software (Audacity® freeware). The unconditioned threatening stimulus (US₂)
consisted of a woman scream sample lasting 4s. All auditory stimuli were binaurally delivered
through headphone speakers (Direct Sound EX29) at 50 dB intensity. All experimental scenarios
were controlled by Presentation® 21.1 software (NeuroBehavioral Systems, Berkeley, CA).

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445 **Preconditioning**

This phase consisted of the presentation of 4 trials of the CS (800Hz) with an inter-trial-interval (ITI) randomly ranging between 21s and 27s. SCRs were recorded during this phase to provide a baseline response pattern to the 800Hz tone for each participant. At the end of this phase, participants were asked to confirm whether the tones were easily audible but not too loud or annoying.

451

452 Unconditioned stimulus calibration procedure

453 Before starting with the calibration procedure, systolic and diastolic blood pressure was measured to prevent possible hypo-arousal reactions caused by basal hypotension. The unconditioned stimulus 454 (US) consisted of a mild electrical shock (train pulse at 50Hz lasting 200ms, with a single pulse 455 456 duration of 1000µs) generated with a direct current stimulator (DS7A Constant Current Stimulator, Digitimer). Impulses were delivered through a bar stimulating electrode connected by a Velcro strap 457 458 on the upper surface of the dominant hand's index finger. The electrical stimulation intensity was individually calibrated through a staircase procedure^{4,50,86}, starting with a low current near the 459 perceptible tactile threshold (~0.5 mA). Participants were asked to rate the painfulness of each train 460 pulse on a scale ranging from 0 (not painful at all), 1 (pain threshold) to 10 (highly painful if 461 protracted in time). At the end of the procedure, the US amplitude was then set at the current level 462 (mA) corresponding to the mean rating of '7' on the subjective analog scale. 463

464

465 **Conditioning**

After a 1-min resting period, participants underwent a single-cue auditory threat conditioning, which consisted of the presentation of 15 trials of the conditioned stimulus (CS, 800Hz), with an ITI randomly ranging between 21s and 27s. The CS co-terminated with the US 12 times (80% reinforcement rate). Subjects were not informed about any possible CS-US contingency. To validate the threat learning experience, immediately following this phase subjects rated the painfulness of
the US using the same analog scale as in the preconditioning calibration procedure (see Table 1 for
all groups' US current intensity and US analog ratings).

473

474 Transcranial Magnetic Stimulation

Transcranial Magnetic Stimulation was performed with a Magstim Rapid² Stimulator (Magstim 475 Co., Whitland, Dyfed, UK). A 70-mm figure-of-eight coil was positioned over the subject's M1 476 cortical area at the optimum scalp position to elicit a contraction of the contralateral abductor 477 pollicis brevis muscle (APB). Resting motor threshold (rMT) was defined as the minimum 478 479 stimulation intensity that induced a visible finger movement in at least 5 out of 10 single pulses over the right-hand area of the left primary motor cortex^{16,87}. After having determined each 480 individual's rMT, we applied a single train of 1Hz-rTMS^{88,89} for a total duration of 10 min (600 481 482 pulses) to the target area. The rTMS intensity was set at 80% of the rMT for subjects whose rMT was \leq 50% of the machine's maximum deliverable power (e.g., the intensity corresponded to 40%) 483 484 of the maximum power when the rMT was equal to 50% of the same parameter). For subjects with an rMT > 50%, the stimulation intensity was always set to a ceiling corresponding to 40% of the 485 machine's maximum deliverable power (see Table 1 for each group's mean rMT and mean 486 stimulation intensity). During the rTMS procedure participants were seated in a comfortable 487 recliner that we adjusted to allow their upper body to be in a sloped position, thus ensuring an 488 optimal positioning of the coil. 489

To target the medial anterior portion of the prefrontal cortex (BA 10; aPFC and aPFC-E groups), the coil was centered over Fpz (10% of nasion-inion distance) according to the international 10–20 electroencephalogram (EEG) system⁴¹ (Figure 1). This placement should –with an rTMS reach of 1.5 to 2 cm beneath the scalp^{90,91}– ensure the targeting of the medial aPFC as in previous studies^{16,42,43} and avoid the targeting of the dorsomedial prefrontal cortex (dmPFC), which would have been localizable with a scalp-based heuristic approach of 25.84% nasion-inion distance⁹². In

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the case of left occipital cortex stimulation (OC group), the coil was positioned over O1 using the 496 10-20 EEG system (BA 18/19), which functionally corresponds to associative visual cortices V3, 497 V4, and $V5^{93,94}$ (Figure 1). For the stimulation of the left dorsolateral prefrontal cortex (dlPFC 498 group), the coil was placed over F3 using the 10–20 EEG system (BA 8/9)^{18,95} (Figure 1). For sham 499 stimulation (sham and sham-E groups), the coil was centered over Fpz and positioned perpendicular 500 to the scalp surface, so that no effective stimulation reached the brain during the procedure but 501 502 allowed subjects to feel a comparable coil-scalp contact and hear the same noise as in real stimulation (Figure 1). 503

All participants were blinded to their experimental condition (i.e., active or sham), and were not informed about the potential cognitive or emotional effects of the stimulation.

506

507 **Discomfort-inducing procedure**

508 The discomfort-inducing procedure mirrored the rTMS protocol and consisted of the delivery of mild electrical shocks (single 1-Hz train of 600 pulses lasting 10min, with a single pulse duration of 509 510 500µs to mimic the duration of a single TMS pulse) generated with a direct current stimulator (DS7A Constant Current Stimulator, Digitimer). Impulses were delivered through two cup-511 stimulating electrodes attached to the surface of the subject's forehead in correspondence with Fpz 512 513 according to the 10-20 EEG system. As in the case of the US calibration, the electrical stimulation intensity was individually calculated through a staircase procedure⁸⁶, starting with a low current 514 near the perceptible tactile threshold (~0.5 mA). Participants were asked to evaluate the perceived 515 discomfort of each pulse on a scale from 0 (no discomfort) to 10 (high discomfort). At the end of 516 the procedure, the shock amplitude was set at the current level (mA) corresponding to the mean 517 rating of '4' on the subjective analog scale. To quantify the habituation to the uncomfortable 518 stimulations, at the end of every minute of the 10-min procedure (i.e. every 60 pulses), subjects 519 were requested to rate the level of the present discomfort on the same scale adopted during the 520 calibration procedure. 521

522

523 Implicit recognition test

After a 1-min resting period, participants underwent this task, which consisted of the presentation of 12 auditory stimuli in a completely random sequence: $4 \times CS$, $4 \times NS_1$, $4 \times NS_2$, with an ITI whose duration randomly ranged between 21s and 27s. SCRs were recorded throughout this phase, and the stimulating electrode was kept attached to create the expectation of receiving the US⁴⁴. Differently from other paradigms^{48,96–98}, here no shocks were delivered to avoid any reacquisition effect^{4,50}.

529

530 Implicit unconditioned threatening test

This task was designed to elicit an unconditioned electrodermal response and consisted of the presentation of 4 trials of a woman scream sample lasting 4s, with an ITI randomly ranging between 21s and 27s. SCRs were recorded throughout this phase, and the stimulating electrode was kept attached.

535

536 Two-alternative forced-choice (2AFC) explicit recognition test

This procedure involves the presentation of two stimuli on each trial and the subject chooses the one that was previously encoded (i.e. the first or the second one). As in our previous works^{4,50}, a 2AFC design was preferred over a new-old paradigm, which involves one single stimulus on each trial, and the subject judges whether the stimulus has been previously encoded (old), or whether it is new. Our choice was motivated by the evidence that a 2AFC task improves recognition performance and discourages response biases such as the familiarity-based decision bias, namely the heuristic to endorse novel cues as 'old' when their familiarity is high⁹⁹.

The task consisted of the presentation of 16 tone-pairs, each composed of the CS (800Hz) and one of the two NSs (NS₁, 1000Hz or NS₂, 600Hz) in a completely random sequence: $4 \times CS$ vs NS₁, $4 \times$ NS₁ vs CS, $4 \times CS$ vs NS₂, $4 \times NS_2$ vs CS. On each trial, the two stimuli were presented with an intra-trial-interval of 1000ms. After each pair offset, an ITI randomly ranging between 21s and 27s occurred. Participants were explained that in each couple of sounds, there was a tone that they had heard on the first session (one week before or, in the case of the follow-up session, two weeks before) and a new tone. Participants were then instructed to recognize and verbally report which one (the first or the second) was the tone heard in the first session, paired with the US-shock (CS). Participants were further asked to verbally provide a confidence rating about each response, on a scale from 0 (completely unsure) to 10 (completely sure). No feedback was supplied. As in the implicit task, the stimulating electrode was kept attached, but no shock was delivered.

