

Prognostic relevance of cell proliferation in head and neck tumors

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Cell proliferative activity has been extensively investigated in head and neck tumors. Ki67/MIB-1 immunostaining, tritiated thymidine or bromodeoxyuridine labeling indices, DNA S-phase fraction, proliferating cell nuclear antigen expression, potential doubling time and analysis of the nucleolar organizer region associated proteins (AgNORs) have shown significant correlation with prognosis in 4806 cases of tumors of the oral cavity, salivary glands, pharynx and larynx. However, this was not observed in 2968 other reported cases. Discrepancies may depend on various factors: the heterogeneity of the series, which include tumors from various anatomic sites and patients treated with different therapy, and the lack of standardization of methods for assessing cell proliferation. Furthermore, none of the methods currently applied can by themselves define the actual proliferative activity, as it depends both on the proportion of cells committed to the cycle (growth fraction) and the speed of the cell cycle. Indeed, the actual proliferative activity of a tumor could well be measured by the equation [$PA = Ki67 \text{ or MIB-1 scores} \times AgNORs$], as we did in pharyngeal carcinoma. Provided that large and homogeneous series are evaluated by standardized methods, cell proliferative activity can still be regarded as an inexpensive and reliable prognostic factor in head and neck tumors.

Key words: cell proliferative activity, head and neck tumors, prognosis

Introduction

Cell proliferation is regarded as one of the most important biological mechanisms in oncogenesis [1]. A survey of the results of a large number of studies has shown that proliferative activity is of high prognostic significance in several types of cancer [2]. A Medline-based search (PubMed of the National Library of Medicine, via Internet) up to July 2003 selected 6305 reports containing the terms 'proliferative activity and tumor'; 4373 reports containing the terms 'proliferative activity' and 'tumor diagnosis', and 1005 containing the terms 'proliferative activity' and 'tumor prognosis'.

Cell proliferation has also been extensively investigated in head and neck tumors: up to July 2003, 306 papers have been published on the proliferative activity in tumors of the oral cavity, salivary glands, pharynx and larynx.

The aim of this review is to discuss and summarize the results obtained so far as to the proliferative activity of head and neck tumors, in an effort to verify if cell proliferation can

really provide useful prognostic information in these tumors or not.

Proliferation markers in tumor pathology

Proliferation markers can be classified into three main categories: (i) growth fraction markers; (ii) markers of specific phases of the cell cycle; and (iii) cell cycle time markers. The growth fraction, i.e. the proportion of the cells committed to the cycle, may be easily assessed by Ki-67 or MIB-1 antibodies, which identify an antigen expressed in G₁, S and G₂ phases of cycling cells [3, 4]. The M-phase can be evaluated by counting the mitotic figures: this is the oldest and, probably, the most popular way of assessing proliferation, even if strict morphological criteria for the recognition of mitotic figures are required [5]. S-phase fraction (SPF) can be assessed by incorporation techniques, such as the *in vivo* or *in vitro* incorporation with tritiated thymidine (TH3) or bromodeoxyuridine (BrdU), which can be regarded as the 'gold standard' marker of S-phase cells [1]. SPF can also be detected by static or flow cytometry (FCM) analysis of the DNA, or the immunohistochemical detection of proliferating cell nuclear antigen (PCNA/Cyclin), a nuclear protein involved in DNA synthesis [6]. The very reliable punctuated labeling of PCNA is identical to the labeling pattern obtained with BrdU [7] and is the method of choice of evaluating the S-phase index in histopathology [8]. Cell cycle time can be

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Table 1. Cell proliferative activity in oral cavity carcinoma

Parameter	Reference	No. of cases	Response
Prognostic value			
Ki67/MIB-1 immunostaining	Girod et al. [41]	216	Survival
	Matsumoto et al. [42]	20	N
	Sittel et al. [34]	56	DFS
	Xie et al. [43] ^a	85	DFS
PCNA immunostaining	Storkel et al. [44] ^a	100	Grade, survival ^b
	Alcalde et al. [45]	33	T, N
	Gasparini et al. [36] ^a	73	OS ^b , DFS
	Tsai and Jin [46]	38	T
	Daniele et al. [47]	54	N
	Welkoborsky et al. [48] ^a	42	OS ^b , DFS
	Sakurai et al. [49]	36	T, N, OS
TH3/BrdU-LI/T _{pot}	Chauvel et al. [50] ^a	87	OS ^b
	Hemmer [51]	33	T, N
	Mukhopadhyay et al. [52]	27	Grade, T, N
	Veneroni et al. [14] ^a	69	OS ^b
	Corvò et al. [33]	35	DFS
DNA-SPF	Gomez et al. [53] ^a	38	OS ^b
	Oya and Ikemura [54] ^a	79	N ^b
AgNORs	Sano et al. [55]	39	OS
	Hirsch et al. [56]	66	Stage
	Teixeira et al. [57] ^a	43	DFS ^b
	Chatterjee et al. [58]	39	Grade, T
	Piffko et al. [15] ^a	80	OS ^b , DFS ^b
	Piffko et al. [59] ^a	94	Metastasis ^b , OS ^b
	Xie et al. [60] ^a	51	OS ^b , DFS ^b
	Xie et al. [61] ^a	80	DFS ^b
Total		1613	
No prognostic value			
Ki67/MIB-1 immunostaining	Roland et al. [23] ^a	79	OS, N
	Valente et al. [28] ^a	31	DFS
	Gonzalez-Moles et al. [62]	74	OS
	Piffko et al. [63] ^a	100	OS
	Stoll et al. [64]	107	OS, DFS
	Bettendorf and Herrmann [65]	329	T, N, OS
	Carinci et al. [66] ^a	25	OS
Okamoto et al. [67]	59	N	
PCNA immunostaining	Sommer and Olofsson [68]	64	T, N, DFS, OS
	Sittel et al. [34]	56	DFS
TH3/BrdU-LI/T _{pot}	Cooke et al. [22]	105	T, N, OS
	Costa et al. [35] ^a	100	DFS
DNA-SPF	Mohr et al. [20]	142	OS
Total		1271	

^aStudies with multivariate analysis.

^bStudies in which the proliferative activity has prognostic significance in multivariate analysis.

DFS, disease-free survival; N, N stage; OS, overall survival; T, T stage.

evaluated by the potential doubling time (T_{pot}), a procedure that requires *in vivo* intravenous BrdU infusion and bivariate FCM [9], or by the quantification of the argyrophilic proteins associated with the nucleolar organizer regions (AgNORs), loops of DNA which transcribe to ribosomal RNA [10]. AgNOR proteins can be easily detected on routinely fixed and paraffin embedded tissues [11]. The AgNOR quantity is strictly related to the rapidity of cell proliferation: the higher the AgNOR quantity, the shorter the doubling time [12, 13].

