



Article Metal Oxalates as a CO₂ Solid State Reservoir: The Carbon Capture Reaction

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Abstract: To maintain the carbon dioxide concentration below the no-return threshold for climate change, we must consider the reduction in anthropic emissions coupled to carbon capture methods applied in synergy. In our recent papers, we proposed a green and reliable method for carbon mineralization using ascorbic acid aqueous solution as the reducing agent for carbon (IV) to carbon (III), thus obtaining oxalic acid exploiting green reagents. Oxalic acid is made to mineralize as calcium (as the model cation) oxalate. Oxalates are solid-state reservoirs suitable for long-term carbon storage or carbon feedstock for manufacturing applications. The carbon mineralization reaction is a double-step process (carbon reduction and oxalate precipitation), and the carbon capture efficiency is invariably represented by a double-slope curve we formerly explained as a decrease in the reducing effectiveness of ascorbic acid during reaction. In the present paper, we demonstrated that the reaction proceeds via a "pure CO₂-capture" stage in which ascorbic acid oxidizes into dehydroascorbic acid and carbon (IV) reduces to carbon (III) and a "mixed" stage in which the redox reaction competes with the degradation of ascorbic acid in producing oxalic acid. Despite the irreversibility of the reduction reaction, that was demonstrated in abiotic conditions, the analysis of costs according to the market price of the reagents endorses the application of the method.

Keywords: carbon mineralization; carbon capture; metal oxalate; ascorbic acid; carbon isotopes

1. Introduction

The concentration of CO₂ in the atmosphere is escalating due to anthropic contributions overlapping with natural outputs, and the trend will not be inverted in the near future. Recently, the IPCC Climate Change 2022 report [1] suggested, among others, carbon mineralization (carbonation in the conventional meaning) as a reliable method for permanent carbon sequestration into solids with foreseeable applications. Carbonation as a CO₂ disposal method exploiting exothermic silicate-weathering reactions has been proposed as a potential alternative method to classical geological storage since the 1990s, originally considering wollastonite (CaSiO₃) as the natural calcium source material [2]. From that point on, many papers have been published, proposing carbonation reactions inspired by the natural weathering of alkaline rocks and naturally occurring at the interface with air or circulating fluids [3,4]. As forecast by Seifritz in 1990, even if spontaneous, the natural weathering process is extremely slow, and the kinetics should be accelerated to apply it to industrial purposes [4-27]. The possibility of capturing and converting CO₂ into target products is a major part of the chemical literature, focusing on the catalytic and photoelectrochemical reduction of CO_2 to produce organic compounds, such as liquid fuels (methanol and dimethylether, formic acid or oxalic acid [28-41]). Transition metal



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Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). complexes have been proposed for CO₂ reduction since the late 1960s [35–38], with waste disposal and energy consumption representing their main disadvantages. In general, photoelectrochemical and catalytical reduction of CO₂ techniques are associated with a significant energy drawback [38]. In this context, the reduction of CO₂ to form oxalate has been accomplished by electrochemical reactions involving transition (Hg, Pb, Cu, Pd, Ag) metal complexes or anion radicals of aromatic hydrocarbons, esters, and nitriles [28,38,42,43]. Transition metal complexes and hydroxides may also be applied to convert carbon dioxide into carbonates [44,45]. Glycol–amine mixtures and polyamine coordination complexes have been reported to bind CO₂ reversibly through the formation of carbamates [46–54]. To the best of our knowledge, only a few authors have proposed CO₂ reduction in C₂O₄^{2−} to precipitate directly oxalate, always resorting to organometallic complexes or metal–sulfide clusters [42,43].

1.1. The Non-Conventional Mineralization Reaction of Carbon Dioxide into Oxalates

Within this framework, we proposed [55–57] a non-conventional carbon mineralization reaction based on the green reduction of C(IV) to C(III) using ascorbic acid (H₂A) in an aqueous solution as the electron donor (irreversibly oxidized). CO₂ is converted into $C_2O_4^{2^-}$, giving rise to the equilibria of H₂C₂O₄ in water (Equation (1)).

$$2CO_2 + H_2A \rightarrow DHA + H_2C_2O_4 \tag{1}$$

The reaction can be described by means of two apparently simple sub-reactions (Equations (2) and (3)).

$$2CO_2 + 2e^- + 2H^+ \to H_2C_2O_4$$
 (2)

$$H_2A \rightarrow DHA + 2H^+ + 2e^- \tag{3}$$

Equation (2) describes the reduction of C(IV) to C(III) by means of the ascorbic acid acting as the electron donor: from two molecules of CO_2 , a molecule of oxalic acid is spontaneously obtained. Equation (3) describes the oxidation reaction of ascorbic acid into dehydroascorbic acid.

In the presence of calcium as the model cation, both in stoichiometric and nonstoichiometric ratios ($Ca^{2+}/H_2A < 1$), along with an initial pH always equal to or lower than 5.5, oxalate anions precipitate into crystalline calcium oxalate, mainly dihydrate (COD, mineral weddellite), while some calcium oxalate monohydrate (COM, mineral whewellite) is found as the only byproduct of the reaction [57]. This behavior is chemically described by Equation (4) whilst the mineralogy of crystalline products was investigated by XRPD.

$$2CO_2 + Ca[HA]_2 \rightarrow 2HA^- + CaC_2O_4 \tag{4}$$

As demonstrated in our previous papers [55,57], the reaction's performance ranges between 6 and 82% of carbon dioxide removed from the atmosphere, depending on the experimental setup, while it is constant and stable for each reactor setup. We carefully addressed this point in our previous works, showing that the larger the reaction surface, the better the yield, reaching a remarkable value of 82% with a fine-divided (sprayed) solution immersed into pure CO_2 atmosphere. We also provided a model for predicting yield increase as a function of the increase in the area-to-volume ratio of gas solution [57] which was consistent with the experimental yield data published subsequently [55].