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556 Two-alternative forced-choice (2AFC) perceptual discrimination test

The task consisted of the presentation of 7 pairs of auditory stimuli (i.e. CS vs NS₁, NS₁ vs CS, CS vs NS₂, NS₂ vs CS, CS vs CS, NS₁ vs NS₁, NS₂ vs NS₂) with a 1000-ms intra-pair-interval in a completely random sequence (ITI randomly ranging between 21s and 27s). For each pair, subjects were asked to report whether the two tones were "the same tone or different tones", and to provide a confidence rating on an analog scale from 0 (completely unsure) to 10 (completely sure). No feedback was supplied, and the stimulating electrode was kept attached.

563

564 Psychophysiological recording and analysis

565 Event-related skin conductance responses (SCRs) were used as an implicit index of defensive responses. To record the autonomic signal, two Ag-AgCl non-polarizable electrodes filled with 566 isotonic paste were attached to the index and middle fingers of the non-dominant hand by Velcro 567 568 straps. The transducers were connected to the GSR100C module of the BIOPAC MP-150 system (BIOPAC Systems, Goleta, CA) and signals were recorded at a channel sampling rate of 1000 Hz. 569 SCR waveforms were analyzed offline using AcqKnowledge 4.1 software (BIOPAC Systems, 570 Goleta, CA), and were performed blindly to the subject's experimental condition and the 571 randomized sequence of stimuli. Each SCR was evaluated as event-related if the trough-to-peak 572 deflection occurred 1-6 s (for the CS and the NSs) or 1-4 s (for the US₂) after the stimulus onset, 573

the duration was comprised between 0.5 and 5.0 s, and the amplitude was greater than 0.02 micro siemens (μ S). Responses that did not fit these criteria were scored zero. To account for interindividual variability, these raw values were then scaled according to each participant's average unconditioned response by dividing each response by the mean unconditioned stimulus (US) response during the conditioning phase^{100,101}. Scaled SCR data were square-root transformed to normalize the distributions¹⁰².

580

581 Statistical analyses

We computed the appropriate sample size based on a power analysis performed through G*Power 3.1.9.2. For the main statistics, i.e. mixed ANOVA (within-between interaction) with two groups and two measurements, with the following input parameters: α equal to 0.05, power (1- β) equal to 0.95, and a hypothesized effect size (f) equal to 0.25, the estimated sample size resulted in n = 30per experimental group.

Since most variables passed the D'Agostino-Pearson omnibus normality test, parametric statistics
were adopted in each experiment.

To test the between-group differences in post-conditioning US ratings, preconditioning mean SCRs levels, mean SCRs to the CS and the US during conditioning, and mean SCRs to the US₂ during the test and the follow-up sessions, we performed Student's unpaired *t* tests. Potential differences in CS-related SCRs over the 15 trials of the conditioning phase were tested through 2×15 mixed ANOVAs with Group (aPFC *vs* sham, aPFC *vs* OC, aPFC *vs* dIPFC) as between-subject variable

594 and Trial (1-15) as within-subject variable.

To test the potential between-group differences in the implicit reactions to the CS during the conditioning session, the test session, and the follow-up session, as well as the within-group differences from conditioning to test/follow-up phases, we computed 2×2 mixed ANOVAs with Group (aPFC *vs* sham, aPFC *vs* OC, aPFC *vs* dlPFC, sham *vs* ctrl discomfort) as between-subject variable and Phase (conditioning *vs* test, conditioning *vs* follow-up) as within-subject variable. Bonferroni adjustment was applied for simple main effects analyses. To compare between-group and within-group responses to the CS and the NSs during the test session, we performed 2×3 mixed ANOVAs with Group (aPFC *vs* sham, aPFC *vs* OC, aPFC *vs* dlPFC) as between-subject variable and Tone (NS₁, CS, and NS₂) as within-subject variable. Bonferroni adjustment was applied for simple main effects analyses.

To test the between-group differences in the explicit recognition and respective confidence ratings, as well as in the perceptual discrimination and respective confidence ratings during the test and the follow-up sessions (aPFC-E *vs* sham-E), we performed Student's unpaired *t* tests. To test whether explicit recognition levels were significantly higher than the 50% chance level for each condition during the test and the follow-up sessions, we calculated Student's one sample *t* tests against 0.50.

For each ANOVA we assessed the Sphericity assumption through Mauchly's Test. Where it wasviolated, we applied the Greenhouse-Geisser correction accordingly.

The null hypothesis was rejected at P < 0.05 significance level. All statistical analyses were performed using SPSS Statistics 22 (IBM) and Prism 9 (GraphPad).

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623

624 **Competing interests**

625 The authors declare no financial interests or potential conflicts of interest.

626 **References**

- DiFazio, L. E., Fanselow, M. & Sharpe, M. J. The effect of stress and reward on encoding
 future fear memories. *Behav. Brain. Res.* 417, 113587 (2022).
- 629 2. Wilker, S., Elbert, T. & Kolassa, I. T. The downside of strong emotional memories: how
 630 human memory-related genes influence the risk for posttraumatic stress disorder-a selective
- 631 review. *Neurobiol. Learn. Mem.* **112**, 75-86 (2014).
- Biol. Psychiatry 78, 336-343 (2015).
- 4. Manassero, E., Mana, L., Concina, G., Renna, A. & Sacchetti, B. Implicit and explicit systems
 differently predict possible dangers. *Sci. Rep.* 9, 1-12 (2019).
- 5. Taylor, S., Abramowitz, J. S. & McKay, D. Non-adherence and nonresponse in the treatment of
 anxiety disorders. *J. Anxiety Disord.* 26, 583-589 (2012).
- 638 6. Fonzo, G. A. et al. PTSD psychotherapy outcome predicted by brain activation during 639 emotional reactivity and regulation. *Am. J. Psychiatry* **174**, 1163-1174 (2017).
- Marković, V., Vicario, C. M., Yavari, F., Salehinejad, M. A. & Nitsche, M. A. A systematic
 review on the effect of transcranial direct current and magnetic stimulation on fear memory and
 extinction. *Front. Hum. Neurosci.* 15, 1-26 (2021).
- 8. Asthana, M. et al. Effects of transcranial direct current stimulation on consolidation of fear
 memory. *Front. Psychiatry* 4, 107 (2013).
- Mungee, A., Kazzer, P., Feeser, M., Nitsche, M. A., Schiller, D. & Bajbouj, M. Transcranial
 direct current stimulation of the prefrontal cortex: a means to modulate fear memories. *Neuroreport* 25, 480-484 (2014).
- Mungee, A., Burger, M. & Bajbouj, M. No effect of cathodal transcranial direct current
 stimulation on fear memory in healthy human subjects. *Brain. Sci.* 6, 55 (2016).
- 650 11. Abend, R. et al. Modulation of fear extinction processes using transcranial electrical
 651 stimulation. *Transl. Psychiatry* 6, e913-e913 (2016).

- Van't Wout, M., Mariano, T. Y., Garnaat, S. L., Reddy, M. K., Rasmussen, S. A. & Greenberg,
 B. D. Can transcranial direct current stimulation augment extinction of conditioned fear? *Brain Stimul.* 9, 529-536 (2016).
- Roesmann, K. et al. Transcranial direct current stimulation of the ventromedial prefrontal
 cortex modulates perceptual and neural patterns of fear generalization. *Biol. Psychiatry Cogn. Neurosci. Neuroimaging* 7, 210-220 (2022).
- Miniussi, C. et al. Efficacy of repetitive transcranial magnetic stimulation/transcranial direct
 current stimulation in cognitive neurorehabilitation. *Brain stimul.* 1, 326-336 (2008).
- Elder, G. J. & Taylor, J. P. Transcranial magnetic stimulation and transcranial direct current
 stimulation: treatments for cognitive and neuropsychiatric symptoms in the neurodegenerative
 dementias? *Alzheimer's Res. Ther.* 6, 1-11 (2014).
- 663 16. Guhn, A. et al. Medial prefrontal cortex stimulation modulates the processing of conditioned
 664 fear. *Front. Behav. Neurosci.* 8, 44 (2014).
- Raij, T., Nummenmaa, A., Marin, M. F., Porter, D., Furtak, S., Setsompop, K. & Milad, M. R.
 Prefrontal cortex stimulation enhances fear extinction memory in humans. *Biol. Psychiatry* 84, 129-137 (2018).
- Borgomaneri, S., Battaglia, S., Garofalo, S., Tortora, F., Avenanti, A. & Di Pellegrino, G.
 State-dependent TMS over prefrontal cortex disrupts fear-memory reconsolidation and prevents
 the return of fear. *Curr. Biol.* **30**, 3672-3679 (2020).
- 19. Su, S. et al. Continuous theta-burst stimulation over the right dorsolateral prefrontal cortex
 disrupts fear memory reconsolidation in humans. *iScience* 25, 103614 (2022).
- 20. Vervliet, B., Craske, M. G. & Hermans, D. Fear extinction and relapse: state of the art. *Annu. Rev. Clin. Psychol.* 9, 215-248 (2013).
- 21. Ramnani, N. & Owen, A. M. Anterior prefrontal cortex: insights into function from anatomy
- 676 and neuroimaging. *Nat. Rev. Neurosci.* **5**, 184-194 (2004).