Oral cavity carcinoma

A large number of papers, reporting a total of 1613 cases, have shown the prognostic value of cell proliferation in squamous cell carcinoma (SCC) of the oral cavity. In general, a high proliferative activity is associated with poor prognosis, even if one series of 69 oral/oropharyngeal SCC patients, who were treated with surgery alone, reported a high TH3-LI

(where LI is labeling index) associated with a better prognosis [14]. Almost 30% of the cases (492 of 1613) were investigated using AgNOR analysis, even if this method of assessing cell proliferation is not extensively applied to tumor pathology. Of particular interest are the results obtained by Piffko et al. [15] in 80 SCC patients, where the quantity of AgNORs at the invasive front of the tumor was reported to be the most significant independent prognostic factor. This finding not only provides a functional background to the well-established clinical relevance of the histopathological tumor front grading [16], but also suggests the use of an aggressive surgical approach for those patients with a high AgNOR quantity. However, in a large number of papers, reporting a total 1271 cases, cell proliferative activity in oral SCC lacks any prognostic value. Literature reports are summarized in Table 1.

Curiously, we noted no paper reporting a lack of prognostic significance for AgNOR analysis. This might well imply that

Table 2. Cell proliferative activity in salivary gland tumors

Parameter	Reference	No. of cases	Response
Prognostic value			
Ki67/MIB-1 immunostaining	Skalova et al. [69]	33	DFS, OS, grade
	Skalova et al. [70]	30	Recurrence
	Hellquist et al. [71]	32	OS
	Nordgard et al. [72] ^a	44	DFS ^b
	Yin et al. [73]	71	OS, grade
	Okabe et al. [74] ^a	31	OS, grade
	Xin and Paulino [75]	18	Grade, metastasis
PCNA immunostaining	Batsakis [40] ^a	43	OS ^b
	Cho et al. [76]	30	OS, DFS
DNA-SPF	Greiner et al. [77]	45	OS
	Hicks et al. [78] ^a	48	OS ^b
	Franzen et al. [79] ^a	51	DFS ^b
	Pinto et al. [80]	26	Grade
	Junquera et al. [81]	44	T, DFS
AgNORs	Chomette et al. [82]	17	OS, DFS
	Chomette et al. [83]	31	OS, DFS
	Vuhahula et al. [84]	34	Metastasis
	Xie et al. [85] ^a	37	DFS ^b
Total		665	
No prognostic value			
PCNA immunostaining	Felix et al. [86] ^a	30	OS
DNA-SPF	Timon et al. [87] ^a	45	Recurrence, N, OS
	Felix et al. [86] ^a	30	OS
AgNORs	Fonseca and Soares [88]	18	Metastasis, OS
	Timon et al. [87] ^a	45	Recurrence, N, OS
Total		168	

^aStudies with multivariate analysis.

^bStudies in which the proliferative activity has prognostic significance in multivariate analysis.

DFS, disease-free survival; N, N stage; OS, overall survival; T, T stage.

the rapidity of cell proliferation, as assessed by AgNOR quantification, is probably the kinetic parameter which correlates better with the outcome of oral cavity SCC.

Salivary gland tumors

A lot of papers, reporting a total of 665 cases, have shown that high proliferative activity is associated with a worse prognosis in salivary gland tumors of various histological type. In particular, high AgNOR counts were indicators of poor overall survival (OS) and/or disease-free survival (DFS) in 119 cases from four different series. Three authors, however, reporting a total of 168 cases, did not confirm the prognostic value of cell proliferation in salivary gland tumors. A summary of the reports in the literature is shown in Table 2.

Pharyngeal carcinoma

To the best of our knowledge, literature reports few papers that have investigated the cell proliferative activity of pharyngeal carcinoma. Eight, for a total of 451 cases, show an association between cell proliferation and prognosis in pharyngeal carcinoma. However, five other authors, for a total of 317 cases, did not confirm the prognostic value of cell proliferation. The results of the literature are shown in Table 3.

Laryngeal carcinoma

Cell proliferation evaluation has been used most extensively in the investigation of laryngeal carcinomas, and a large number

of publications, reporting a total of 2077 cases, have shown its prognostic value. In general, high proliferative activity is associated with a worse prognosis, even if high Ki67 scores were associated with a favorable response to radiotherapy in one series of 36 T2 laryngeal cancers [17], and a low DNA-SPF was found to predict recurrence in 23 glottic carcinomas that achieved complete local remission after curative radiotherapy [18]. However, a few studies, reporting a total of 1212 cases, failed to demonstrate the prognostic value of cell proliferative activity in laryngeal carcinoma. Table 4 shows a summary of the literature reports.

Personal findings

A total of 94 cases of pharyngeal carcinoma were retrospectively investigated in our laboratory. Patient age ranged from 29 to 87 years (mean, 60.1 years) of which 68 were male and 26 female. Forty-two carcinomas were from the oro-rhinopharynx and 52 from hypopharynx-pyiform sinus. All cases were squamous cell carcinoma; according to UICC [19], 34 were grade 2 and 60 grade 3; 12 were stage T1, 25 T2, 36 T3 and 21 T4; 43 were N0, 18 N1, 5 N2 and 28 N3. Out of 42 oro-rhinopharyngeal carcinomas, 10 (23.8%) were stages 1–2 and 32 (76.2%) stages 3–4. Out of 52 hypopharynx-pyiform sinus carcinomas, seven (13.5%) were stages 1–2 and 45 (86.5%) stages 3–4. After diagnosis, all patients underwent only external ⁶⁰Co radiotherapy. A minimum follow-up of 3 years for censored (surviving) patients or until death was available in all cases. Initial biopsies, fixed in 10% formalin and embedded in paraffin, were stained with the AgNOR

Table 3. Cell proliferative activity in pharyngeal carcinoma

Parameter	Reference	No. of cases	Response
Prognostic value			
Ki67/MIB-1 immunostaining	Pich et al. [89] ^a	28	OS ^b
	Raybaud et al. [31] ^a	56	Recurrence ^b
PCNA immunostaining	Pich et al. [26]	45	OS
	TH3/BrdU-LI/T _{pot}	87	OS ^b
DNA-SPF	Corvò et al. [21]	31	Recurrence
	Benazzo et al. [90]	52	N
	Myers et al. [91]	91	OS, N, grade
AgNORs	Pich et al. [25] ^a	61	OS ^b
Total		451	
No prognostic value			
Ki67/MIB-1 immunostaining	Roland et al. [23] ^a	79	Metastasis, OS
	Shnayder et al. [92]	54	Recurrence OS
BrdU-LI/T _{pot}	Bourhis et al. [29] ^a	70	Recurrence T, N, DFS, OS
DNA-SPF	Del Valle-Zapico et al. [93] ^a	51	T, OS
AgNORs	Pak et al. [37] ^a	63	OS, DFS
Total		317	

^aStudies with multivariate analysis.

^bStudies in which the proliferative activity has prognostic significance in multivariate analysis.

DFS, disease-free survival; N, N stage; OS, overall survival; T, T stage.

method [11] and PCNA and MIB-1 immunostaining using PC10 and MIB-1 monoclonal antibodies.