The products of the degradation of ascorbic acid were analyzed to ensure that the reaction does not produce any harmful substances [57]. It was demonstrated [57,58] that the byproducts follow the sequence: dehydroascorbic acid \rightarrow diketogulonic acid \rightarrow 2,3,4 trihydroxybutanoic acid + oxalic acid. The first product represents the early step of H₂A degradation, and the second an intermediate product, while the last ones are both common metabolites in plants. To the best of our knowledge, safety concerns are not an issue with any of the molecules produced.

As reported in our previous papers and summarized in Figure 1, the experimental curves recorded from the simplest experimental setup (B-setup in Figure S1 in Supplementary materials) show a common trend with a double slope in the CO₂ capture rate [55,57]. We preliminarily related this trend to the progress of the redox reaction and, accordingly, to the concentration of the oxalic acid produced by the C(IV) to C(III) reaction. The steep slope at the beginning of the experiment was correlated with the highest reducing power of the fresh H₂A solution, corresponding to the first peak in calcium oxalate supersaturation and, consequently, to the highest nucleation frequency for COD (and, eventually, COM as a crystalline byproduct). Then, along with H₂A degradation, the carbon capture efficiency slope decreased along with the C(IV) to C(III) reduction rate.



Figure 1. General trend in the carbon capture curves as determined during capture experiments (B-setup, Figure S1 in the Supplementary Materials). The double slope was roughly associated with the kinetics of the redox reaction, i.e., fast at the initial stages when H₂A-reducing power was high and then slowed down by the degradation of H₂A [55,57].

The weddellite crystals obtained are stable crystalline pools for carbon dioxide longterm storage. We characterized their structure and thermal stability, expecting their potential reuse as feedstocks [56,57]. In fact, weddellite is a carbon feedstock for retrieval and reprocessing in industry (food and beverages, for instance) but can also be applied for metal recovery [59], biomedical applications [60–62], pre- and post-harvesting of farming soils [63–70], ceramics, chemicals, and pharmaceutics. Albeit beyond our scientific purposes, the economical sustainability of the capture reaction was positively assessed by estimating the main operational outlays (ascorbic acid supply) versus revenues from the sale of the calcium oxalate produced, with both referred to as technical-grade quality chemicals (Figure S2). In so doing, we concluded that a positive income per CO_2 -kg stored into Ca-oxalate in a pilot plant is achievable, even more so when considering no toxic waste discharge costs.

1.2. The Spontaneousness of the Reaction and Oxidation of H_2A

It must be emphasized that, even if the topic is exceptionally important for biomedical research because of the powerful antioxidant and free radical scavenger properties of H_2A , an exhaustive definition of the reactive pathway of H_2A has not yet been described in the literature. The simplified reaction as found in most articles can be written as reported in Equation (3).

Some important contributions to the comprehension of the complex reaction mechanisms of H_2A have been published in recent times. The simplified reaction must be compared to the complex deprotonation and oxidation sequence of reactions as reported in some papers and summarized in Table S1 and Figure S3 in Supplementary materials [71–74].

According to Tu and Njus [73,74], H₂A can lose two electrons in two successive steps: a first oxidation step forming an intermediate radical product (HA[•]) at the equilibrium with the ascorbate radical $A^{-\bullet}$ [73–80], and a second oxidation step that produces the completely oxidated dehydroascorbic acid (DHA). DHA in water, as usually represented in the literature, is energetically unstable because of its tricarbonyl structure (it can exist only in aprotic non-aqueous solvents) and reacts with water, keeping the γ -lactone ring of ascorbic acid or forming bicyclic hydrated forms. The free energies of the many configurations calculated ab initio differ slightly [74], the most stable being a bicyclic conformer (as demonstrated by NMR data [72]), proving that in aqueous media, hydration and cyclization of the fully oxidized dehydroascorbic acid are energetically favored. In comparison, hydration and cyclization of the ascorbate radical A^{-•} are not energetically favored. Eventually, complete oxidation can occur in a single step without intermediate products by disproportionation of two $A^{-\bullet}$ radicals into one ascorbate (HA⁻) and one dehydroascorbate (DHA4). The vitamin C redox system is extremely unstable and, in the presence of strongly oxidizing agents, oxidation may continue beyond the DHA formation, giving birth to biologically useless products and inactive compounds (the "degradation cascade" of H_2A). The rate of oxidation and the nature of the reaction's products is pH- and ionic force-dependent [73,74,81-84]. The last product of oxidation, DHA, has a pKa ~8 [85] and its E^0 has been evaluated to be about -0.2 V, depending on the authors (-0.14 V [86]; -0.174 V [80]; -0.21 V [75]).

There is a general agreement in describing the further stages of the degradation of the DHA. The irreversible delactonization of the γ -lactone ring of ascorbic acid leads to 2,3-diketogulonic acid (DKGA) [84,87–91]. DKGA is considered the earliest waste product of the H₂A degradation chain since it has no specific function in biological systems. Starting from DKGA and following its fate, the molecule continues degrading through a multiple-step process that originates several reaction products depending on the path (oxidative or non-oxidative) and the presence of enzymes [89]. In fully abiotic conditions, the decay rate of DHA depends on the pH and temperature of the solution. Still, it is not affected by the presence of oxygen, stressing that the central reaction in DHA degradation is hydration and cyclization [72,74,87]. On the contrary, oxygen enhances the oxidation of H₂A. L-lisonic acid, L-xylonic acid, and L-threonic acid are some inactive products of the degradation of DKGA, but some other products are still unknown [92].

