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The link between altered cholesterol metabolism and Alzheimer’s disease

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Alzheimer’s disease (AD), the most common form of dementia, is characterized by the progressive loss of neurons and synapses, and by extracellular deposits of amyloid-β (Aβ) as senile plaques, Aβ deposits in the cerebral blood vessels, and intracellular inclusions of hyperphosphorylated tau in the form of neurofibrillary tangles. Several mechanisms contribute to AD development and progression, and increasing epidemiological and molecular evidence suggests a key role of cholesterol in its initiation and progression. Altered cholesterol metabolism and hypercholesterolemia appear to play fundamental roles in amyloid plaque formation and tau hyperphosphorylation. Over the last decade, growing evidence supports the idea that cholesterol oxidation products, known as oxysterols, may be the missing link
between altered brain cholesterol metabolism and AD pathogenesis, as their involvement in neurotoxicity, mainly by interacting with Aβ peptides, is reported.
Alzheimer’s disease (AD), a neurodegenerative disorder, is the most common form of dementia in developed countries. It is a complex and genetically heterogeneous disease, characterized by progressive memory deficit, cognitive impairment and personality changes, accompanied by specific structural abnormalities in the brain. The main histological features of AD are extracellular deposits of amyloid-β (Aβ) in the form of senile plaques, Aβ deposits in the cerebral blood vessels, and intracellular inclusions of hyperphosphorylated tau in the form of neurofibrillary tangles (NFT). The loss of neurons and synapses in the neocortex, hippocampus and other subcortical regions of the brain is also a common feature of AD.¹²

AD begins with the abnormal processing of amyloid precursor protein (APP) by the sequential enzymatic actions of two enzymes of the amyloidogenic pathway: beta-site amyloid precursor protein-cleaving enzyme 1 (BACE1), a β-secretase, and γ-secretase; these actions lead to excess of production and/or reduced clearance of Aβ peptides, which comprise 39-43 amino acids. An imbalance between production and clearance of Aβ in the brain, and their aggregation, causes Aβ to accumulate, and this excess may be the initiating factor in AD. Monomers of Aβ₄₀ are usually much more prevalent than the aggregation-prone and damaging Aβ₄₂ species, but an increased proportion of Aβ₄₂ appears sufficient to cause early onset of AD. Additionally, insoluble oligomers and intermediate amyloids are the most neurotoxic forms of Aβ₄₂.³

Several mechanisms (e.g. perturbation of brain metabolism, oxidative stress, inflammation, presence of the apolipoprotein E (ApoE) ε4 allele, impaired cholesterol metabolism) contribute to the development and progression of AD. Among these, a growing body of epidemiological and molecular evidence suggests a mechanistic link between cholesterol and AD progression. A number of genes involved in cholesterol homeostasis have been identified as susceptibility loci for sporadic or late-onset AD,⁴⁻⁶ and altered cholesterol metabolism seems to play a fundamental role in the formation of amyloid plaques and in tau hyperphosphorylation.⁷,⁸ In addition, hypercholesterolemia
is unanimously recognized to be a risk factor for sporadic AD, a form that accounts for the great majority of cases.\textsuperscript{4,9-11} Finally, this evidence is supported by epidemiological studies indicating that cholesterol-lowering agents belonging to the family of statins reduce the prevalence of AD,\textsuperscript{12-14} a conclusion not yet fully accepted because of the contradictory results reported by prospective clinical studies.\textsuperscript{15-17}

**Apolipoprotein E and its receptors in AD**

ApoE is the brain’s principal cholesterol-carrier protein, mainly transporting it from astrocytes to neurons. The association between ApoE polymorphism and AD is presumably related to the disturbance of cholesterol transport. Of note, subjects who are homozygous for the ApoE $\varepsilon$4 genotype express an increased AD risk, versus those carrying $\varepsilon$2 or $\varepsilon$3, evidence that is consistently confirmed by numerous independent studies.\textsuperscript{18-21} In addition, receptors recognizing ApoE are also widely expressed in the AD brain.\textsuperscript{4,20,22}