In carcinomas of the oro-rhinopharynx, the median AgNOR count was 10.32 AgNORs/cell and the median PCNA score 36.12. No association was found between AgNOR counts or

PCNA scores and tumor histological grade or stage. The median OS for the whole series was 48 months, and the 3- and 5-year survival rates were 60% and 49%, respectively. Cell proliferation was the only significant prognostic factor: using the median values as the cut-off, 3-year survival rates

Table 4. Cell proliferative activity in laryngeal carcinoma

Parameter	Reference	No. of cases	Response	
Prognostic value				
Ki67/MIB-1 immunostaining	Hake et al. [94]	47	Recurrence, DFS	
	Welkoborsky et al. [95] ^a	40	Recurrence ^b	
	Franchi et al. [96] ^a	60	N ^b	
	Kropveld et al. [17]	36	Recurrence	
	Golusinski et al. [97]	120	T, N, grade	
	Valente et al. [98] ^a	102	DFS ^b	
	Sittel et al. [38]	47	Recurrence, DFS	
	Lazaris et al. [99] ^a	96	Recurrence, N ^b	
	Wozniak et al. [100]	55	T, N	
	PCNA immunostaining	Munck-Wikland et al. [27]	28	Recurrence
		Welkoborsky et al. [95] ^a	40	OS ^b
		Franchi et al. [96] ^a	60	N ^b
		Dobros et al. [101]	90	N, OS
		Krecicki and Jelen [102] ^a	154	OS ^b
Sarac et al. [103] ^a		92	Grade, N, DFS ^b	
Dong et al. [104] ^a		54	OS, DFS	
Sittel et al. [38]		47	Recurrence, DFS	
BrdU- T_{pot}	Dong et al. [105]	102	OS	
	Alsner et al. [32]	58	OS	
DNA-SPF	Tennvall et al. [18]	23	Recurrence	
	Tomasino et al. [106] ^a	99	OS ^b , DFS ^b	
	Danic et al. [107] ^a	36	OS ^b	
	Myers et al. [91]	91	OS, Grade, N	
	Dobros et al. [108] ^a	90	OS ^b	
	Bazan et al. [109] ^a	64	OS	
	AgNORs	Bockmuhl et al. [110] ^a	30	OS ^b , DFS ^b
Xie et al. [111] ^a		93	DFS ^b	
Xie et al. [112]		69	DFS	
Krecicki et al. [113] ^a		154	OS ^b	
Total		2077		
No prognostic value				
Ki67/MIB-1 immunostaining	Roland et al. [23] ^a	79	N, OS	
	Resnick et al. [24]	88	N, OS	
	Spafford et al. [114] ^a	70	Metastasis, OS	
	Hirvikoski et al. [115] ^a	103	DFS, OS	
	Pulkkinen et al. [116] ^a	100	OS	
	Krecicki et al. [117] ^a	154	T, N	
PCNA immunostaining	Resnick et al. [24]	88	N, OS	
	Tomasino et al. [106] ^a	99	OS, DFS	
	Hirvikoski et al. [115] ^a	103	DFS, OS	

Table 4. (Continued)

Parameter	Reference	No. of cases	Response
DNA-SPF	Resnick et al. [24]	88	N, OS
AgNORs	Yamamoto et al. [118]	73	OS
	Barnes et al. [119]	53	T, N, recurrence, OS
	Pulkkinen et al. [120]	66	DFS, OS
	Klatka and Skomra [121]	48	T, N, OS
Total		1212	

^aStudies with multivariate analysis.

^bStudies in which the proliferative activity has prognostic significance in multivariate analysis.

DFS, disease-free survival; N, N stage; OS, overall survival; T, T stage;

were 81% for patients with ≤ 10.32 AgNORs/cell but only 38% for those with higher counts ($P=0.0002$); 87% for patients with PC10 scores ≤ 36.12 , but only 40% for those with higher scores ($P=0.0002$). In Cox multivariate survival analysis, only PCNA appeared as an independent variable ($\chi^2=13.56$; $P<0.001$; $rr=8.16$). When PCNA was excluded, AgNOR count emerged as an independent variable ($\chi^2=13.79$; $P<0.001$; $rr=5.71$).

In carcinomas of the hypopharynx-pyiform sinus, the median AgNOR count was 11.27 AgNORs/cell and the median PCNA score 40. No association was found between AgNOR counts or PCNA scores and tumor histological grade or stage. The median MIB-1 score was 33.87 and was higher in G_3 than in G_2 carcinomas ($P=0.008$). The median OS for the whole series was 20 months, and the 3- and 5-year survival rates were 26% and 22%, respectively. Of the classical parameters, histological grade ($P=0.009$) and T stage ($P=0.04$) were associated with OS. Using the median values as a cut-off, the 3-year survival rates were 48% for patients with ≤ 11.27 AgNORs/cell, but only 4% for those with higher counts ($P<0.0001$); 71% for patients with PC10 scores ≤ 40 but only 14% for those with higher scores ($P=0.01$); 47% for patients with MIB-1 scores ≤ 33.87 and 0% for those with higher counts ($P<0.0001$). In Cox multivariate survival analysis, only MIB-1 scores ($\chi^2=18.53$; $P<0.001$; $rr=6.06$) and AgNOR counts ($\chi^2=3.89$; $P=0.04$; $rr=3.34$) appeared as independent prognostic variables.

Comments

Even if most of the papers published to date show a correlation between high proliferative activity and poor prognosis in head and neck tumors, a large number of reports failed to show any significant association with prognosis. There are several reasons which may account for this discrepancy.

Anatomic site

In the last edition of the TNM classification of malignant tumors [19], head and neck tumors include tumors of the lip, oral cavity, pharynx, larynx, maxillary sinus, nasal cavity and ethmoid sinus, salivary gland and thyroid gland. Therefore, the series which generically deal with tumors of the 'head and

neck' [20–23] are heterogeneous, mixing tumors of different histological type, biological activity and, consequently, clinical behavior. Our experience with pharyngeal carcinoma indicates that it is better not to consider pharyngeal tumors from different anatomic sites as a whole. In fact, the median OS was 23 months when the whole series of 94 cases was considered; however, when cases were subdivided according to the anatomic site, the median OS of the 42 oro-rhinopharyngeal carcinomas was 48 months, significantly longer than that of the 52 hypopharynx-pyiform sinus cases (20 months, $P=0.002$). The different survival rates were not dependent on differences in clinical stage, since the proportion of low and advanced cases was comparable in the two anatomic sites ($P=0.3$).

Type of therapy

Literature reports some series of head and neck tumors treated with surgery alone [14, 24], and others with radiotherapy alone [18, 21, 25–32]. Other series were treated with combined surgery and radiotherapy [33, 34], combined surgery and chemotherapy [35], or combined radio- and chemotherapy [36, 37], and there were also a few cases treated with laser therapy [38]. Since different therapies, especially chemo- or radiotherapy, interfere with biological tumor activities, only homogeneously treated series of head and neck carcinomas should be compared if the prognostic value of cell proliferation is to be assessed reliably. For instance, TH3-LI provided independent prognostic information in oral and oropharyngeal carcinomas treated with surgery alone [14], but did not appear to be a predictive factor in patients with carcinoma of the same anatomic site submitted to primary chemotherapy followed by surgery [35].