Koshiishi [93] demonstrated the participation of a CO_2 – H_2CO_3 – $NaHCO_3$ buffer in the hydrolysis of DHA in plasma and bicarbonate-containing cell culture media (Dulbecco's modified Eagle's medium). From the literature, the final product of the H_2A to DHA degradation path is always oxalic acid.

The presence of oxalic acid at the end of the degradation path of H_2A/DHA casts shadows on the trustworthiness of the carbon capture method proposed. However, experimental data presented in our early papers [55,57] factually comply with the hypothesis of an efficient carbon capture effect.

The redox potential of H_2A is evaluated in the literature considering its physiological action and, generally, for the simplified reaction reported in Equation (3).

Borsook and Keighley [81] assessed this value at variable pH (ranging between 2.04 and 5.75) and H₂A/DHA activity ratios. The experiments were performed at 35.5 $^{\circ}$ C

because of the biochemical importance of vitamin C, and the oxidation of H_2A was obtained using iodine. At pH values higher than 5.75, the rate of decomposition of DHA was too high for reliable measurements. Thus, the redox potential values at basic pH were extrapolated from the experimental data fitting. From this early paper, the figures of formal redox potential (embedding the activities of the species, the equilibrium constants of the reactions, and with pH values other than zero) for DHA | H_2A range from -0.283 V (pH = 2) to -0.006 V (pH = 7), thus demonstrating once more the strong dependence of the redox potential on the pH and proton availability, accordingly. The redox mechanism is claimed to be represented by electron transfer followed by a rapid chemical step. The oxidized form (DHA) is known to decompose quickly [81].

Many other authors described the reaction mechanisms of ascorbic acid and their dependence on the proton availability by performing cyclic voltammetry measurements using variously finished electrodes. To cite some, in 1982, Falat and Cheng evaluated the formal redox potential of vitamin C at 0.1494 V at pH = 7.4 using an electrochemically treated graphite/epoxy electrode [94], while in 1985, Hu and coworkers determined a formal potential of 0.824 V at pH = 2 using glassy carbon electrodes [95]. A few years later, Wring and its team evaluated a formal potential of 0.594 V at pH = 5.0 using a graphite–epoxy composite electrode chemically modified with cobalt phthalocyanine [96] and, in 2006, Ren published a value of 0.299 V at a pH of 7.4 [97], measured using a glassy carbon electrode modified with caffeic acid. A recent paper by Lovander and coworkers [98] reported the values of the electrochemical properties of some water-soluble and fat-soluble vitamins at physiological pH. In this paper, the vitamin C redox potential evaluated at pH = 7.4 is set at 0.39 V (H₂A | DHA).

Matsui et al. [99] published the standard redox potential of vitamin C and other biologically critical molecules calculated considering a linear dependence between the experimental pKa values and the theoretical Gibbs energy differences in their deprotonation reactions. Both L-H₂A and DHA's calculated redox potentials range between 0.40 V and 0.55 V, in good agreement with many experimental results.

The variability in the redox potential values reported in the literature referring to the simplified Equation (2) is possibly related to the oversimplification of the reactive system and calls for an accurate characterization with reference to the application.

1.3. Aims of the Present Work

The performance of the carbon capture reaction via H_2A was thoroughly examined [55,57] in our previously published papers, and even more work is addressed on the optimization of the setup to move from the laboratory to the application. Still, some points need an explanation for a full grasp of the underlying chemical mechanism. In such a view, the aim of the present investigation is threefold:

- 1. The spontaneousness of the reaction in the abiotic conditions we applied (no enzymes, no biologically induced oxidative stress) remains a key point to be reasserted to assure the scaling-up of the reaction and its transferability to anthropic CO₂ emitters;
- 2. An unquestionable confirmation of the fact that calcium oxalate is built by carbon from the atmosphere is pivotal, so as to rule out any possibility that CaC_2O_4 may form from oxalic acid originated by the degradation of ascorbic acid;
- 3. The meaning of the carbon capture efficiency curve, and especially of the double slope shown in Figure 1, must be explained to elucidate the process mechanisms and make the system either scalable-up to the CO₂ anthropic emitters, or tunable as a function of the required Ca-oxalate crystal properties.

2. Materials and Methods

2.1. Carbon Capture Experiments

All the tests have been performed on the B-setup as described in our previous papers [55,57]. The B-setup, even if less efficient than other designs tested, such as BD or SPRAY (for details about the experimental geometries, please refer to our previous work), is

the most reliable in variable control and carbon capture effectiveness evaluation. A scheme of the B-setup is reported in Figure S1 in Supplementary Materials.

Carbon capture was operated in a thermostatic reactor at 20 °C using a 1 M stoichiometric calcium ascorbate (Ca[HA]₂) solution prepared under an inert atmosphere, if not otherwise stated, using reagent-grade ascorbic acid (Merck Group, Darmstadt, Germany) by adding 150 mL of ultrapure vacuum-degassed water (18 M Ω water ultra-purification system with resistivity measure device, Maina SrL, Torino, Italy) and carefully mixing until a clear solution was obtained. In the meanwhile, a suspension of Ca(OH)₂ was prepared by thoroughly mixing under an inert atmosphere Ca(OH)₂ powder (Merck Group, Darmstadt, Germany) with 50 mL of ultrapure and vacuum-degassed water. The suspension was added dropwise to the H₂A solution to rein in the degradation of the ascorbic acid induced by the sudden pH increase. Before the capture experiment started, the Ca[HA]₂ solution was mixed for 30 min under an inert Ar atmosphere to ensure the complete dissolution of Ca(OH)₂. The pH of the stoichiometric solution was slightly acidic (5.5), assuring no base-related carbon capture.