Although ApoE mediates Aβ clearance by binding Aβ and forming a stable complex, ApoE may also stimulate Aβ aggregation and amyloid deposition, as well as tau hypersphorylation.\textsuperscript{20,22-25} Moreover, among the mechanisms that might explain the effects of ApoE on the brain of AD subjects, ApoE $\varepsilon$4 and its receptors are also reported to be involved in APP trafficking and its processing to Aβ. Additionally, ApoE may mediate Aβ cell internalization, by binding to the LDL receptor-related protein (LRP).\textsuperscript{26} ApoE might also modulate the distribution and metabolism of cholesterol in neuronal membranes, and regulate the role of cholesterol in synapse formation and function, through ApoE receptors.\textsuperscript{20,25,27} Moreover, $\gamma$-secretase cleavage of APP could thus regulate ApoE metabolism through the LRP1 receptor.\textsuperscript{27}

A number of epidemiological studies also report that individuals with high levels of blood cholesterol have an increased susceptibility to AD, apparently influenced by the ApoE $\varepsilon$4 genotype,
which may influence cholesterol metabolism and the formation of cholesterol oxidation products, known as oxysterols.  

**Role of cholesterol in AD**

The brain is the organ with the highest concentration of cholesterol, which is essential for its normal function, being a major component of neuronal cell membranes and a determinant of membrane fluidity. In the brain, cholesterol is mostly present in the free form and is derived from *de novo* biosynthesis from acetyl-coenzyme A mediated by 3-hydroxy-3-methyl-glutaryl-coenzyme A (HMG-CoA) reductase, rather than from plasma lipoproteins, which are prevented by the blood-brain barrier (BBB) from crossing from the peripheral circulation into the brain. Further, the astrocytic compartment meets neuronal cholesterol demands by secreting ApoE-cholesterol complexes, which are transported to the neurons.  

The mechanism by which cholesterol affects Aβ production and metabolism is not fully understood; however, a change in membrane properties has been suggested. Cholesterol is mainly concentrated in membrane microdomains termed “lipid rafts”, where considerable evidence indicates that the amyloidogenic pathway takes place. In this connection, it has been reported that cellular cholesterol, especially when it is elevated in the membrane, binds directly to APP and thus promotes APP’s insertion into the phospholipid monolayers of the “lipid rafts” and other organelles where β- and γ-secretases reside, and favors the amyloidogenic pathway. Amyloidogenic activity is thus linked to cholesterol levels: β- and γ-secretase activities are positively regulated by high and inhibited by low levels of cholesterol. Since APP processing and Aβ generation are associated with cholesterol-rich microdomains, and both are present in “lipid rafts”, Aβ production may require raft integrity and a lipid component as optimal conditions. An alteration in raft components could thus change the configuration of either the enzymes or the substrate associated with the rafts, leading to an alteration in Aβ generation. Conversely, in the non-
amyloidogenic pathway, APP is processed by \( \alpha \)-secretase in non-raft domains, and this event is promoted by a decreased cellular cholesterol level.\(^39\) Moreover, the amyloidogenic pathway is inactivated when \( \alpha \)-secretase is forced to associate with “lipid rafts”.\(^40\)

Cholesterol also enhances A\( \beta \) to form neurotoxic aggregates.\(^41\) In addition, fibrillogensis of A\( \beta \) has been proposed to take place in “lipid rafts” at ganglioside clusters, where A\( \beta \) displays a specific affinity to cholesterol which binds avidly to A\( \beta \) protofibrils.\(^42,43\) Moreover, it has been shown that increased cholesterol levels in the lipid bilayers facilitate binding of A\( \beta \) to the membranes, promoting A\( \beta \) conformational change from a helix-rich to a \( \beta \)-sheet-rich structure, and thus becoming an endogenous seed for amyloid formation.\(^44\) The conversion of soluble and nontoxic monomeric A\( \beta \) to insoluble and toxic oligomeric and aggregate A\( \beta \) is thus the critical step in AD development.

Given the above considerations, it seems that cholesterol distribution and trafficking within brain cells, rather than total cholesterol levels in the neurons, are the relevant factors in the APP processing and A\( \beta \) accumulation in the AD progression.\(^45\)

It has also been observed that high concentrations of free cholesterol alone do not affect APP processing or A\( \beta \) production; rather, the conversion of excess free cholesterol into cholesterol ester has a profound effect on APP and A\( \beta \) enhancing their upregulation.\(^46\) Consequently, clearance of A\( \beta \) from the brain is reduced when an overabundance of esterified cholesterol decreases membrane lipid turnover. Conversely, inhibition of the enzyme acyl-coenzyme A:cholesterol acyltransferase 1 (ACAT1), which esterifies cholesterol, leads to the reduction of both cholesteryl esters and A\( \beta \).\(^47\) These data suggest that the balance between free cholesterol and cholesterol esters is a key parameter controlling amyloidogenesis, although the molecular mechanisms underlying this relationship are still unclear.