- Semendeferi, K., Armstrong, E., Schleicher, A., Zilles, K. & van Hoesen, G. W. Prefrontal
 cortex in humans and apes: a comparative study of area 10. *Am. J. Phys. Anthropol.* 114, 224241 (2001).
- Volman, I. et al. Reduced serotonin transporter availability decreases prefrontal control of the
 amygdala. *J. Neurosci.* 33, 8974-8979 (2013).
- Koch, S. B. J., Mars, R. B., Toni, I. & Roelofs, K. Emotional control, reappraised. *Neurosci. Biobehav. Rev.* 95, 528-534 (2018).
- Bramson, B. et al. Human lateral frontal pole contributes to control over emotional approach–
 avoidance actions. *J. Neurosci.* 40, 2925-2934 (2020).
- 686 26. Folloni, D. et al. Dichotomous organization of amygdala/temporal-prefrontal bundles in both
 687 humans and monkeys. *Elife* 8, 1-23 (2019).
- Peng, K., Steele, S. C., Becerra, L. & Borsook, D. Brodmann area 10: collating, integrating and
 high level processing of nociception and pain. *Prog. Neurobiol.* 161, 1-22 (2018).
- 690 28. Klumpers, F. et al. Prefrontal mechanisms of fear reduction after threat offset. *Biol. Psychiatry*691 68, 1031-1038 (2010).
- 29. Lanius, R. A. et al. Functional connectivity of dissociative responses in posttraumatic stress
 disorder: a functional magnetic resonance imaging investigation. *Biol. Psychiatry* 57, 873-884
 (2005).
- 30. Morey, R. A., Petty, C. M., Cooper, D. A., LaBar, K. S. & McCarthy, G. Neural systems for
 executive and emotional processing are modulated by symptoms of posttraumatic stress
 disorder in Iraq War veterans. *Psychiatry Res. Neuroimaging* 162, 59-72 (2008).
- 31. Sadeh, N. et al. Neurobiological indicators of disinhibition in posttraumatic stress disorder. *Hum. Brain Mapp.* 36, 3076-3086 (2015).
- 32. Sadeh, N. et al. SKA2 methylation is associated with decreased prefrontal cortical thickness
 and greater PTSD severity among trauma-exposed veterans. *Mol. Psychiatry* 21, 357-363
 (2016).

- Xaldewaij, R., Koch, S. B., Hashemi, M. M., Zhang, W., Klumpers, F. & Roelofs, K. Anterior
 prefrontal brain activity during emotion control predicts resilience to post-traumatic stress
 symptoms. *Nat. Hum. Behav.* 5, 1055-1064 (2021).
- 706 34. Fonzo, G. A. et al. Selective effects of psychotherapy on frontopolar cortical function in PTSD.
 707 *Am. J. Psychiatry* 174, 1175-1184 (2017).
- 35. Resnik, J. & Paz, R. Fear generalization in the primate amygdala. *Nat. Neurosci.* 18, 188-190
 (2015).
- 36. Wong, A. H. & Lovibond, P. F. Rule-based generalisation in single-cue and differential fear
 conditioning in humans. *Biol. Psychol.* 129, 111-120 (2017).
- 37. Grosso, A., Santoni, G., Manassero, E., Renna, A. & Sacchetti, B. A neuronal basis for fear
 discrimination in the lateral amygdala. *Nat. Commun.* 9, 1-12 (2018).
- 38. Concina, G., Cambiaghi, M., Renna, A. & Sacchetti, B. Coherent activity between the
 prelimbic and auditory cortex in the slow-gamma band underlies fear discrimination. *J. Neurosci.* 38, 8313-8328 (2018).
- 39. Grosso, A., Cambiaghi, M., Milano, L., Renna, A., Sacco, T. & Sacchetti, B. Region-and layerspecific activation of the higher order auditory cortex Te2 after remote retrieval of fear or
 appetitive memories. *Cereb. Cortex* 27, 3140-3151 (2017).
- 40. Hanlon, C. A., Dowdle, L. T., Gibson, N. B., Li, X., Hamilton, S., Canterberry, M. & Hoffman,
 M. Cortical substrates of cue-reactivity in multiple substance dependent populations:
 transdiagnostic relevance of the medial prefrontal cortex. *Transl. Psychiatry* 8, 1-8 (2018).
- 41. Jasper, H. H. The ten-twenty electrode system of the International Federation.
 Electroencephalogr. *Clin. Neurophysiol.* 10, 370-375 (1958).
- 42. Herrmann, M. J., Katzorke, A., Busch, Y., Gromer, D., Polak, T., Pauli, P. & Deckert, J.
 Medial prefrontal cortex stimulation accelerates therapy response of exposure therapy in
 acrophobia. *Brain Stimul.* 10, 291-297 (2017).

- 43. Karmann, A. J., Maihöfner, C., Lautenbacher, S., Sperling, W., Kornhuber, J. & Kunz, M. The
 role of prefrontal inhibition in regulating facial expressions of pain: a repetitive transcranial
 magnetic stimulation study. *J. Pain.* 17, 383-391 (2016).
- 44. Ameli, R., Ip, C. & Grillon, C. Contextual fear-potentiated startle conditioning in humans:
 Replication and extension. *Psychophysiology* 38, 383-390 (2001).
- 45. Maren, S., Phan, K. L. & Liberzon, I. The contextual brain: implications for fear conditioning,
 extinction and psychopathology. *Nat. Rev. Neurosci.* 14, 417-428 (2013).
- 46. Sacco, T. & Sacchetti, B. Role of secondary sensory cortices in emotional memory storage and
 retrieval in rats. *Science* 329, 649-656 (2010).
- 47. Sacchetti, B., Baldi, E., Tassoni, G. & Bucherelli, C. Memorization of contextual and CS
 conditioned fear response (freezing) in a one-trial acquisition paradigm. *Arch. Ital. Biol.* 137,
 235-248 (1999).
- 48. Onat, S. & Büchel, C. The neuronal basis of fear generalization in humans. *Nat. Neurosci.* 18, 1811-1818 (2015).
- 49. Laufer, O., Israeli, D. & Paz, R. Behavioral and neural mechanisms of overgeneralization in
 anxiety. *Curr. Biol.* 26, 713-722 (2016).
- 50. Manassero, E., Giordano, A., Raimondo, E., Cicolin, A. & Sacchetti, B. Sleep deprivation
- during memory consolidation, but not before memory retrieval, widens threat generalization to
 new stimuli. *Front. Neurosci.* 781, 1-12 (2022).
- 51. Hiser, J. & Koenigs, M. The multifaceted role of the ventromedial prefrontal cortex in emotion,
 decision making, social cognition, and psychopathology. *Biol. Psychiatry* 83, 638-647 (2018).
- 52. Catani, M. et al. Short frontal lobe connections of the human brain. *Cortex* **48**, 273-291 (2012).
- 53. Orr, J. M., Smolker, H. R. & Banich, M. T. Organization of the human frontal pole revealed by
- ⁷⁵¹ large-scale DTI-based connectivity: implications for control of behavior. *PloS one* **10**, 1-23
- 752 (2015).