Pure CO₂ (Sapio Srl, Milano, Italy) was bubbled into the sealed B-reactor three times at 150 mL/min to wash it from Ar, then the reactor was loaded (150 mL/min) with carbon dioxide at a fixed pressure setpoint slightly higher than the room pressure (1.042 bar) to avoid the accidental air loading by the reaction vessel. The reaction was then left to proceed. Flow rate and pressure setpoint were controlled using an EL-FLOW valve coupled to an EL-PRESS pressure meter (Bronkhorst High-Tech B.V., Ruulo, The Netherlands). The pressure drop induced by CO₂ consumption by mineralization was continuously measured using the pressure meter during the whole reaction time.

Operating conditions of temperature and pressure were kept constant for the B-setup unless otherwise stated.

2.2. Characterization of the Precipitates

XRPD characterization was carried out using a Rigaku (Rigaku Corporation, Tokyo, Japan) MiniFlex 600 benchtop X-ray diffractometer (Bragg–Brentano geometry; CuK α radiation and an X-ray source operating at 40 kV and 15 mA; D/teX Ultra2 silicon strip detector; $2^{\circ} < 2\theta < 50^{\circ}$, step width 0.005°, and scan speed 0.3°/min). XRPD patterns were qualitatively evaluated using the EVA software (Bruker, Siemens, Germany) and the free software Profex v. 5.2.8 [100] to confirm the nature of the reaction's products.

Selected samples were imaged using a Tescan VEGA (TESCAN Brno s.r.o., Brno, Czech Republic) scanning electron microscope (W filament, 15 keV, and 10 pA working conditions).

2.3. Evaluation of the Redox Potential of Ascorbic Acid in Abiotic Conditions

As revealed by the erraticism of data from the literature, the redox potential of a reaction can be affected by several parameters, such as the nature of the solvent, the ionic strength of the solution, the speciation in the solution, the pH and processes involved, and the evaluation technique and equipment. Moreover, due to the complex sequence of deprotonation and oxidation reactions described in Section 1.2, an averaged measure is likely to allow us to state the spontaneousness of the reaction. E^0 data collected under physiological or biological-like conditions cannot be a steady point in our abiotic experiments. The boundary conditions are utterly different from those of the traditional biochemical applications of H₂A and DHA. Nevertheless, it was indispensable to our purposes to define a value of the redox potential of H₂A in our experimental conditions to solve any doubt about the effectiveness of the vitamin C in reducing carbon dioxide to mineralize it in stable oxalates, the reduction potential for carbon dioxide to produce oxalic acid being reported in the literature [101] as 0.47 V.

As a first rough step, we indirectly evaluated the redox potential of a solution of ascorbic acid 1 M, hypothesizing the simplified Equation (2) as occurring in our system.

Then, we considered Equation (5) reported as the reference reaction, forcing the hypothesis of both electrons exchanged at the same time, as follows:

$$\mathrm{Me}^{\mathrm{n}+} + 2\mathrm{e}^{-} \to \mathrm{Me}^{(\mathrm{n}-2)+} \tag{5}$$

where Meⁿ⁺ is a metal with a well-known and tabulated redox potential [101].

For this indirect evaluation of the redox potential in the experimental conditions of our CO₂ capture system, we tested the reaction of a 1 M H₂A solution prepared using ultrapure water (18 M Ω) on an inventory of chemicals starting from a list (Table S2 in the Supplementary Materials) of well-established redox potentials to determine the oxidation potential of ascorbic acid experimentally in our abiotic conditions.

 H_2A can be oxidized by transition metals via two different mechanisms: (i) inner sphere electron transfer and (ii) outer sphere electron transfer. Both mechanisms can occur, but the first oxidation step invariably involves the formation of the ascorbate radical [71]. Nitrates, sulfates, and phosphates were avoided because of the variable oxidation state of nitrogen, sulfur, and phosphorous, respectively, which may result in competition over the reduction power of H_2A . Details of the reactions applied are reported in Table S2.

The reactions were performed in 10 mL polycarbonate tubes filled with solution, leaving no headspace to ensure no oxygen participated in the reaction, and then sealed. The sealed tubes were left to react from a few minutes to a week to ensure a satisfactory reaction time for the slowest reduction processes.

At the end of the reaction, the resulting solid fraction was measured by XRPD to check the presence of crystalline phases of the reduced species.

2.4. Cyclic Voltammetry (CV)

CV measurements were performed at different scan rates to investigate the electrochemical processes which can occur at the electrode surface to better evaluate the onset of the oxidation of H₂A under inorganic conditions. Cyclic voltammetry experiments were carried out with a standard electrochemical setup, comprising a computer-controlled potentiostat, Autolab PGSTAT12 (Metrohm, Herisau, Switzerland). The electrochemical cell was a conventional three-electrode cell equipped with a 5 mm diameter glassy carbon disc (working electrode), glassy carbon rod (counter electrode), and an Ag/AgCl/KCl 3 M (reference electrode). The electrolyte was a 1.0 M HA, 0.30 M CaCl₂ solution at pH 4.2. Measurements were performed at room temperature and pressure at an aging time ranging from 0 to 26 h, in a CO₂ atmosphere, with continuous CO₂ flow of 50 mL min⁻¹ at scan rates of 50, 100, and 200 mV s⁻¹.