Standardization

There is a lack of standardization in the various methods for assessing cell proliferation. The number of mitotic figures can be evaluated in several ways: (i) the mitotic index (MI) (total number of mitoses in 10 high power fields); (ii) the mitotic rate (MR) (number of mitoses per 1000 cells); or (iii) the number of mitoses for the area percentage of epithelium, which yields the mitoses per volume (M/V) index [1]. Of course, the different procedures lead to different mitosis

counts. The use of various antibody dilutions, or antigen retrieval, can lead to different results in immunohistochemical staining e.g. most of our PC10 immunonegative pharyngeal carcinomas became positive after microwave irradiation. Moreover, different DNA indices and SPF values are obtained when fresh or fixed tissues are processed. Lastly, the positivity or negativity of a case may depend on the different cut-off values of the proliferation markers. For instance, we chose the median MIB-1 value (33.87) as a cut-off in our series of hypopharyngeal carcinomas and found 14 (50%) negative cases; however, only between one and three cases would have been negative if a cut-off of 15–20% (similar to the cut-off currently used for mammary carcinomas) had been applied.

Site of measurement

The site of measurement is another point, as heterogeneity is one of the hallmarks of malignancy, and heterogeneity for several proliferation markers has been documented for many types of tumors. A large tumor specimen may be classified positive or negative depending on whether the proliferation markers are evaluated only at the most ‘active’ sites (e.g. the invasive tumor front) [15], in randomly selected areas [25, 26] or in the maximally positive tumor field [17].

Identification of neoplastic cells

There is a difficulty in identifying the neoplastic cells. It is often hard to distinguish by MIB-1 or PCNA immunostaining small neoplastic cells from reactive proliferating lymphocytes in poorly differentiated rhinopharyngeal carcinomas. AgNOR staining is particularly suitable in such cases, since it allows for an easy distinction between neoplastic and non-neoplastic cells, avoiding the need for double (Cytokeratin/MIB-1) staining.

Proliferative activity

Finally, what was indeed measured in most studies reported is not the actual proliferative activity of a tumor, in as much as the mechanisms responsible for proliferative activity are the proportion of cells committed to cycle (growth fraction, or G) and the speed of the cycle, which is inversely proportional to the generation time (T) [1]. The mathematical relationship between proliferative activity (PA), growth fraction (G) and generation time (T) is $PA = G/T$ [1]. It is clear from this equation that neither the growth fraction nor the generation time can by itself define the actual proliferative activity of a tumor [39]. The tumor growth fraction can be easily assessed on routinely processed tissues by the Ki67 or MIB-1 labeling index, since Ki67 or MIB-1 antibodies recognize an antigen expressed in all cycling cells [3, 4]. On the contrary, evaluation of the speed of cell cycle by the potential doubling time (T_{pot}) is a rather lengthy procedure and is not suitable for retrospective studies. However, since the AgNOR quantity reflects the rapidity of cell proliferation [12, 13], and can be detected in routinely fixed and embedded tissues [11], AgNOR analysis can be regarded as an easy and reliable tech-

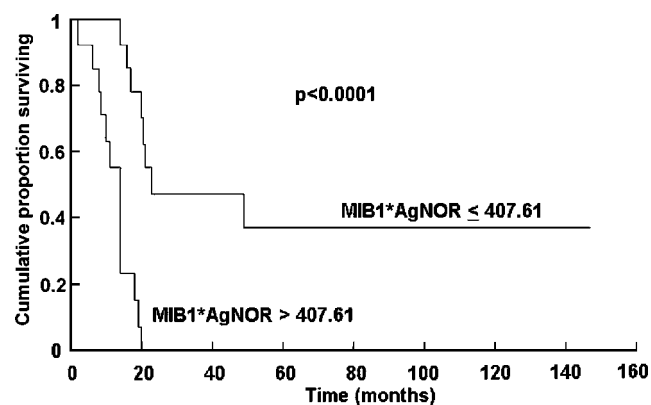


Figure 1. Kaplan–Meier survival curves (OS) for 28 hypopharynx-pyriform sinus carcinomas categorized according to the median (MIB-1 × AgNOR) value.

nique to evaluate the tumor cell doubling time on histological preparations. Therefore, the ‘actual’ proliferative activity of a tumor can be simply expressed by the equation ($PA = Ki67$ or $MIB-1$ scores × AgNOR quantity). To the best of our knowledge, this equation, which represents the ‘true’ proliferative activity of a tumor, has never been applied to head and neck tumors. We used this formula to assess cell proliferation in 28 carcinomas of the hypopharynx-pyriform sinus. The median OS for the whole series was 18 months, and the 3- and 5-year survival rates were 24% and 19%, respectively. Using the median (MIB-1 × AgNOR) value as a cut-off, the median survival for cases with (MIB-1 × AgNOR) ≤ 407.61 was 23 months, but only 11 months for those with higher values ($P < 0.0001$). Noteworthy, after 20 months of follow-up, 79% of patients with a low (MIB-1 × AgNOR) index were alive, whereas patients with higher values were not (Figure 1). We are aware that the number of our cases is rather small and this approach should be validated in a large series of patients.

Summary

Despite the many negative reports in the literature, cell proliferative activity is a reliable prognostic factor in head and neck tumors, especially when studies are performed with well standardized methods that take into account the type of treatment used. Indeed, a high cell proliferation, as expressed by the MIB-1 labeling index, was a significant indicator for treatment failure in a large matched-pair study design of recurrent and non-recurrent oral and oropharyngeal carcinomas initially treated with primary surgery combined with curative post-operative radiation [34]. Also, in another large matched-pair study on recurrent and non-recurrent laryngeal carcinomas, homogeneous for site (glottis), stage (T1 and T2) and treatment (transoral laser surgery alone), investigated using well standardized MIB-1 and PCNA staining and scoring, high proliferative activity appeared to be a significant prognostic factor. Lastly, the standardized AgNOR analysis showed the strong independent prognostic value of cell proliferation in a large series of oral squamous cell carcinoma [15].

An increasing number of sophisticated techniques, such as molecular biology, are currently being applied to tumor pathology to investigate the oncogene alterations responsible for cell proliferation changes. However, since the proliferation changes during the cell cycle reflect the net effect of genetic damage and, therefore, include the accumulated changes in genes, they may be regarded as more useful prognostic indicators than individual oncogene alterations [40].

In conclusion, provided that large series of cases, homogeneous for anatomic site and type of treatment, are evaluated by well standardized methods, cell proliferative activity still seems to be an inexpensive and reliable prognostic factor in head and neck tumors.