A regulatory role for APP and \( \gamma \)-secretase in cholesterol metabolism, jointly acting to lower cellular cholesterol levels, has also been reported.\(^27\) The effect of A\( \beta \) and of the intracellular domain
of APP (AICD) on cellular cholesterol metabolism have also been investigated. Aβ, in particular the oligomeric rather than the monomeric form, alters intracellular trafficking and cholesterol homeostasis, by promoting the release of cholesterol and some other lipids from cells, in the form of Aβ-lipid particles. The fibrillar Aβ then down-regulates cholesterol biosynthesis. The AICD, which is released upon γ-secretase cleavage of APP, down-regulates cellular cholesterol uptake by acting as a transcriptional suppressor of LRPI gene, a major ApoE receptor in the brain. Furthermore, the peptide products of the amyloidogenic pathway ultimately reduce both cholesterol uptake and its biosynthesis, completing a negative feedback loop. A decrease in cellular cholesterol levels then results in an enhancement of tau phosphorylation and synaptic failure. It has also been demonstrated that extracellular cholesterol accumulates in the senile plaques and neurofibrillary tangles of AD patients, and in transgenic mice expressing the Swedish Alzheimer mutation APP751, as well as ApoE, and that cholesterol, ApoE and Aβ all colocalize in the core of fibrillar plaques. This shows that both ApoE and cholesterol may be essential for plaque formation, and that extracellular cholesterol, by binding to aggregated Aβ, may be the seed for its deposition.

**Role of oxysterols in AD**

Because there is little synthesis of cholesterol in the adult brain, and because the brain cannot degrade cholesterol, it must be excreted from the brain in order to prevent its accumulation. The most important mechanism whereby the brain eliminates excess cholesterol is through the formation and excretion into the circulation of oxysterols, a class of cholesterol oxidation products, which are thus important to balance the local synthesis of sterols.

Cholesterol is primarily converted into the oxysterol 24-hydroxycholesterol (24-OH), also known as cerebrosterol, which is produced almost exclusively in the brain by CYP46A1 (cholesterol 24-hydroxylase) and which, unlike cholesterol itself, can easily cross the BBB.
Following its secretion from the brain, 24-OH enters the circulation and reaches the liver, where it is taken up and metabolized. The oxysterol 24-OH also plays an important role in the regulation of cholesterol homeostasis in the brain. Neuronal cells have a lower rate of cholesterol synthesis than glial cells; for this reason, an increased flux of 24-OH from neurons to glial cells causes an increased flux of cholesterol to the neuronal cells, by means of activation of the nuclear liver X receptor (LXR) and upregulation of ApoE in the glial cells (Fig. 1).\(^6\) Another oxysterol, 27-hydroxycholesterol (27-OH), has been found to be produced in situ in the brain by CYP27A1, although in small amounts, and then metabolized by the enzyme CYP7B to 7α-hydroxy-3-oxo-4-cholestenoic acid (7-OH-4-C), which, crossing the BBB, reaches the liver where it is eliminated.\(^5\)\(^4\),\(^5\)\(^5\),\(^5\)\(^7\) However, it has been observed that most 27-OH flows from the circulation into the brain, since, unlike cholesterol, it can cross the BBB.\(^5\)\(^8\) Summarizing, these considerations indicate that there are fluxes of oxysterols in opposite directions across the BBB: two fluxes out of the brain (24-OH and 7-OH-4-C) and one flux into the brain (27-OH). A further compound, 7β-hydroxycholesterol (7β-OH), may also derive in the brain from oxidation of cholesterol following cholesterol interaction with Aβ and APP (Fig. 2).\(^5\)\(^9\)

During the last decade, the idea that oxysterols might be the missing link between altered brain cholesterol metabolism and AD pathogenesis has been increasingly supported by research pointing to the involvement of 24-OH and 27-OH in neurotoxicity, mainly by interacting with Aβ peptides.