2.5. Liquid Chromatography (LC) Coupled to Tandem Mass Spectrometry Detection of H_2A and DHA

LC measurements have been performed on a 1 M solution of H_2A to avoid calcium, which hinders the measurements. For this reason, the results of the analysis are not perfectly aligned with those of the mineralization reaction and should be considered as a general line of interpretation of the degradation of the ascorbate. The first run was performed on H_2A reacted with CO_2 in the B-setup reactor and exposed to the air only immediately before the measurement. The second run was performed on H_2A reacted with CO_2 in the B-setup reactor and exposed to the air only immediately before the measurement. The second run was performed on H_2A reacted with CO_2 in the B-setup reactor and protected from the air during the whole experiment. In both cases, the carbon capture performance of the solution was verified and confirmed the data that we had previously published [55].

Here, 2-furfural (1 mg/mL) in MeOH was used as the internal standard (ISTD). H_2A and DHA stock solutions (1 mg/mL) diluted in disodium EDTA 0.05% (w/v) were used to evaluate the degradation rates of the samples. Samples were diluted 1/10,000 in formic acid (0.2%) and 1/2 in disodium EDTA (0.05%). The calibration curve was obtained for H_2A and DHA 2, 5, 10, 20 ppm and internal standard 10 ppm.

The chromatographic separation was achieved using an Accela System (Thermo Fisher Scientific, Waltham, MA, USA) and using a Luna-C18 column (150 mm \times 2.1 mm; 3 μ m

from Phenomenex, Torrance, CA, USA). Furthermore, 0.1% formic acid (A) and methanol (B) were used as the mobile phase. Gradient elution was programmed as follows: 3 min isocratic elution at 100%, linear decrease up to 30% in 8 min, and isocratic for 1 min. The total run time, including re-conditioning, was 17 min. Detection was obtained by the SRM method on an LCQ Fleet (Thermo Fisher Scientific, Waltham, MA, USA), equipped with the electrospray source operating in positive mode.

For quantification, the calibration curve was built by plotting the area of the analyte's peak divided by the area of the internal standard (furfural) peak towards H_2A and DHA concentrations (Figure S4 in Supplementary materials). The limit of detection (LOD) was calculated as $(3 \times SD)/m$, and the limit of quantification (LOQ) was calculated as $(10 \times SD)/m$, where SD is the standard deviation of the instrumental response and m is the slope of the calibration curve.

2.6. Carbon and Oxygen Isotopes Fractioning during Carbon Capture

The carbon and oxygen isotopic ratio of the CO₂ from the canister and the H₂A reagent were measured to be used as internal standards. The CO₂ from the canister was sampled and cryo-purified using a mix of alcohol and liquid nitrogen (T = -70 °C) before measuring, while carbon from H₂A was extracted by burning the reagent-grade H₂A used during carbon capture experiments in a sealed, free-from-air glass capsule at 620 °C. The values of d¹³C and d¹⁸O for the reagents are reported in Table S3.

The isotopic composition of the carbon dioxide dissolved in Ca[HA]₂ solution was obtained by modifying a protocol for solid-state carbonate isotopic analysis [102]: a 1 M Ca[HA]₂ solution was prepared for the B-setup experiment under an inert Ar atmosphere, as described in Section 2.1, to avoid air contamination. Carbon and oxygen were extracted by acidifying the Ca[HA]₂ solution with pure phosphoric acid (title \geq 98%). The reaction continued for 12 h at a fixed temperature of 25 °C. The CO₂ produced by the reaction was extracted and cryo-purified using liquid nitrogen (T = 196 °C) in a vacuum line. Finally, the gas was fed to the spectrometer.

The isotopic composition of the calcium oxalates precipitated during the reaction was obtained by following the same protocol for solid-state carbonate isotopic analysis: the solid sample was ground and homogenized, then directly acidified at room temperature using pure phosphoric acid and extracted in a vacuum line.

d¹³C and d¹⁸O ratios were measured by mass spectrometry IRMS (Finnigan MAT250, Thermo Fisher Scientific, Waltham, MA, USA). The internal laboratory standard is a Carrara marble sample (M43) calibrated over the international Pee Dee Belemnite (PDB) standard. The measurement uncertainty is ± 0.2 d‰.

We performed a provisional set of B-setup trials using degassed water to prepare the Ca[HA]₂ solution in an inert Ar atmosphere and filling the reactor with CO₂ (d¹³C = -43.26) up to a pressure setpoint of 1.043 bar, as described for the regular carbon capture experiment in Section 2.1, aiming to define the best experimental procedure for isotopic characterization. After 24 h, the pressure drop stood at 0.08 bar (pure CO₂ atmosphere), corresponding to a carbon capture yield of 6.5%, perfectly fitting the performance of the B-setup reported in our previous works [55]. The solution was then filtered and divided into five flasks (50 mL each) without headspace to avoid oxygen contamination, and the d¹³C and d¹⁸O were measured at increasing reaction times.

For the second set of trials, a 1 M Ca[HA]₂ solution was prepared from vacuumdegassed water. As in the other experiments, the setpoint pressure of 1.043 bar of pure CO₂ was loaded. At the end of the experiment (20 h), the pressure drop measured was -0.07 bar (pure CO₂ atmosphere), corresponding to a capture yield of 6%. The solution was then stored under an N₂ atmosphere without filtering to evaluate fractioning. This modification in the sampling procedure allowed for avoiding artifacts introduced by handling, as occurred in the first trial.

3. Results

3.1. Spontaneousness of the Reaction

The value of the oxidation potential of H_2A resulting from the raw series of metal reduction experiments ranges between 0.44 V and 0.55 V, comparable with that necessary to guarantee the spontaneousness of the reaction and fitting other experimental data and predictions from computational methods [99]. However, the difference in redox potential between H_2A and carbon dioxide to oxalate is modest, explaining the slowness in triggering the carbon reduction reaction.