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References

- van Diest PJ, Brugal G, Baak JPA. Proliferation markers in tumours: interpretation and clinical value. *J Clin Pathol* 1998; 51: 716–724.
- Tubiana M, Courdi A. Cell proliferation kinetics in human solid tumors: relation to probability of metastatic dissemination and long term survival. *Radiother Oncol* 1989; 15: 1–18.
- Gerdes J, Schwab U, Lemke H, Stein H. Production of a monoclonal antibody reactive with a human nuclear antigen associated with cell proliferation. *Int J Cancer* 1983; 31: 13–20.
- Cattoretti G, Becker MHG, Key G et al. Monoclonal antibodies against recombinant parts of the Ki-67 antigen (MIB-1 and MIB-3) detect proliferating cells in microwave-processed formalin-fixed paraffin sections. *J Pathol* 1992; 168: 357–363.
- Baak JPA. Mitosis counting in tumours. *Hum Pathol* 1990; 21: 683–685.
- Mathews MB, Bernstein RM, Franza BR, Garrels JI. Identity of the proliferating cell nuclear antigen and cyclin. *Nature* 1984; 303: 374–376.
- Humbert C, Santisteban MS, Usson Y, Robert-Nicoud M. Intranuclear co-location of newly replicated DNA and PCNA by simultaneous immunofluorescent labelling and confocal microscopy in MCF-7 cells. *J Cell Sci* 1992; 103: 97–103.
- Galand P, Degraef C. Cyclin/PCNA immunostaining as an alternative to tritiated thymidine pulse labelling for marking S phase cells in paraffin sections from animal and human tissues. *Cell Tissue Kinet* 1989; 22: 383–392.
- Riccardi A, Danova M, Wilson G et al. Cell kinetics in human malignancies studied with *in vivo* administration of bromodeoxyuridine and flow cytometry. *Cancer Res* 1988; 48: 6238–6245.
- Gall JG, Pardue ML. Formation and detection of RNA-DNA hybrid molecules in cytological preparations. *Proc Natl Acad Sci USA* 1969; 63: 378–383.
- Ploton D, Menager M, Jeannesson P et al. Improvement in the staining and in the visualisation of the argyrophilic proteins of the nucleolar organizer region at the optical level. *Histochem J* 1986; 18: 5–14.
- Derenzini M, Sirri V, Trere D, Ochs RL. The quantity of nucleolar proteins nucleolin and protein B23 is related to cell doubling time in human cancer cells. *Lab Invest* 1995; 73: 497–502.
- Dong H, Bertler C, Schneider E, Ritter MA. Assessment of cell proliferation by AgNOR scores and Ki-67 labeling indices and a comparison with potential doubling times. *Cytometry* 1997; 28: 280–288.
- Veneroni S, Silvestrini R, Costa A et al. Biological indicators of survival in patients treated by surgery for squamous cell carcinoma of the oral cavity and oropharynx. *Oral Oncol* 1997; 33: 408–413.
- Piffko J, Bankfalvi A, Öfner D et al. Standardized AgNOR analysis of the invasive tumour front in oral squamous cell carcinomas. *J Pathol* 1997; 182: 450–456.
- Bryne M, Koppang HS, Lilleng R, Kjaerheim A. Malignancy grading of the deep invasive margins of oral squamous cell carcinomas has high prognostic value. *J Pathol* 1992; 166: 375–381.
- Kropveld A, Slootweg PJ, Blankenstein MA et al. Ki-67 and p53 in T2 laryngeal cancer. *Laryngoscope* 1998; 108: 1548–1552.
- Tennvall J, Wennerberg J, Willen R et al. T3N0 glottic carcinoma: DNA S-phase as a predictor of the outcome after radiotherapy. *Acta Otolaryngol* 1993; 113: 220–224.
- Sobin LH, Wittekind CH. *TNM Classification of Malignant Tumors*, 6th edition. New York, NY: Wiley-Liss 2002.
- Mohr C, Molls M, Streffer C, Pelzer T. Prospective flow cytometric analysis of head and neck carcinomas. Prognostic relevance of DNA-content and S-fraction. *J Craniomaxillofac Surg* 1992; 20: 8–13.
- Corvò R, Giaretti W, Sanguineti G et al. Potential doubling time in head and neck tumors treated by primary radiotherapy: preliminary evidence for a prognostic significance in local control. *Int J Radiat Oncol Biol Phys* 1993; 27: 1165–1172.
- Cooke LD, Cooke TG, Forster G et al. Prospective evaluation of cell kinetics in head and neck squamous carcinoma: the relationship to tumour factors and survival. *Br J Cancer* 1994; 69: 717–720.
- Roland NJ, Caslin AW, Bowie GL, Jones AS. Has the cellular proliferation marker Ki67 any clinical relevance in squamous cell carcinoma of the head and neck? *Clin Otolaryngol* 1994; 19: 13–18.
- Resnick JM, Uhlman D, Niehans GA et al. Cervical lymph node status and survival in laryngeal carcinoma: prognostic factors. *Ann Otol Rhinol Laryngol* 1995; 104: 685–694.
- Pich A, Pisani P, Krengli M et al. Argyrophilic nucleolar organizer region counts and prognosis in pharyngeal carcinoma. *Br J Cancer* 1991; 64: 327–332.
- Pich A, Chiusa L, Pisani P et al. Argyrophilic nucleolar organizer region counts and proliferating cell nuclear antigen scores are two reliable indicators of survival in pharyngeal carcinoma. *J Cancer Res Clin Oncol* 1992; 119: 106–110.
- Munck-Wikland E, Fernberg JO, Kuylensstierna R et al. Proliferating cell nuclear antigen (PCNA) expression and nuclear DNA content in predicting recurrence after radiotherapy of early glottic cancer. *Eur J Cancer B Oral Oncol* 1993; 29B: 75–79.
- Valente G, Orecchia R, Gandolfo S et al. Can Ki67 immunostaining predict response to radiotherapy in oral squamous cell carcinoma? *J Clin Pathol* 1994; 47: 109–112.
- Bourhis J, Dendale R, Hill C et al. Potential doubling time and clinical outcome in head and neck squamous cell carcinoma treated with 70 Gy in 7 weeks. *Int J Radiat Oncol Biol Phys* 1996; 35: 471–476.
- Toffoli G, Franchin G, Barzan L et al. Brief report: prognostic importance of cellular DNA content in T1-2 N0 laryngeal squamous cell carcinomas treated with radiotherapy. *Laryngoscope* 1995; 105: 649–652.
- Raybaud H, Fortin A, Bairati I et al. Nuclear DNA content, an adjunct to p53 and Ki-67 as a marker of resistance to radiation therapy in oral cavity and pharyngeal squamous cell carcinoma. *Int J Oral Maxillofac Surg* 2000; 29: 36–41.
- Alsner J, Hoyer M, Sorensen SB, Overgaard J. Interaction between potential doubling time and TP53 mutation: predicting radiotherapy