Several studies have found higher levels of 24-OH in the peripheral circulation and cerebrospinal fluid (CSF) of AD patients during the early stages than in unaffected individuals, suggesting that cholesterol turnover in the brain increases during the neurodegenerative changes of AD.\(^6\)\(^0\)-\(^6\)\(^2\) Conversely, plasma levels of 24-OH were decreased in patients with later stages of AD than in the respective controls, suggesting that the rate of cholesterol transport lowers as the disease progresses.\(^6\)\(^2\),\(^6\)\(^3\) These contradictory results might be rationalized by considering that increased plasma levels of 24-OH reflect ongoing neurodegeneration and/or demyelinization, whereas
decrease

dplasma levels reflect a selective loss of neurons expressing CYP46A1.\textsuperscript{30} However, in glial cells of AD brains there is some ectopic induction of CYP46A1, and consequently some 24-OH production, which may overlap with decreased neuronal expression in the presence of increased glial expression,\textsuperscript{64,65} although this induction of CYP46A1 cannot compensate for the loss of 24-hydroxylase activity due to the neuronal degeneration. Another study, however, has found that plasma levels of 24-OH in AD patients are not significantly different than in control subjects.\textsuperscript{66}

It has also been reported that, in all brain areas of deceased AD patients, as well as in aged mice expressing the Swedish Alzheimer mutation APP751, the amount of 24-OH decreases and 27-OH increases.\textsuperscript{67} A marked accumulation of 27-OH was also found in the brain of patients carrying the Swedish APP 670/671 mutation.\textsuperscript{68} Whereas the decreased levels of 24-OH in the AD brain are presumably due to the loss of neuronal cells and consequent loss of the enzyme CYP46A1, the increased levels of 27-OH may be due to increased flux of this oxysterol across the BBB, because of hypercholesterolemia\textsuperscript{69} or a damage of BBB integrity.\textsuperscript{70} An alternative explanation for the high levels of 27-OH is a reduced metabolism of the oxysterol into 7-OH-4-C, by the enzyme CYP7B, which is reduced in the brain of AD patients.\textsuperscript{71} However, increased levels of both 24-OH and 27-OH have been observed in the CSF in patients with advanced AD.\textsuperscript{72}

From these considerations, the hypothesis has been formulated that the balance between 24-OH and 27-OH is important for amyloidogenesis,\textsuperscript{54,69} and the increased ratio of 27-OH to 24-OH in AD brains is consistent with this hypothesis.\textsuperscript{67} Thus, the shift in balance between the two oxysterols might lead to increased generation and accumulation of A\textsubscript{\beta} and regulation of the 24-OH/27-OH ratio could be an important strategy in controlling A\textsubscript{\beta} levels in AD. However, opinions still differ about the involvement of these two oxysterols in the APP processing and A\textsubscript{\beta} generation.

According to several studies, induction of CYP46A1 activity has beneficial effects, directly preventing A\textsubscript{\beta} generation by modulating cholesterol homeostasis and reducing cellular cholesterol. Indeed, astrocytes are sensitive to 24-OH-mediated upregulation of the LXR-responsive genes involved in cholesterol efflux, i.e. ATP-binding cassette transporter A1 and G1 (ABCA1 and
ABCG1) and ApoE. Conversely, the low levels of 27-OH in the brain might also be expected not to affect amyloidogenesis. However, since the flux of 27-OH across the BBB increases under conditions of hypercholesterolemia, or in the case of reduced BBB integrity, the inhibitory effect of 24-OH on Aβ generation is consequently reduced. In this connection, the high flux of 27-OH from the peripheral circulation to the brain, and changes in the brain cholesterol/oxysterol balance, may partially explain the link between hypercholesterolemia and AD. Nevertheless, in murine primary neuronal cells both 24-OH and 27-OH were found to inhibit Aβ formation and secretion, 24-OH being about 1000-fold more potent than 27-OH. The study showed that the distribution of the enzymes CYP46A1 and CYP27A1 is altered in the brain of subjects with AD. CYP46A1, whose expression increases in astrocytes and decreases in neurons, is selectively expressed in degenerating neuritis around senile plaques, whereas CYP27A1 expression, which is mostly expressed in neurons but also to a lesser extent in astrocytes and oligodendrocytes, is increased in white-matter oligodendrocytes. Moreover, it has been shown that 27-OH significantly reduces Aβ peptide generation from primary human neurons, not by affecting α-, β-, or γ-secretase but by upregulating LXR responsive genes (ABCA1, ABCG1 and ApoE). Of note the LXR-mediated gene regulation of cholesterol efflux and metabolism not only modulates neurodegeneration and Aβ peptide transport and clearance but also inflammation in the brain. On the basis of these data it cannot be excluded that 27-OH, as an LXR ligand, might exert anti-amyloidogenic effects by reducing extracellular Aβ and inflammation.