Applying the reaction with H_2A , some elements, such as Cu, Ag, Hg, Cd, Mn, and S, precipitated in their metallic form directly from the solution (as copper in Figure S5 in the Supplementary Materials, [103]), allowing their straight removal from wastewater, as a side-effect of the proposed method.

It is worth stressing here that calcium is kept as a model cation to precipitate oxalates. Still, other divalent cations can substitute it (e.g., transition metals) to achieve the double goal of carbon capture and metal trapping using carbon dioxide from anthropic activities to trap metals from wastewater.

From CV measurements, the onset of the oxidation of H_2A is detectable at 0.15 V vs. Ag/AgCl, and then the current increases almost linearly until the potential scan is reversed. This value fits the data reported for the oxidation of the ascorbate radical to DHA (please refer to Table S1 and Figure S3) [74]. No anodic current peak (no complete oxidation) is observed until after 7 h of reaction. Upon the potential scan reversal, the current decreases again linearly with the applied potential; nevertheless, it remains anodic, meaning that we reached the irreversible degradation stage. No cathodic peaks are observed. The anodic current decreases monotonically with the reaction time: since it is directly proportional to the concentration of the oxidized species in the electrolyte, we can confirm that H_2A is consumed during the reaction forming its oxidized counterpart. From the current values, we can estimate that 30% to 40% of the initial ascorbic acid has been consumed after 26 h. Evidence that ascorbic acid concentration declines with time comes from the appearance of an anodic peak in the voltammetry after 21.5 h and especially at slower scan rates, i.e., 50 mV s⁻¹ (Figure S6 in the Supplementary Materials).

The oxidation of ascorbic acid is confirmed to be completely irreversible, suggesting no possibility of reagent recovery and reuse after carbon capture. Nevertheless, neither the formation of a precipitate, nor visual changes in the electrolyte, which remained clear and colorless, were observed during the reaction. This last point was also observed in mineralization experiments, and is most likely related to the supersaturation values reached during the early part of the experiment with respect to calcium oxalate. This point will be discussed more carefully in the following paragraph.

Along with CV measurements, LC confirmed that the first step of H₂A degradation into DHA begins immediately after preparing the sample. In two days and with the presence of air, a solution 1 M of H_2A is completely degraded (Figure S7a in the Supplementary Materials), the concentration of DHA being consistently lower than that theoretically expected, indicating the delactonization of the DHA ring followed by the formation of waste products of the degradation cascade. Data from CV and LC set the onset of the oxidation reaction of H_2A at the very beginning of the experiment, immediately after the preparation of the solution. CV moreover confirms that the solution is stable and effectively acts as a good reducing agent for carbon during the first day of reaction, even if its oxidation is already detectable. These data perfectly match the trend evaluated during carbon mineralization experiments where the steep slope (maximum efficiency) of the carbon capture curve in Figure 1 is restrained to the early two hours of reaction. Then the curve flattens $(H_2A \text{ oxidation detectable but with mineralization still occurring})$. From the LC data, we can infer that the reaction will work with lower performance for up to two days of carbon mineralization. Figure S7b (in the Supplementary Materials) shows the increase in DHA in the solution, matching the trend shown in Figure S7a (in the Supplementary Materials). The absence of oxygen delays the degradation by about two days.

3.2. Carbon Isotope Fractioning during CO₂ Capture/CaC₂O₄·nH₂O Precipitation

Experiments with ¹³C-labelled CO_2 have been performed to trace the carbon path within the reaction. As previously stressed, the double slope of the carbon mineralization curve in B-setup represents a recurring feature and must be explained to scale-up the method.

As described in Section 2.6, we evaluated the isotopic fingerprint of raw materials used for the reaction as our reference values.

The d¹³C and d¹⁸O ratios for commercial CO₂ were, respectively, -43.26 and -47.31, while for commercial H₂A, they were, respectively, -11.17 and -9.62, demonstrating that carbon dioxide from the canister is strongly enriched in ¹²C, and that the H₂A isotopic composition is heavier than carbon dioxide from the canister (Table S3).

The isotopic determination was performed on the CO₂ dissolved into the Ca[HA]₂ solution and the solid fraction precipitated, in order to evaluate the evolution of the isotopic ratios of carbon and oxygen with time.

The first trial's results are reported in Figure S8a in the Supplementary Materials: both the solid phase and the solution show a general ¹³C enrichment trend with time, but some values obtained for the solution are clearly affected by artifacts. In general, the lightest carbon isotope is removed from the gaseous phase dissolved in solution at the early stages of the reaction. The average partition coefficient α calculated as reported by Hayes [104] between products (calcium oxalate) and reagents (carbon dioxide in solution) equals 0.41 (±0.04), confirming the bias of the lightest carbon isotope (coming from the CO₂ canister) for the solid phase. However, isotopic data from the solid phase as collected in the first run are deceitful because oxalate crystals precipitate in many generations, as we described in our previous papers, and the isotopic zoning of crystals may be common. Both phenomena average, over time, the isotopic content of the solid fraction. Moreover, carbon contamination from the air during the transfer of the solution from the reactor to the flasks must be considered, meaning that the system is open compared to our standard experiment.

In the second experiment, we avoided carbon contamination from the air by keeping the system closed until the end of the experiment. Thus, we were not able to evaluate the evolution of the isotopic composition of the solid phase as a function of time and it was averaged over the whole experiment duration ($d^{13}C = -16.95$, Figure S8b in the Supplementary Materials). The partition coefficient α of ¹³C between products and reagents in this second run is equal to 0.5 (evaluated over the end-of-experiment point). Even if this is higher in value compared to the previous result, the bias of the lighter carbon isotope for the solid fraction is confirmed.