- outcome in squamous cell carcinoma of the head and neck. *Int J Radiat Oncol Biol Phys* 2001; 49: 519–525.
33. Corvò R, Margarino G, Sanguineti G et al. Cell kinetics analysis in patients affected by squamous cell carcinoma of the head and neck treated with primary surgery and adjuvant radiotherapy. *Tumori* 2000; 86: 53–58.
 34. Sittel C, Ruiz S, Volling P et al. Prognostic significance of Ki-67 (MIB 1), PCNA and p53 in cancer of the oropharynx and oral cavity. *Oral Oncol* 1999; 35: 583–589.
 35. Costa A, Licitra L, Veneroni S et al. Biological markers as indicators of pathological response to primary chemotherapy in oral-cavity cancers. *Int J Cancer* 1998; 79: 619–623.
 36. Gasparini G, Bevilacqua P, Bonoldi E et al. Predictive and prognostic markers in a series of patients with head and neck squamous cell invasive carcinoma treated with concurrent chemoradiation therapy. *Clin Cancer Res* 1995; 1: 1375–1383.
 37. Pak MW, To KF, Leung SF, van Hasselt CA. Prognostic significance of argyrophilic nucleolar organizer regions in nasopharyngeal carcinoma. *Eur Arch Otorhinolaryngol* 2000; 257: 517–520.
 38. Sittel C, Eckel HE, Damm M et al. Ki-67 (MIB1), p53, and Lewis-X (LeuM1) as prognostic factors of recurrence in T1 and T2 laryngeal carcinoma. *Laryngoscope* 2000; 110: 1012–1017.
 39. Brugal G. Interpretation of proliferation markers. In Hofstädter F, Knüchel R, Rüschoff J (eds): *Cell Proliferation Assessment in Oncology*. Virchows Arch 1995; 427: 323–341.
 40. Batsakis JG. Staging of salivary gland neoplasms: role of histopathologic and molecular factors. *Am J Surg* 1994; 168: 386–390.
 41. Girod SC, Pfeiffer P, Ries J, Pape HD. Proliferative activity and loss of function of tumour suppressor genes as ‘biomarkers’ in diagnosis and prognosis of benign and preneoplastic oral lesions and oral squamous cell carcinoma. *Br J Oral Maxillofac Surg* 1998; 36: 252–260.
 42. Matsumoto M, Komiyama K, Okae M et al. Predicting tumor metastasis in patients with oral cancer by means of the proliferation marker KI67. *J Oral Sci* 1999; 41: 53–56.
 43. Xie X, De Angelis P, Clausen OP, Boysen M. Prognostic significance of proliferative and apoptotic markers in oral tongue squamous cell carcinomas. *Oral Oncol* 1999; 35: 502–509.
 44. Storkel S, Reichert T, Reiffen KA, Wagner W. EGFR and PCNA expression in oral squamous cell carcinomas—a valuable tool in estimating the patient’s prognosis. *Eur J Cancer B Oral Oncol* 1993; 29B: 273–277.
 45. Alcalde RE, Shintani S, Yoshihama Y, Matsumura T. Cell proliferation and tumor angiogenesis in oral squamous cell carcinoma. *Anticancer Res* 1995; 15: 1417–1422.
 46. Tsai ST, Jin YT. Proliferating cell nuclear antigen (PCNA) expression in oral squamous cell carcinomas. *J Oral Pathol Med* 1995; 24: 313–315.
 47. Daniele E, Rodolico V, Leonardi V, Tralongo V. Prognosis in lower lip squamous cell carcinoma: assessment of tumor factors. *Pathol Res Pract* 1998; 194: 319–324.
 48. Welkoborsky HJ, Gluckman JL, Jacob R et al. Tumor biologic prognostic parameters in T1N0M0 squamous cell carcinoma of the oral cavity. *Laryngorhinootologie* 1999; 78: 131–138.
 49. Sakurai K, Urade M, Takahashi Y et al. Increased expression of c-erbB-3 protein and proliferating cell nuclear antigen during development of verrucous carcinoma of the oral mucosa. *Cancer* 2000; 89: 2597–2605.
 50. Chauvel P, Courdi A, Gioanni J et al. The labelling index: a prognostic factor in head and neck carcinoma. *Radiother Oncol* 1989; 14: 231–237.
 51. Hemmer J. In vitro bromodeoxyuridine labelling of squamous cell carcinomas of the oral cavity. *Eur J Cancer* 1990; 26: 113–115.
 52. Mukhopadhyay D, Chatterjee R, Chakraborty RN. Cytokinetic studies of oral cancer cells using bromodeoxyuridine labelling in relation to factors influencing prognosis. *Eur J Cancer B Oral Oncol* 1995; 31B: 32–36.
 53. Gomez R, el-Naggar AK, Byers RM et al. Squamous carcinoma of oral tongue: prognostic significance of flow-cytometric DNA content. *Mod Pathol* 1992; 5: 141–145.
 54. Oya R, Ikemura K. Can flow cytometrically determined DNA ploidy and S-phase fraction predict regional metastasis in squamous cell carcinoma of the oral cavity? *Head Neck* 2002; 24: 136–142.
 55. Sano K, Takahashi H, Fujita S et al. Prognostic implication of silver-binding nucleolar organizer regions (AgNORs) in oral squamous cell carcinoma. *J Oral Pathol Med* 1991; 20: 53–56.
 56. Hirsch SM, DuCanto J, Caldarelli DD et al. Nucleolar organizer regions in squamous cell carcinoma of the head and neck. *Laryngoscope* 1992; 102: 39–44.
 57. Teixeira G, Antonangelo L, Kowalski L et al. Argyrophilic nucleolar organizer regions staining is useful in predicting recurrence-free interval in oral tongue and floor of mouth squamous cell carcinoma. *Am J Surg* 1996; 172: 684–688.
 58. Chatterjee R, Mukhopadhyay D, Chakraborty RN, Mitra RB. Evaluation of argyrophilic nucleolar organizer regions (AgNORs) in oral carcinomas in relation to human papillomavirus infection and cytogenetics. *J Oral Pathol Med* 1997; 26: 310–314.
 59. Piffko J, Bankfalvi A, Öfner D et al. Prognostic value of histobiological factors (malignancy grading and AgNOR content) assessed at the invasive tumour front of oral squamous cell carcinomas. *Br J Cancer* 1997; 75: 1543–1546.
 60. Xie X, Clausen OP, Sudbo J, Boysen M. Diagnostic and prognostic value of nucleolar organizer regions in normal epithelium, dysplasia, and squamous cell carcinoma of the oral cavity. *Cancer* 1997; 79: 2200–2208.
 61. Xie X, Boysen M, Clausen OP, Bryne MA. Prognostic value of Le(y) and H antigens in oral tongue carcinomas. *Laryngoscope* 1999; 109: 1474–1480.
 62. Gonzalez-Moles MA, Caballero R, Rodriguez-Archilla A et al. Prognosis value of the expression of Ki-67 for squamous cell carcinoma of the oral cavity. *Acta Stomatol Belg* 1996; 93: 159–165.
 63. Piffko J, Bankfalvi A, Öfner D et al. In situ assessment of cell proliferation at the invasive front of oral squamous cell carcinomas. *Virchows Arch* 1996; 429: 229–234.
 64. Stoll C, Baretton G, Ahrens C, Lohrs U. Prognostic significance of apoptosis and associated factors in oral squamous cell carcinoma. *Virchows Arch* 2000; 436: 102–108.
 65. Bettendorf O, Herrmann G. Prognostic relevance of Ki-67 antigen expression in 329 cases of oral squamous cell carcinoma. *ORL J Otorhinolaryngol Relat Spec* 2002; 64: 200–205.
 66. Carinci F, Stabellini G, Calvitti M et al. CD44 as prognostic factor in oral and oropharyngeal squamous cell carcinoma. *J Craniofac Surg* 2002; 13: 85–89.
 67. Okamoto M, Nishimine M, Kishi M et al. Prediction of delayed neck metastasis in patients with stage I/II squamous cell carcinoma of the tongue. *J Oral Pathol Med* 2002; 31: 227–233.
 68. Sommer T, Olofsson J. Significance of p53, PCNA and Ki-67 in the prognosis of squamous cell carcinoma of the oral cavity. *Laryngorhinootologie* 1997; 76: 189–196.
 69. Skalova A, Lehtonen H, von Boguslawsky K, Leivo I. Prognostic significance of cell proliferation in mucoepidermoid carcinomas of the salivary gland: clinicopathological study using MIB 1 antibody in paraffin sections. *Hum Pathol* 1994; 25: 929–935.
 70. Skalova A, Leivo I, Von Boguslawsky K, Saksela E. Cell proliferation correlates with prognosis in acinic cell carcinomas of salivary