By contrast, other studies are consistent with the possibility that 27-OH may accelerate neurodegeneration. In human SH-SY5Y neuroblastoma cells, 24-OH directly increases α-secretase activity as well as elevating the α/β activity ratio, whereas 27-OH counteracts the inhibitory effect of 24-OH on the generation of amyloid. In addition, recent studies underlined the different effects of 24-OH and 27-OH on APP levels and processing in human neuroblastoma SH-SY5Y cells and in the brain tissue: 24-OH may favor the non-amyloidogenic pathway, whereas 27-OH is thought to
enhance production of Aβ42 by upregulating APP and BACE1, and tau hyperphosphorylation. Another study has found that 27-OH increases Aβ accumulation by reducing insulin-like growth factor 1 (IGF1) levels, a neurotrophic factor that promotes neurogenesis and has a neuroprotective effect, in hippocampal slices from adult rabbits; in addition, 27-OH reduces the production of the “memory protein” activity-regulated cytoskeleton-associated protein (Arc) in mouse brain.

Alongside altered cholesterol metabolism and hypercholesterolemia, inflammatory response and oxidative stress also significantly contribute to neuronal damage in AD.

The importance of inflammatory processes has been pointed out during the past decade by the intensive investigation of inflammatory mediators and microglia activation in the brain of AD, although it remains unclear whether inflammation represents a cause or a consequence of AD. It has been reported that intraneuronal Aβ and soluble Aβ oligomers activate microglia in the earliest stages of the disease, even before plaque and tangle formation, in particular when cells are stressed. Fibrillar Aβ also can activate microglia by binding to cells via specific receptors, in particular through a multireceptor complex involving CD36, α6β1-integrin and CD47. The induction of a microglia-driven inflammatory response results in the release of various inflammatory mediators, including a whole array of neurotoxic cytokines and free radicals. Once activated, microglia cells may also recruit astrocytes, which actively enhance the inflammatory response to extracellular Aβ deposits that intensify neuronal dysfunction and cell death: inflammatory mediators and other components of the immune system are often found near areas of amyloid plaques.

It has also been postulated that oxidative stress may be either a cause or a consequence of the neuropathology associated with AD and, in support of its being a consequence, Aβ can stimulate the production of reactive oxygen species (ROS). Conversely, oxidative stress has been shown to contribute to the formation of amyloid plaques, since Aβ peptide has been found in the oxidized form. Since the brain has a high lipid content, it is extremely vulnerable to free radicals
and ROS, which are responsible for enhancing lipid peroxidation, including cholesterol oxidation and oxysterol formation as well as tau hyperphosphorylation and NFT formation. mitochondrial insufficiency and neuronal cell death.

Oxysterols have been shown to enhance Aβ aggregation and its neurotoxicity, by modifying specific sites of Aβ peptide. Following Aβ modification at Lys-16, peptide aggregates were formed faster than in the case of modification at Lys-28 or Asp-1. Moreover, 7β-OH has been found to be neurotoxic at nanomolar concentrations in cultured rat hippocampal neuronal cells, and may therefore contribute to Aβ-related neurodegeneration in the brain of AD patients. Another oxysterol that might derive from the autooxidation of cellular cholesterol released during neurodegeneration, is 7α-hydroperoxycholesterol, which has also been found to be responsible for necrotic cell death of SH-SY5Y cells, and a further possibility is 7-ketocholesterol. Additionally, 24-OH has been shown to enhance the neurotoxic effect of the Aβ₄₂ peptide in the human differentiated neuroblastoma cell line MSN, as well as augmenting ROS generation.