- 54. Wu, Y., Sun, D., Wang, Y. & Wang, Y. Subcomponents and connectivity of the inferior frontooccipital fasciculus revealed by diffusion spectrum imaging fiber tracking. *Front. Neuroanat.*10, 1-13 (2016).
- 55. Pavlov, I. P. Conditioned Reflexes. (London: Oxford Univ. Press., 1927).
- 757 56. Rachman, S. Studies in desensitization III: speed of generalization. *Behav. Res. Ther.* 4, 7-15
 758 (1966).
- 759 57. Pascual-Leone, A., Tormos, J. M., Keenan, J., Tarazona, F., Cañete, C. & Catalá, M. D. Study
 760 and modulation of human cortical excitability with transcranial magnetic stimulation. *Clin.*761 *Neurophysiol.* 15, 333-343 (1998).
- 58. Luber, B. & Deng, Z. D. Application of non-invasive brain stimulation in psychophysiology.
 In: Berntson, G. G., Cacioppo, J. T. & Tassinary, L. G. (Eds.). *Handbook of Psychophysiology, 4th ed.* (Cambridge University Press, Cambridge, pp. 116-150, 2016).
- 59. Beynel, L., Powers, J. P. & Appelbaum, L. G. Effects of repetitive transcranial magnetic
 stimulation on resting-state connectivity: A systematic review. *NeuroImage* 211, 116596
 (2020).
- 60. Eisenegger, C., Treyer, V., Fehr, E. & Knoch, D. Time-course of "off-line" prefrontal rTMS
 effects—a PET study. *NeuroImage* 42, 379-384 (2008).
- 61. Nahas, Z. et al. Unilateral left prefrontal transcranial magnetic stimulation (TMS) produces
 intensity-dependent bilateral effects as measured by interleaved BOLD fMRI. *Biol. Psychiatry*50, 712-720 (2001).
- Dunsmoor, J. E., Bandettini, P. A. & Knight, D. C. Impact of continuous versus intermittent
 CS-UCS pairing on human brain activation during Pavlovian fear conditioning. *Behav. Neurosci.* 121, 635-642 (2007).
- 63. Dunsmoor, J. E., Bandettini, P. A. & Knight, D. C. Neural correlates of unconditioned response
- diminution during Pavlovian conditioning. *NeuroImage* **40**, 811-817 (2008).

- 64. Vicario, C. M., Salehinejad, M. A., Felmingham, K., Martino, G. & Nitsche, M. A. A
 systematic review on the therapeutic effectiveness of non-invasive brain stimulation for the
 treatment of anxiety disorders. *Neurosci. Biobehav. Rev.* 96, 219-231 (2019).
- 65. Pape, H. C. & Pare, D. Plastic synaptic networks of the amygdala for the acquisition,
 expression, and extinction of conditioned fear. *Physiol. Rev.* **90**, 419-463 (2010).
- 66. Sacchetti, B., Baldi, E., Lorenzini, C. A. & Bucherelli, C. Cerebellar role in fear-conditioning
 consolidation. *PNAS* 99, 8406-8411 (2002).
- 67. Sacchetti, B., Scelfo, B., Tempia, F. & Strata, P. Long-term synaptic changes induced in the
 cerebellar cortex by fear conditioning. *Neuron* 42, 973-982 (2004).
- 787 68. Zhu, L., Sacco, T., Strata, P. & Sacchetti, B. Basolateral amygdala inactivation impairs
 788 learning-induced long-term potentiation in the cerebellar cortex. *PloS one* 6, e16673 (2011).
- 69. Grosso, A. et al. The higher order auditory cortex is involved in the assignment of affective
 value to sensory stimuli. *Nat. Comm.* 6, 1-14 (2015).
- 70. Manassero, E., Renna, A., Milano, L. & Sacchetti, B. Lateral and basal amygdala account for
 opposite behavioral responses during the long-term expression of fearful memories. *Sci. Rep.* 8,
 1-12 (2018).
- 794 71. Cambiaghi, M. et al. Higher-order sensory cortex drives basolateral amygdala activity during
 795 the recall of remote, but not recently learned fearful memories. *J. Neurosci.* 36, 1647-1659
 796 (2016a).
- 797 72. Cambiaghi M, Grosso A, Renna A. & Sacchetti B. Differential recruitment of auditory cortices
 798 in the consolidation of recent auditory fearful memories. *J. Neurosci.* 36, 8586-8597 (2016b).
- 799 73. Concina, G., Renna, A., Grosso, A. & Sacchetti, B. The auditory cortex and the emotional
 valence of sounds. *Neurosci. Biobehav. Rev.* 98, 256-264 (2019).
- 74. You, Y., Brown, J. & Li, W. Human sensory cortex contributes to the long-term storage of
 aversive conditioning. *J. Neurosci.* 41, 3222-3233 (2021).

- 803 75. You, Y., Novak, L. R., Clancy, K. J. & Li, W. Pattern differentiation and tuning shift in human
 804 sensory cortex underlie long-term threat memory. *Curr. Biol.* 32, 2067-2075 (2022).
- 76. Ojala, K. E., Staib, M., Gerster, S., Ruff, C. C. & Bach, D. R. Inhibiting human aversive
 memory by transcranial theta-burst stimulation to the primary sensory cortex. *Biol. psychiatry*92, 149-157 (2022).
- 808 77. Monfils, M. H. The high road to inhibiting fear memories. *Biol. Psychiatry* **92**, 102-103 (2022).
- 809 78. Motzkin, J. C., Philippi, C. L., Wolf, R. C., Baskaya, M. K. & Koenigs, M. Ventromedial
 810 prefrontal cortex is critical for the regulation of amygdala activity in humans. *Biol. Psychiatry*811 77, 276-284 (2015).
- 812 79. Faran, Y. A comment on the connection between BA10 and episodic memory. *Front. Behav.*813 *Neurosci.* 17, 1-3 (2023).
- 80. Bechara, A., Tranel, D., Damasio, H., Adolphs, R., Rockland, C. & Damasio, A. R. Double
 dissociation of conditioning and declarative knowledge relative to the amygdala and
 hippocampus in humans. *Science* 269, 1115-1118 (1995).
- 817 81. LaBar, K. S. & Cabeza, R. Cognitive neuroscience of emotional memory. *Nat. Rev. Neurosci.*818 7, 54-64 (2006).
- 819 82. Knight, D. C., Waters, N. S. & Bandettini, P. A. Neural substrates of explicit and implicit fear
 820 memory. *NeuroImage* 45, 208-214 (2009).
- 821 83. Rossi, S. et al. Safety and recommendations for TMS use in healthy subjects and patient
- populations, with updates on training, ethical and regulatory issues: Expert Guidelines. *Clin. Neurophysiol.* 132, 269-306 (2021).
- 824 84. Spielberger, C. D., Gorsuch, R. L., Lushene, R., Vagg, P. R. & Jacobs, G. A. *Manual for the*825 *State-Trait Anxiety Inventory*. (Palo Alto: Consulting Psychologists Press, 1983).
- 826 85. Pedrabissi, L. & Santinello, M. *Nuova versione italiana dello STAI forma Y [New Italian version of the STAI form Y]*. (Firenze: Organizzazioni Speciali, 1989).
- 828 86. Cornsweet, T. N. The staircase-method in psychophysics. Am. J. Psychol. 75, 485-491 (1962).

- 829 87. Westin, G. G., Bassi, B. D., Lisanby, S. H. & Luber, B. New York State Psychiatric Institute,
- N. Y. Determination of motor threshold using visual observation overestimates transcranial
 magnetic stimulation dosage: safety implications. *Clin. Neurophysiol.* **125**, 142-147 (2014).
- 832 88. Ando, A. et al. Embodied simulation and ambiguous stimuli: The role of the mirror neuron
 833 system. *Brain Res.* 1629, 135-142 (2015).
- 834 89. Salatino, A. et al. Transcranial magnetic stimulation of posterior parietal cortex modulates line835 length estimation but not illusory depth perception. *Front. Psychol.* 10, 1169 (2019).
- 836 90. Epstein, C. M., Schwartzberg, D. G., Davey, K. R. & Sudderth, D. B. Localizing the site of
 837 magnetic brain stimulation in humans. *Neurology* 40, 666-670 (1990).
- 838 91. Rudiak, D. & Marg, E. Finding the depth of magnetic brain stimulation: a re-evaluation.
 839 *Electroencephalogr.* 93, 358-371 (1994).
- 92. Mir-Moghtadaei, A., Giacobbe, P., Daskalakis, Z. J., Blumberger, D. M. & Downar, J.
 Validation of a 25% nasion–inion heuristic for locating the dorsomedial prefrontal cortex for
 repetitive transcranial magnetic stimulation. *Brain. Stimul.* 9, 793-795 (2016).
- 843 93. Rojas, G. M., Alvarez, C., Montoya, C. E., De la Iglesia-Vaya, M., Cisternas, J. E. & Gálvez,
 844 M. Study of resting-state functional connectivity networks using EEG electrodes position as
 845 seed. *Front. Neurosci.* 12, 235 (2018).
- 846 94. Brighina, F., Ricci, R., Piazza, A., Scalia, S., Giglia, G. & Fierro, B. Illusory contours and
 847 specific regions of human extrastriate cortex: evidence from rTMS. *Eur. J. Neurosci.* 17, 2469848 2480 (2003).
- 95. Mir-Moghtadaei, A. et al. Concordance between BeamF3 and MRI-neuronavigated target sites
 for repetitive transcranial magnetic stimulation of the left dorsolateral prefrontal cortex. *Brain. Stimul.* 8, 965-973 (2015).
- 96. Lissek, S., Bradford, D. E., Alvarez, R. P., Burton, P., Espensen-Sturges, T., Reynolds, R. C. &
- 853 Grillon, C. Neural substrates of classically conditioned fear-generalization in humans: a
- parametric fMRI study. Soc. Cogn. Affect. Neurosci. 9, 1134-1142 (2014).