- gland origin. Immunohistochemical study of 30 cases using the MIB 1 antibody in formalin-fixed paraffin sections. *J Pathol* 1994; 173: 13–21.
71. Hellquist HB, Sundelin K, Di Bacco A et al. Tumour growth fraction and apoptosis in salivary gland acinic cell carcinomas. Prognostic implications of Ki-67 and bcl-2 expression and of in situ end labelling (TUNEL). *J Pathol* 1997; 181: 323–329.
 72. Nordgard S, Franzen G, Boysen M, Halvorsen T. Ki-67 as a prognostic marker in adenoid cystic carcinoma assessed with the monoclonal antibody MIB1 in paraffin sections. *Laryngoscope* 1997; 107: 531–536.
 73. Yin HF, Okada N, Takagi M. Apoptosis and apoptotic-related factors in mucoepidermoid carcinoma of the oral minor salivary glands. *Pathol Int* 2000; 50: 603–609.
 74. Okabe M, Inagaki H, Murase T et al. Prognostic significance of p27 and Ki-67 expression in mucoepidermoid carcinoma of the intraoral minor salivary gland. *Mod Pathol* 2001; 14: 1008–1014.
 75. Xin W, Paulino AF. Prognostic factors in malignant mixed tumors of the salivary gland: correlation of immunohistochemical markers with histologic classification. *Ann Diagn Pathol* 2002; 6: 205–210.
 76. Cho KJ, Lee SS, Lee YS. Proliferating cell nuclear antigen and c-erbB-2 oncoprotein expression in adenoid cystic carcinomas of the salivary glands. *Head Neck* 1999; 21: 414–419.
 77. Greiner TC, Robinson RA, Maves MD. Adenoid cystic carcinoma. A clinicopathologic study with flow cytometric analysis. *Am J Clin Pathol* 1989; 92: 711–720.
 78. Hicks MJ, el-Naggar AK, Byers RM et al. Prognostic factors in mucoepidermoid carcinomas of major salivary glands: a clinicopathologic and flow cytometric study. *Eur J Cancer B Oral Oncol* 1994; 30B: 329–334.
 79. Franzen G, Nordgard S, Boysen M et al. DNA content in adenoid cystic carcinomas. *Head Neck* 1995; 17: 49–55.
 80. Pinto AE, Fonseca I, Soares J. The clinical relevance of ploidy and S-phase fraction determination in salivary gland tumors: a flow cytometric study of 97 cases. *Cancer* 1999; 85: 273–281.
 81. Junquera L, Alonso D, Sampedro A et al. Pleomorphic adenoma of the salivary glands: prospective clinicopathologic and flow cytometric study. *Head Neck* 1999; 21: 652–656.
 82. Chomette G, Auriol M, Wann A, Guilbert F. Acinic cell carcinomas of salivary glands histoprognosis. Value of NORs stained with AgNOR technique and examined with semi-automatic image analysis. *J Biol Buccale* 1991; 19: 205–210.
 83. Chomette GP, Auriol MM, Labrousse F, Vaillant JM. Mucoepidermoid tumors of salivary glands: histoprognostic value of NORs stained with AgNOR technique. *J Oral Pathol Med* 1991; 20: 130–132.
 84. Vuhahula EA, Nikai H, Ogawa I et al. Prognostic value of argyrophilic nucleolar organizer regions (AgNOR) count in adenoid cystic carcinoma of salivary glands. *Pathol Int* 1994; 44: 368–373.
 85. Xie X, Nordgard S, Halvorsen TB et al. Prognostic significance of nucleolar organizer regions in adenoid cystic carcinomas of the head and neck. *Arch Otolaryngol Head Neck Surg* 1997; 123: 615–620.
 86. Felix A, El-Naggar AK, Press MF et al. Prognostic significance of biomarkers (c-erbB-2, p53, proliferating cell nuclear antigen, and DNA content) in salivary duct carcinoma. *Hum Pathol* 1996; 27: 561–566.
 87. Timon CI, Dardick I, Panzarella T et al. Acinic cell carcinoma of salivary glands. Prognostic relevance of DNA flow cytometry and nucleolar organizer regions. *Arch Otolaryngol Head Neck Surg* 1994; 120: 727–733.
 88. Fonseca I, Soares J. Adenoid cystic carcinoma: a study of nucleolar organizer regions (AgNOR) counts and their relation to prognosis. *J Pathol* 1993; 169: 255–258.
 89. Pich A, Chiusa L, Margaria E et al. p53 overexpression correlates with proliferative activity and prognosis in carcinomas of the pyriform sinus. *Int J Oncology* 1995; 6: 1053–1058.
 90. Benazzo M, Mevio E, Occhini A et al. Proliferative characteristics of head and neck tumors. *In vivo* evaluation by bromodeoxyuridine incorporation and flow cytometry. *ORL J Otorhinolaryngol Relat Spec* 1995; 57: 39–43.
 91. Myers EN, Sampedro A, Alvarez C et al. Cell proliferation activity and kinetic profile in the prognosis and therapeutic management of carcinoma of the pharynx and larynx. *Otolaryngol Head Neck Surg* 1999; 121: 476–481.
 92. Shnayder Y, Kuriakose MA, Yee H et al. Adhesion molecules as prognostic factors in nasopharyngeal carcinoma. *Laryngoscope* 2001; 111: 1842–1846.
 93. Del Valle-Zapico A, Fernandez FF, Suarez AR et al. Prognostic value of histopathologic parameters and DNA flow cytometry in squamous cell carcinoma of the pyriform sinus. *Laryngoscope* 1998; 108: 269–272.
 94. Hake R, Eckel H, von Pritzbuere E et al. Value of monoclonal antibodies (PC 10, MIB1, p53 and LeuM 1) for assessing the prognosis of patients with squamous epithelial carcinoma of the larynx after partial laser resection. *Pathologie* 1995; 16: 197–203.
 95. Welkoborsky HJ, Hinni M, Dienes HP, Mann WJ. Predicting recurrence and survival in patients with laryngeal cancer by means of DNA cytometry, tumor front grading, and proliferation markers. *Ann Otol Rhinol Laryngol* 1995; 104: 503–510.
 96. Franchi A, Gallo O, Boddi V, Santucci M. Prediction of occult neck metastases in laryngeal carcinoma: role of proliferating cell nuclear antigen, MIB-1, and E-cadherin immunohistochemical determination. *Clin Cancer Res* 1996; 2: 1801–1808.
 97. Golusinski W, Olofsson J, Szmeja Z et al. A comprehensive analysis of selected diagnostic methods with respect to their usefulness in evaluating the biology of neoplastic cells in patients with laryngeal cancer. *Eur Arch Otorhinolaryngol* 1999; 256: 306–311.
 98. Valente G, Giusti U, Kerim S et al. High prognostic impact of growth fraction parameters in advanced stage laryngeal squamous cell carcinoma. *Oncol Rep* 1999; 6: 289–293.
 99. Lazaris ACh, Rigopoulou A, Tseleni-Balafouta S et al. Immunodetection and clinico-pathological correlates of two tumour growth regulators in laryngeal carcinoma. *Histol Histopathol* 2002; 17: 131–138.
 100. Wozniak A, Golusinski W, Kaczmarek E et al. Prognostic significance of Ki 67 and PCNA expression in laryngeal squamous cell carcinoma (morphometric evaluation of labelling index-L1). *Otolaryngol Pol* 2002; 56: 437–443.
 101. Dobros W, Rys J, Niezabitowski A, Olszewski E. The prognostic value of proliferating cell nuclear antigen (PCNA) in the advanced cancer of larynx. *Auris Nasus Larynx* 1998; 25: 295–301.
 102. Krecicki T, Jelen M. Proliferating cell nuclear antigen in laryngeal cancer. *J Laryngol Otol* 1998; 112: 310–313.
 103. Sarac S, Ayhan A, Hosal AS, Kaya S. Prognostic significance of PCNA expression in laryngeal cancer. *Arch Otolaryngol Head Neck Surg* 1998; 124: 1321–1324.
 104. Dong Y, Sui L, Tai Y et al. Prognostic significance of cyclin E overexpression in laryngeal squamous cell carcinomas. *Clin Cancer Res* 2000; 6: 4253–4258.
 105. Dong Y, Sui L, Tai Y et al. The overexpression of cyclin-dependent kinase (CDK) 2 in laryngeal squamous cell carcinomas. *Anticancer Res* 2001; 21: 103–108.
 106. Tomasino RM, Daniele E, Bazan V et al. Prognostic significance of cell kinetics in laryngeal squamous cell carcinoma: clinicopathological associations. *Cancer Res* 1995; 55: 6103–6108.