Data related to $d^{13}C$ in CO₂ dissolved in the capturing solution showed a first-order decay trend reported both in Figure S8b (Supplementary Materials) and Figure 2. Comparing the data coming from the first and second run, the calcium oxalate average $d^{13}C$ ratio ranges over a narrow interval of values between -12 and -17. The gas phase in solution is progressively enriched in ¹³C with time, as confirmed by the α values.

Going back to the general behavior of the capture curve (Figure 1), we can finally describe it as a double-sloped capture curve in which the first part (in the light-grey field in Figure 3), with a steeper slope that suggests an efficient carbon capture process, corresponds to the pure capture mechanism in which Ca[HA]₂ acts only as the reductant for C(IV) in C(III) and the concentration of the degradation products into the solution is low, as demonstrated by the CV and LC measurements. CO₂ reduces into the oxalate anion and forms calcium oxalate at various degrees of hydration [56,105].

The second part of the curve (the dark-grey shadowed field in Figure 3) represents a less efficient CO_2 capture process in the presence of a double oxalate source: (i) CO_2 reduction into $C_2O_4^{2-}$ by the action of the H₂A and (ii) degradation of the H₂A into its waste products and finally into the oxalic acid.



Figure 2. Trend of $d^{13}C$ measured in the mineralizing system (solid diamond-, liquid circles-, and gas squares phases) vs. time. No handling in the second run allowed a better quality of data, highlighting the first-order decay of $d^{13}C$ in the dissolved CO₂. The points related to the $d^{13}C$ of the CO₂ from the canister and H₂A (squares) are reported to be compared with the trend of the dissolved carbon. A single value for the solid fraction is reported (diamond) because of the experimental procedure intended to limit the artifacts' appearances in the measurements as explained in the text.



Figure 3. The general carbon capture curve via carbon mineralization into oxalates explained: in the light-grey area, the efficient pure carbon capture process during the "low-oxidation stage" of the H_2A as determined by CV and LC measurements; in the dark-grey area, the mixed process of capture and H_2A degradation occurring when the H_2A reaches higher degree of oxidation (30–40% from CV measurements) and the H_2A degradation cascade is running.

The increase in oxalic acid's concentration due to the presence of degradation products of H_2A in the second part of the capture curve leads to an abrupt rise in calcium oxalate concentration in the solution. At supersaturation, calcium oxalate (ksp_{COM} = 2.32×10^{-9} [101]) homogeneously nucleates in the volume of the solution. The formation of the early nuclei lowers the supersaturation, and the precipitation continues by heterogeneous nucleation on the homoepitaxial substrate, explaining the absence of visible precipitate during the early stages of mineralization experiments and in CV experiments.

The crystalline product of the mineralization reaction is pure weddellite. Figure 4 reports the PDXR pattern of the solid phase obtained from a standard carbon capture experiment (blue pattern). The calculated pattern of diffraction of weddellite is reported in red. The reference crystal structure is that of weddellite published in 1980 by Tazzoli and Domeneghetti [105] (tetragonal I 4/m, a = 12.3710 Å, c = 7.3570 Å). The perfect match between the experimental pattern and the calculated pattern is remarkable. Occasionally, some whewellite (the monohydrate form of calcium oxalate) is found as a byproduct, as reported in our early studies [57]. For more details about the crystal quality and structure stability, please refer to the literature [56].



Figure 4. Calcium oxalate is the only product of the mineralization reaction. (**a**) Calcium oxalate crystals from a B-setup carbon mineralization experiment; (**b**) XRPD pattern of the precipitate (red bars: weddellite pattern from Tazzoli and Domeneghetti, 1980 [105]).

Nevertheless, the final capture efficiency is not hindered by the double oxalate source, as was demonstrated in our previous work. Moreover, in all the capture experiments performed to verify the d¹³C, the final pressure into the vessel was lower than the room pressure, as occurred in the yield evaluation experiments [55], proving that carbon dioxide was efficiently removed from the atmosphere. Finally, from the LC experiments, we can infer that the carbon capture process will last for up to two days of mineralization.

4. Conclusions

In our previous work [55,57], the performance of the carbon capture reaction using H_2A as the electron donor was systematically examined to optimize the experimental setup towards a sizeable apparatus. The goal was to tailor the capture process parameters to obtain the maximum yield of carbon capture and marketable Ca-oxalate with specific crystallographic and morphologic features.

In the present paper, some key points of the reaction that we proposed, which were still not completely understood, have been addressed:

(i) In our previous work, we already described the process as a double-step reaction whose kinetics are controlled by the first (carbon reduction) step. Here, we confirmed that at the first reaction (Equation (1)) stage, H₂A oxidizes into DHA, and in the

counterpart, C(IV) reduces to C(III). In the present paper, we were able to assess, using CV measurements, the onset of the oxidation of H_2A at 0.15 V, while the oxidation potential of H_2A was indirectly evaluated in the range from 0.44 to 0.55 V, in agreement with former experimental data and theoretical modelling. These values demonstrate the spontaneousness of the carbon reduction reaction. On the other hand, the small redox potential gap between DHA oxidation and C reduction results in the slow triggering of the reaction, which is confirmed to be the rate-determining step. During the second stage of the reaction, the fastest step, atmospheric carbon that has been dissolved and reduced enters the crystal structure of oxalates along with the oxalic acid produced at the end of the degradation cascade of DHA. Stable isotope analysis provides d¹³C values in the oxalate crystal phase comparable to those of carbon from the CO_2 cylinder, confirming that this reaction phase is a "pure carbon capture" phase. Progressive ¹³C isotope enrichment of the solution reflects the bias of the heavy isotope of carbon for the liquid fraction as evaluated by d¹³C trend curves in the liquid and fractionation calculations (a partition coefficient of $\alpha \sim 0.5$ in the closed system).