- 97. Dunsmoor, J. E., Kroes, M. C., Braren, S. H. & Phelps, E. A. Threat intensity widens fear
 generalization gradients. *Behav. Neurosci.* 131, 168-175 (2017).
- 98. Holt, D. J., Boeke, E. A., Wolthusen, R. P., Nasr, S., Milad, M. R. & Tootell, R. B. A
 parametric study of fear generalization to faces and non-face objects: relationship to
 discrimination thresholds. *Front. Hum. Neurosci.* 8, 1-12 (2014).
- 99. Macmillan, N. A. & Creelman, C. D. Detection theory: A user's guide. (New York: Psychology press, 2004).
- 100.Schiller, D., Monfils, M. H., Raio, C. M., Johnson, D. C., LeDoux, J. E. & Phelps, E. A.
- Preventing the return of fear in humans using reconsolidation update mechanisms. *Nature* 463,
 49-53 (2010).
- 101.Battaglia, S., Garofalo, S. & Di Pellegrino, G. Context-dependent extinction of threat
 memories: Influences of healthy aging. *Sci. Rep.* 8, 1-13 (2018).
- 102.Lykken, D. T. & Venables, P. H. Direct measurement of skin conductance: A proposal for
 standardization. *Psychophysiology* 8, 656-672 (1971).

869	Table 1. Experimental groups' descriptive, experimental, and clinical data. The table reports,
870	for each experimental condition: sample size (N), sex distribution (F = Female, M = Male), mean
871	age, State-Trait Anxiety Inventory Form Y (STAI-Y) State subscale score during session 1 (S1),
872	session 2 (S2), and session 3 (S3), and Trait subscale score, US current intensity (mA), post-
873	conditioning US rating, rTMS resting motor threshold (rMT), rTMS power, and discomfort
874	stimulation (DS) current intensity (mA). All data are mean \pm standard deviation.

Group	N	Sex	Age	STAI-Y State (S1)	STAI-Y State (S2)	STAI-Y State (S3)	STAI-Y Trait	US (mA)	US rating	rTMS rMT	rTMS power	DS (mA)
aPFC	30	18F 12M	24.45 ± 3.78	30.97 ± 4.07	32.47 ± 7.16	30.60 ± 6.04	39.27 ± 6.18	4.92 ± 2.06	5.28 ± 0.90	58.20 ± 6.40	39.73 ± 1.11	-
sham	30	18F 12M	23.35 ± 2.35	33.23 ± 5.86	32.70 ± 7.74	31.87 ± 6.51	38.77 ± 4.02	4.88 ± 2.45	5.47 ± 0.88	-	-	-
OC	30	18F 12M	24.14 ± 2.62	32.33 ± 5.51	31.53 ± 7.57	30.60 ± 6.75	39.03 ± 5.12	4.99 ± 3.17	5.28 ± 1.06	60.90 ± 6.67	39.70 ± 1.47	-
dIPFC	30	18F 12M	23.91 ± 3.15	31.70 ± 5.40	30.83 ± 7.04	30.13 ± 5.88	39.17 ± 5.85	5.16 ± 2.43	5.57 ± 1.45	58.77 ± 5.89	39.90 ± 0.40	-
aPFC-E	21	13F 8M	24.39 ± 2.43	31.71 ± 4.89	30.90 ± 5.66	30.48 ± 4.96	38.29 ± 6.21	5.13 ± 1.86	5.43 ± 0.94	58.67 ± 7.16	39.52 ± 1.54	-
sham-E	21	13F 8M	23.83 ± 2.73	33.10 ± 5.59	31.48 ± 5.54	30.38 ± 7.73	38.29 ± 5.22	5.27 ± 3.19	5.31 ± 1.31	-	-	-
ctrl discomfort	10	5F 5M	22.34 ± 3.67	34.40 ± 4.20	36.50 ± 6.47	34.20 ± 5.98	39.70 ± 4.03	6.97 ± 4.14	5.65 ± 1.11	-	-	6.65 ± 2.25

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Figure 1. Schematic diagram depicting the experimental outline and rTMS conditions. In the 879 first session (day 1, context A), participants underwent a single-cue threat conditioning in which a 880 tone (CS) was paired with a mild electrical shock (US). In the second session (day 8, context B), a 881 1Hz-rTMS procedure was actively applied over the medial anterior prefrontal cortex (aPFC, n = 30; 882 883 aPFC-E, n = 21), sham-applied over the same site (sham, n = 30; sham-E, n = 21), actively applied over the left occipital cortex (OC, n = 30) and over the left dorsolateral prefrontal cortex (dlPFC, n884 = 30). In the implicit conditions (aPFC, sham, OC, dlPFC), subjects underwent an implicit test 885 886 during which they were presented with the CS and two new stimuli (NS₁ and NS₂) and then an unconditioned threat test while being recorded in their SCRs. In the explicit conditions (aPFC-E, 887 sham-E), participants underwent an explicit 2AFC recognition task during which they were 888 889 presented with tone pairs each composed of the CS and one of the two NSs, and they were asked to recognize the CS providing a confidence level for each choice. Last, participants underwent a 2AFC 890 891 perceptual discrimination test, in which they had to judge whether the two tones in each pair (CS and/or NSs) were "the same tone" or "different tones". The third session (day 15, context A) was 892 identical to the second one except for the absence of the rTMS. 893

Figure 2. Effects of rTMS over the aPFC on immediate and remote implicit threat memory, 894 895 threat generalization to new stimuli, and overall electrodermal responsivity. (A) Simulation of rTMS effects on the neural tissue of the medial aPFC (medial BA 10), performed with SimNIBS 896 4.0 software. The magnitude of the electric field is expressed in V/m. (**B**, **C**) Dot plot and line chart 897 representing the mean SCRs elicited by the CS during the conditioning session and test session in 898 the two different conditions. Groups' reactions were not different during the conditioning phase, 899 900 whereas during the test phase the group stimulated over the aPFC showed attenuated implicit reactions relative to the sham condition. The aPFC group displayed reduced autonomic reactions to 901 the CS from conditioning to test, while the sham group showed an increase in defensive responses. 902 903 (\mathbf{D}, \mathbf{E}) Implicit reactions to all the tones $(NS_1, CS, and NS_2)$ during the test session were decreased 904 in the aPFC group relative to the sham group. Although we found a significant main effect of Group and no Group \times Tone interaction effect, we reported the statistical significance marks of simple 905 main effects. (F) Implicit reactions to the US_2 during the test session were not different between 906 conditions, showing no rTMS effects on the overall electrodermal responsivity. (G, H) In the 907 follow-up session, the aPFC group enduringly demonstrated reduced implicit reactions to the CS 908 909 relative to the sham group and to the conditioning phase. (I) Implicit reactions to the US₂ during the follow-up session were not different between groups. * P < 0.05, ** P < 0.01, *** P < 0.001. All 910 911 data are mean and SEM. 2×2 mixed ANOVA followed by Bonferroni-adjusted post hoc comparisons (B, C, G, H); 2×3 mixed ANOVA followed by Bonferroni-adjusted post hoc 912 comparisons (D, E); Student's unpaired t test (F, I). 913

914 Figure 3. Effects of rTMS over the aPFC on immediate and remote explicit threat memory.

915 (A) During the test session, explicit recognition patterns were not different between the group 916 stimulated over the aPFC and the sham group. (B) During the test session, confidence ratings did 917 not differ between the two conditions. (C) During the follow-up session, aPFC-E and sham-E 918 groups identified the CS between the NSs in a not different manner. (D) During the follow-up 919 session, aPFC-E and sham-E groups were not differently confident about their explicit choices. All 920 data are mean and SEM. Student's unpaired *t* test (A, B, C, D).