107. Danic D, Milicic D, Prgomet D, Leovic D. Prognostic factors in carcinoma of the larynx: relevance of DNA ploidy, S-fractions and localization of the tumour. *J Laryngol Otol* 1999; 113: 538–541.
108. Dobros W, Lackowska B, Rys J et al. DNA analysis of laryngeal carcinoma cells by flow cytometry: the histoclinical factors and their significance. *J Otolaryngol* 2000; 29: 371–376.
109. Bazan V, Zanna I, Migliavacca M et al. Prognostic significance of p16INK4a alterations and 9p21 loss of heterozygosity in locally advanced laryngeal squamous cell carcinoma. *J Cell Physiol* 2002; 192: 286–293.
110. Bockmuhl U, Bockmuhl F, Dimmer V, Kunze KD. 'Nucleolar organizer regions' as a factor for the prognosis of laryngeal cancer? *Laryngorhinootologie* 1992; 71: 137–141.
111. Xie X, Stenersen TC, Clausen OP, Boysen M. Nucleolar organizer regions and prognosis in glottic squamous cell carcinoma. *Head Neck* 1997; 19: 20–26.
112. Xie X, Clausen OP, De Angelis P, Boysen M. Bax expression has prognostic significance that is enhanced when combined with AgNOR counts in glottic carcinomas. *Br J Cancer* 1998; 78: 100–105.
113. Krecicki T, Jelen M, Zalesska-Krecicka M et al. Prognostic value of nucleolar organiser regions (AgNORs) in laryngeal cancer. *Acta Otorhinolaryngol Belg* 1998; 52: 215–221.
114. Spafford MF, Koeppe J, Pan Z et al. Correlation of tumor markers p53, bcl-2, CD34, CD44H, CD44v6, and Ki-67 with survival and metastasis in laryngeal squamous cell carcinoma. *Arch Otolaryngol Head Neck Surg* 1996; 122: 627–632.
115. Hirvikoski P, Kumpulainen E, Virtaniemi J et al. p53 expression and cell proliferation as prognostic factors in laryngeal squamous cell carcinoma. *J Clin Oncol* 1997; 15: 3111–3120.
116. Pulkkinen JO, Penttinen M, Jalkanen M et al. Syndecan-1: a new prognostic marker in laryngeal cancer. *Acta Otolaryngol* 1997; 117: 312–315.
117. Krecicki T, Jelen M, Zalesska-Krecicka M, Szkudlarek T. Ki-67 immunostaining and prognosis in laryngeal cancer. *Clin Otolaryngol* 1998; 23: 539–542.
118. Yamamoto Y, Itoh T, Saka T et al. Nucleolar organizer regions in glottic carcinomas: comparison of DNA cytofluorometry and clinicopathological analysis. *Eur Arch Otorhinolaryngol* 1995; 252: 499–503.
119. Barnes P, Sagar S, Laybolt K et al. Quantitation of nucleolar organizer regions by image analysis in glottic squamous cell carcinoma. *Clin Invest Med* 1999; 22: 36–43.
120. Pulkkinen JO, Klemi P, Martikainen P, Grenman R. Apoptosis in situ, p53, bcl-2 and AgNOR counts as prognostic factors in laryngeal carcinoma. *Anticancer Res* 1999; 19: 703–707.
121. Klatka J, Skomra D. Nucleolar organizer regions in laryngeal cancer. *Ann Univ Mariae Curie Sklodowska [Med]* 2001; 56: 417–421.