(ii) In the simplified B-system, the curve describing the carbon capture performance invariably shows a double slope. The isotopic analysis explains this behavior and demonstrates the proneness of ¹²C from a canister to be preferentially trapped into the solid phase (and, in contrast, of ¹³C to be accumulated into the liquid phase). The carbon capture curve is steep at the beginning, during the "pure carbon capture phase" and flattens afterwards, when carbon coming from both CO_2 (reduced carbon) and ascorbic acid (finally degraded into oxalate) are stored in calcium oxalate crystals. From this point on, the reaction still works, but the carbon capture rate decreases because of the mixed carbon source for oxalates.

Oxalates, in turn, are crystalline reservoirs for long-term carbon dioxide storage and carbon raw materials for many manufacturing applications. Their structure and thermal stability are well known and are characterized [56] to estimate their reuse or storage behaviors. The presence of a substrate (understandably, a homoepitaxial substrate will be the best scenario and may be envisaged in future applications) assists their precipitation during mineralization reactions.

(iii) As expected, the degradation of H_2A into DHA and then into the products belonging to its degradation cascade, already described in our previously published work [57], is completely irreversible in abiotic conditions, when enzymatic processes are not present. CV data confirmed the irreversibility of the process during the carbon capture reaction. Therefore, reusing the H₂A after the carbon mineralization reaction is not possible once the lactone ring is opened. Nevertheless, we must recall from our previous studies [55,57] that the performance of the carbon mineralization was strongly dependent on the reaction surface and can reach levels up to 82% of the carbon dioxide in a 100% CO₂ atmosphere in less than 24 h, when working with a finely divided solution dispersed into a carbon dioxide atmosphere. This is a relevant yield, and is even more impressive if one considers that CO₂ in oxalates is permanently stored, making the methodology an interesting and valuable addition to the suite of available carbon capture techniques. We do not need to mention that the IPCC [1] has advocated for a portfolio of carbon capture techniques in its most recent publications. Operational costs can nevertheless be significantly reduced by properly tuning the concentration of the ascorbic acid to fulfill the application's requirements: carbon capture by means of ascorbic acid has been hitherto performed in a pure carbon dioxide atmosphere, far from the real-case scenarios, wherein the upper limit of CO₂ in a gas mixture falls to about 20%. Therefore, the concentration of ascorbic acid can be tuned to match the expected yield of carbon capture, cutting wastage without affecting performance. Moreover, applying H_2A to wastewater-containing metals suggests several appealing implications, such as the removal of metals from wastewaters.

Finally, we must stress that the irreversible process of delactonization could be possibly hindered by applying mixtures of reducing sugars [106]. This point will be addressed in further experiments.

Supplementary Materials: The following supporting information can be downloaded at: https: //www.mdpi.com/article/10.3390/cleantechnol6040066/s1, Figure S1: The B-setup. Carbon dioxide bubbles into the calcium ascorbate solution. The pressure setpoint is controlled using a pressure meter with a feedback loop on the flow meter. and it is chosen to be slightly higher than the atmospheric pressure. After loading, the system is closed and left to react. Figure S2: Flow rate vs. net revenue at variable reaction yield. The model is built on the yields experimentally determined in our previous papers from laboratory setups [55,57]. Yields 5% and 45% are not experimentally determined and represent a hypothetical very low carbon capture performance (comparable to the average performance of the B-setup) and a cautionary average performance of a BD/SPRAY setup, respectively. From the model, we can infer that the method is economically affordable for yields larger than 10% and CO₂ feeding equal or higher to 20 mL/min for a diluted Ca[HA]₂ solution (200 mL, 0.1 M). Table S1: Deprotonation and oxidation reactions following the model proposed by Tu et al. [74] (see also Supporting Information Figure S2 for visual description of the reactions). pKa and E0 values are reported as in the literature: a [75], b [76], c [77], d [78], e [79]. Figure S3: Scheme of the electrochemical equilibria of ascorbic acid and its derivatives in water solution. From Tu et al., modified [3]. Table S2: The list of the metals with tabulated values of the redox potentials used to roughly evaluate the redox potential of a 1 M aqueous ascorbic acid solution. Figure S4: (a) Calibration curve for H₂A; calculated LOD and LOQ were, respectively, 0.48 and 1.6 ppm; (b) Calibration curve for DHA; calculated LOD and LOQ were, respectively, 0.51 and 1.7 ppm. Figure S5: Cu0 crystals (single euhedral crystals and spherulitic aggregates) from the spontaneous redox reaction of CuO with H₂A. Figure S6: Cyclic voltammogram at 50 mV/s scan rate of 1.0 M AA, 0.30 M CaCl₂ solution at pH 4.2 under a CO_2 atmosphere. Figure S7: LC evaluation of the H_2A to DHA oxidation reaction: (a) H_2A concentration as a function of time; (b) DHA concentration as a function of time. Table S3: $d^{13}C$ and $d^{18}O$ ratios in the commercial CO₂ canister and the H₂A reagent. Figure S8: (a) General 13 C enrichment trend for both the solid phase and the solution. Some artifacts affect values from the solution. The general trend is an enrichment of the ¹²C in the gas fraction. Many crystal generations and zoning of crystals bring artifacts into the isotopic content of the solid phase. (b) In the second experiment, we avoided the contamination from the air that could arise from the transfer of the solution from the reactor to the flasks. No sampling of the solid phase was performed until the end of the experiment. The single solid composition is averaged over the whole experiment time $(d^{13}C = -16.95).$

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