Figure 4. Selective effects of rTMS over the aPFC and the left OC on the defensive responses 921 922 to threat-predictive cues. (A) Simulation of rTMS effects on the neural tissue of the left OC (BA 18/19), performed with SimNIBS 4.0 software. The magnitude of the electric field is expressed in 923 V/m. (B, C) Dot plot and line chart representing the mean SCRs elicited by the CS during the 924 conditioning session and test session in the OC group, compared with the same aPFC group of Fig. 925 2. The two groups did not differently respond during the conditioning phase, but during the test 926 927 phase the group stimulated over the aPFC showed weaker reactions than the OC group. While the defensive reactions of the aPFC group decreased from conditioning to test, those of the OC group 928 remained not differently high. (D, E) Implicit reactions to NSs during the test session did not differ 929 930 between groups. In the OC group, the responses elicited by the NS₂ were lower than those evoked by the CS and the NS_1 . (F) Implicit reactions to the US_2 during the test session were not different 931 between groups. (G, H) In the follow-up session, the aPFC group persisted in showing reduced 932 933 implicit reactions to the CS relative to the OC group. Defensive reactions of both groups decreased from the conditioning phase. Although we found a significant main effect of Phase and no Group \times 934 Phase interaction effect, we reported the statistical significance marks of simple main effects. (I) 935 Implicit reactions to the US₂ during the follow-up session were not different between groups. * P <936 0.05, *** P < 0.001. All data are mean and SEM. 2×2 mixed ANOVA followed by Bonferroni-937 adjusted post hoc comparisons (B, C, G, H); 2×3 mixed ANOVA followed by Bonferroni-adjusted 938 post hoc comparisons (D, E); Student's unpaired t test (F, I). 939

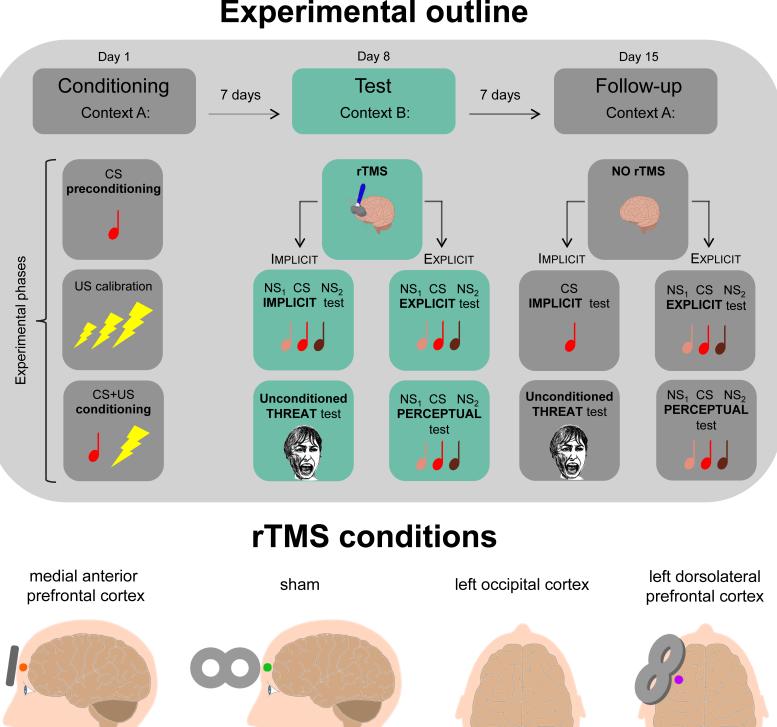
Figure 5. Different effects of rTMS over the aPFC and the left dlPFC on immediate and 940 941 remote implicit threat memory. (A) Simulation of rTMS effects on the neural tissue of the left dlPFC (BA 8/9), performed with SimNIBS 4.0 software. The magnitude of the electric field is 942 expressed in V/m. (B, C) Dot plot and line chart representing the mean SCRs elicited by the CS 943 during the conditioning session and test session in the dIPFC group, compared with the same aPFC 944 group of Fig. 2. The two conditions did not differently react during the conditioning phase, whereas 945 946 during the test phase the group stimulated over the aPFC displayed lower reactions than the dlPFC group. Implicit reactions of the aPFC group decreased from conditioning to test, while those of the 947 dlPFC group increased. (D, E) Implicit reactions to NSs during the test session did not differ 948 949 between groups. In the dIPFC group, the responses elicited by the NS₂ were lower than those evoked by the CS. (F) The two groups did not differently react to the US_2 during the test session. 950 (G, H) In the follow-up session, the aPFC group persisted in more dimly reacting to the CS relative 951 952 to the dlPFC group and to the conditioning phase. (I) Implicit reactions to the US_2 during the follow-up session were not different between groups. * P < 0.05, ** P < 0.01, *** P < 0.001. All 953 954 data are mean and SEM. 2×2 mixed ANOVA followed by Bonferroni-adjusted post hoc comparisons (B, C, G, H); 2×3 mixed ANOVA followed by Bonferroni-adjusted post hoc 955 comparisons (D, E); Student's unpaired t test (F, I). 956

957	Figure 2-figure supplement 1. Implicit reactions during preconditioning (CS) and
958	conditioning (CS, US) in the aPFC and sham groups. (A) Dot plot representing the mean SCRs
959	elicited by the CS during the preconditioning phase in the aPFC group ($n = 30$) compared with the
960	sham group ($n = 30$). Implicit reactions were not significantly different. (B) Mean SCRs elicited by
961	the US during the conditioning phase in the aPFC group compared with the sham group. Responses
962	were not significantly different. (C) Mean SCRs evoked by the CS over the 15 trials of the
963	conditioning phase, and averaged SCRs (trials 1-15) in the aPFC and sham groups. Autonomic
964	reactions were not significantly different. All data are mean and SEM. Student's unpaired t test (A,
965	B, C); 2×15 mixed ANOVA (C).

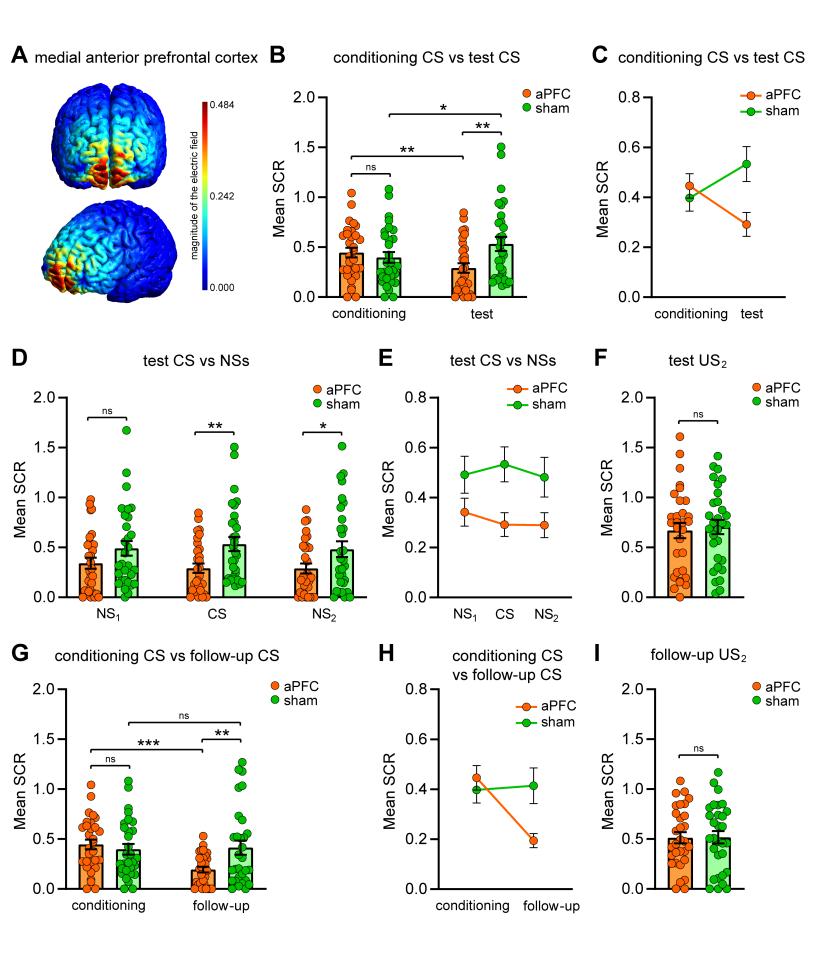
Figure 2-figure supplement 2. Effects of a discomfort-inducing procedure on immediate and 966 remote implicit threat memory. (A) Mean discomfort ratings provided by the subjects of the ctrl 967 discomfort group (n = 10) during the 10-min discomfort-inducing procedure. (B) Dot plot 968 representing the mean SCRs elicited by the CS during the conditioning phase and the test phase in 969 the ctrl discomfort group and the sham group (n = 30). The groups' reactions were not significantly 970 different during the conditioning phase as well as during the test phase (2×2 mixed ANOVA; main 971 effect of group: $F_{(1,38)} = 0.712$, P = 0.404; main effect of phase: $F_{(1,38)} = 1.713$, P = 0.198; group × 972 phase interaction: $F_{(1,38)} = 0.956$, P = 0.335). (C) In the follow-up session, SCRs to the CS did not 973 differ between groups (2 × 2 mixed ANOVA; main effect of group: $F_{(1,38)} = 1.335$, P = 0.255; main 974 effect of phase: $F_{(1,38)} = 0.042$, P = 0.838; group × phase interaction: $F_{(1,38)} = 0.175$, P = 0.678). All 975 data are mean and SEM. 976

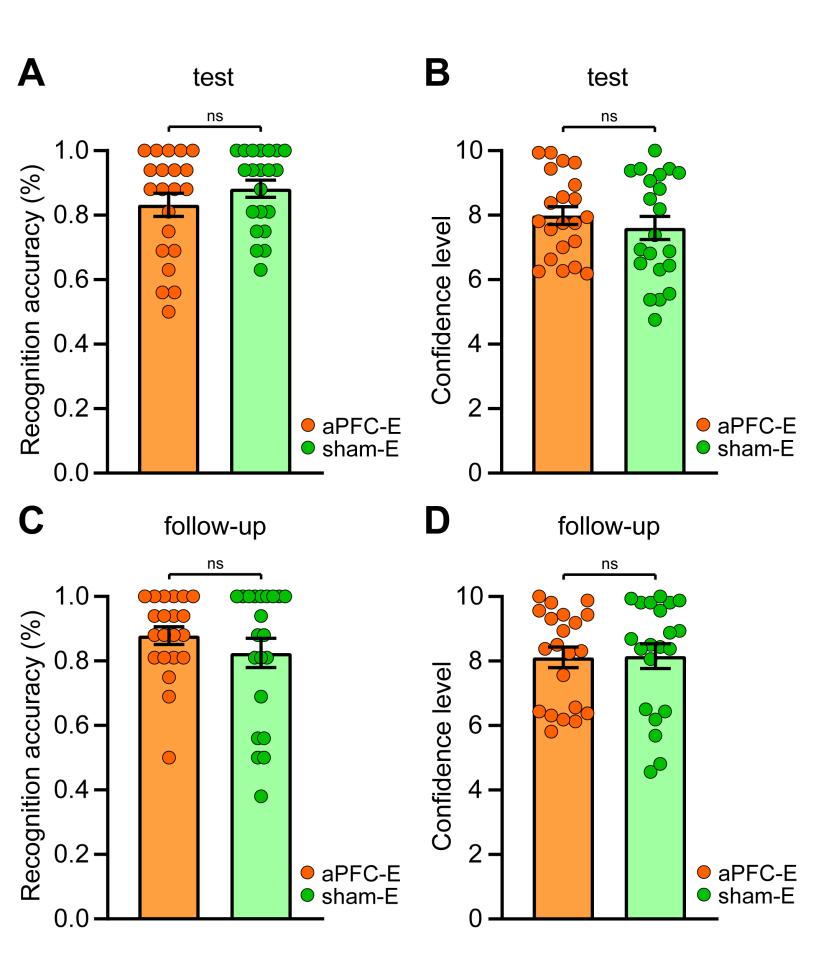
977	Figure 4-figure supplement 1. Implicit reactions during preconditioning (CS) and
978	conditioning (CS, US) in the aPFC and OC groups. (A) Dot plot representing the mean SCRs
979	elicited by the CS during the preconditioning phase in the aPFC group ($n = 30$) compared with the
980	OC group ($n = 30$). Implicit reactions were not significantly different. (B) Mean SCRs elicited by
981	the US during the conditioning phase in the aPFC group compared with the OC group. Responses
982	were not significantly different. (C) Mean SCRs evoked by the CS over the 15 trials of the
983	conditioning phase, and averaged SCRs (trials 1-15) in the aPFC and OC groups. Autonomic
984	reactions were not significantly different. All data are mean and SEM. Student's unpaired t test (A,
985	B, C); 2×15 mixed ANOVA (C).

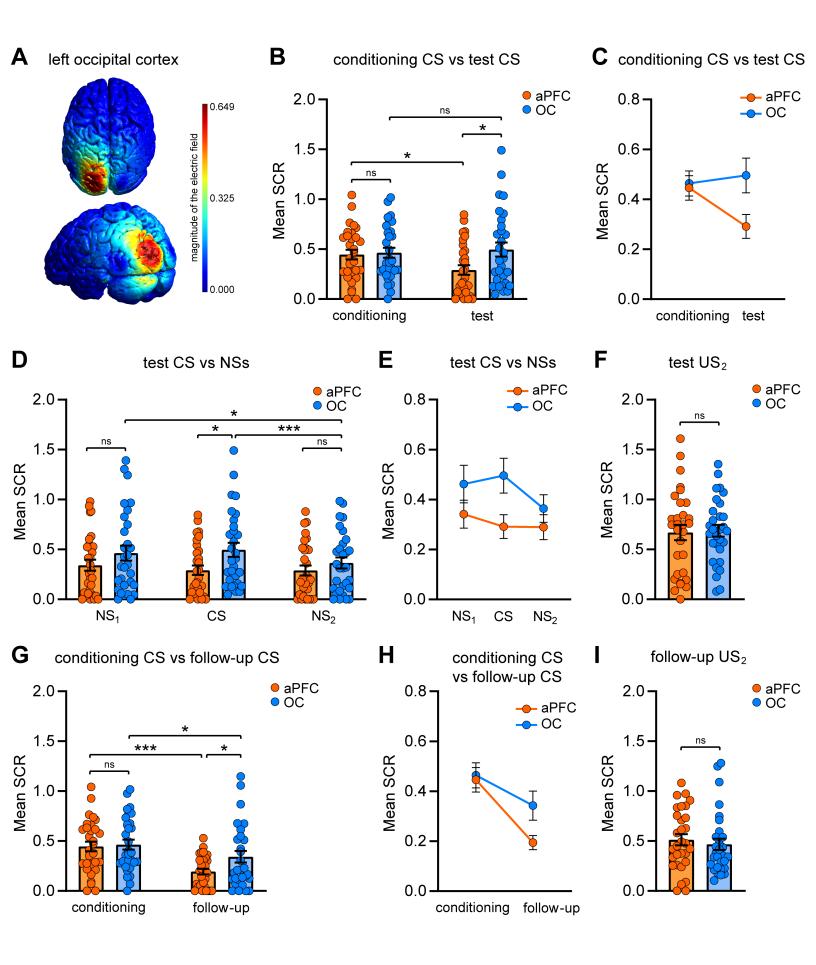
Figure 5-figure supplement 1. Implicit reactions during preconditioning (CS) and 986 conditioning (CS, US) in the aPFC and dIPFC groups. (A) Dot plot representing the mean SCRs 987 elicited by the CS during the preconditioning phase in the aPFC group (n = 30) compared with the 988 dlPFC group (n = 30). Implicit reactions were not significantly different. (**B**) Mean SCRs elicited by 989 the US during the conditioning phase in the aPFC group compared with the dlPFC group. 990 Responses were not significantly different. (C) Mean SCRs evoked by the CS over the 15 trials of 991 the conditioning phase, and averaged SCRs (trials 1-15) in the aPFC and dlPFC groups. Autonomic 992 reactions were not significantly different. All data are mean and SEM. Student's unpaired t test (A, 993 B, C); 2×15 mixed ANOVA (C). 994

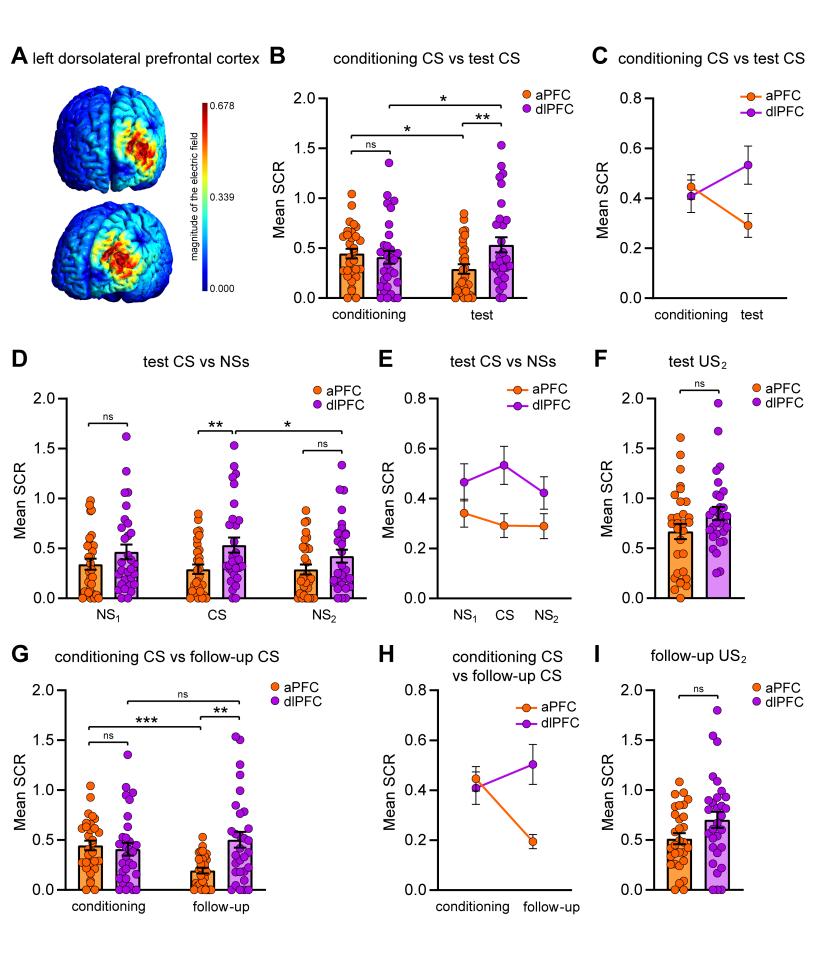


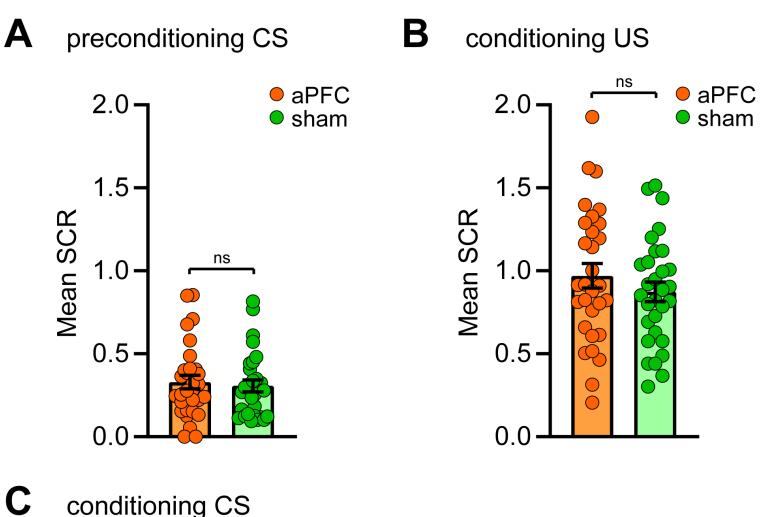
Experimental outline



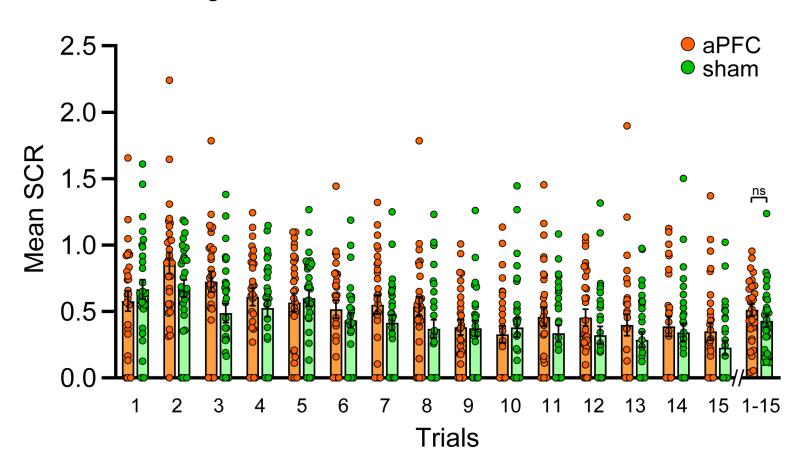


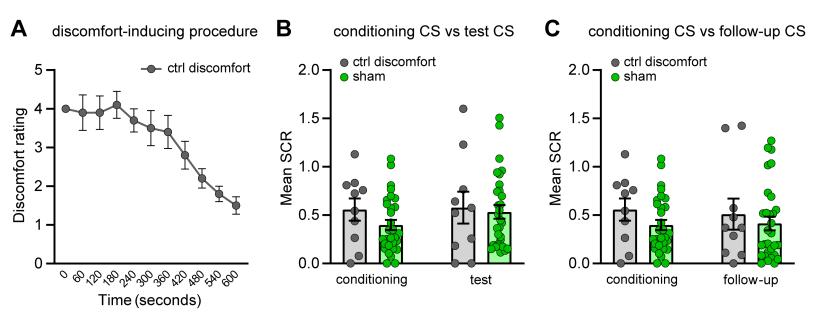


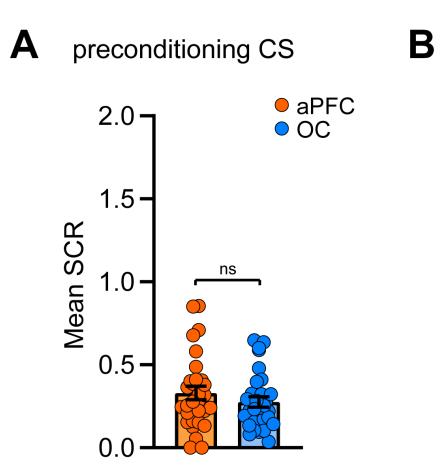


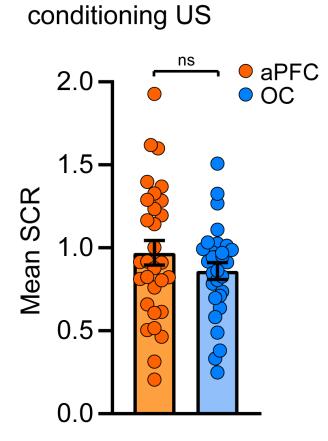


conditioning CS

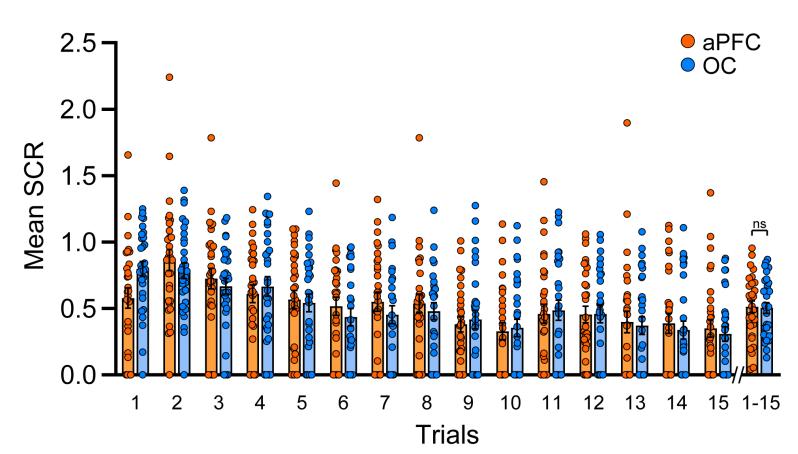


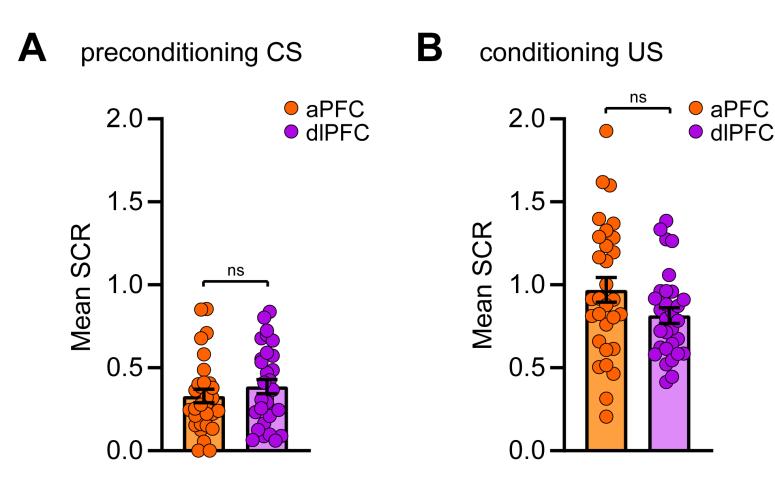






C conditioning CS





conditioning CS

С