In our recent study, we examined the ways in which the oxysterols 24-OH, 27-OH, and 7β-OH, specifically implicated in brain pathophysiology, may modulate and possibly amplify the expression of AD. All three oxysterols strongly enhanced the binding and concentration of Aβ₄₂ on membranes of human differentiated neuronal cell lines (SK-N-BE and NT-2) by markedly upregulating expression and synthesis of CD36 and β1-integrin receptors, two components of the multireceptor complex CD36/β1-integrin/CD47, through which Aβ peptide binds to membrane cells. An interesting finding of the same study is that only 24-OH significantly potentiates both the necrogenic and the apoptotic effects exerted by Aβ₄₂ peptide on these cells (Fig. 3A and B). These effects were inhibited when 24-OH-treated neuronal cells were incubated with anti-CD36 and anti-β1-integrin antibodies before Aβ₄₂ addition, since Aβ peptide binding to the cell surface was prevented. One significant reason for this selective behavior of 24-OH appears to be its marked pro-oxidant action on neuronal cells, by locally increasing ROS generation, an action not exerted by
either 27-OH or 7β-OH (Fig. 3C). The 24-OH-dependent potentiation of Aβ neurotoxicity was completely inhibited by incubation of differentiated SK-N-BE or NT-2 cells with either the flavonol quercetin or the isoflavone genistein.105

However, despite the present knowledge demonstrates that cholesterol and some oxystetrols play a key role in the AD pathogenesis interacting with Aβ (Fig. 4), elucidation of the precise mechanisms will clearly require further study.

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References


Figure legends

**Figure 1.** Cholesterol homeostasis in the mature brain. Excess cholesterol is converted into 24-hydroxycholesterol (24-OH) in neuronal cells by CYP46A1. Most of the 24-OH goes directly from the brain into the blood circulation, a small quantity entering the cerebrospinal fluid (CSF). 24-OH may also be caught by astrocytes, where it upregulates the nuclear receptor liver X receptor (LXR)-responsive genes involved in cholesterol efflux, i.e. ATP-binding cassette transporter A1 and G1 (ABCA1 and ABCG1) and apolipoprotein E (ApoE). Synthesized cholesterol is loaded by astrocytes onto ApoE, and the ApoE/cholesterol complex is then internalized by neurons via low-density lipoprotein receptors (LDLR).

**Figure 2.** Fluxes of 24-hydroxycholesterol (24-OH) and 27-hydroxycholesterol (27-OH) through the blood brain barrier (BBB). The enzymes CYP46A1 and CYP27A1, located in the neuronal cells, are respectively responsible for generating 24-OH and 27-OH in the brain. However, most of the 27-OH flows from the circulation into the brain since, unlike cholesterol, it can cross the BBB, as can 24-OH. In the brain, 27-OH is also metabolized to $7\alpha$-hydroxy-3-oxo-4-cholestenoic acid (7-OH-4-C), which crosses the BBB to reach the liver. In conclusion, there is a complex of fluxes of oxysterols in opposite directions at the BBB: two fluxes out of the brain (24-OH and 7-OH-4-C) and one flux into the brain (27-OH). Of note, the shift in balance between 24-OH and 27-OH is important for amyloidogenesis since it might lead to increased generation and accumulation of Aβ in AD brain.

**Figure 3.** (A) Necrotic effects of 27-hydroxycholesterol (27-OH), 7β-hydroxycholesterol (7β-OH), or 24-hydroxycholesterol (24-OH) evaluated in terms of lactate dehydrogenase (LDH) release. SK-N-BE cells were treated with the oxysterol (1 µM) for 48 h, and then for 24 h with Aβ$_{42}$ (1 µM). Histograms represent the mean values ± SD of 3 experiments. **P < 0.01 vs. control (untreated cells).** (B) Apoptotic effects of 27-OH, 7β-OH, or 24-OH on SK-N-BE cells observed by DAPI
staining to determine apoptotic nuclei formation. Cells were treated with the oxysterol for 48 h and then for 24 h with Aβ42. (C) Pro-oxidant effects of 27-OH, 7β-OH, or 24-OH. Intracellular generation of reactive oxygen species (ROS) was examined in SK-N-BE cells using 2’,7’-dichlorodihydrofluorescein (DCFH-DA) as an intracellular probe. Cells were incubated with oxysterol for 1 h, or simultaneously with oxysterol plus Aβ42 for 1 h.

**Figure 4.** Interaction between lipid metabolism and amyloid-β (Aβ) processing. The major source of cerebral cholesterol is *de novo* synthesis from acetyl-coenzyme A mediated by 3-hydroxy-3-methyl-glutaryl-coenzyme A (HMG-CoA) reductase. Excess cholesterol is converted into cholesteryl esters by acyl-coenzyme A:cholesterol acyl-transferase (ACAT) and into the oxysterols 24- and 27-hydroxycholesterol (24-OH, 27-OH) by CYP47A1 and CYP27A1, respectively; the oxysterol 7β-hydroxycholesterol (7β-OH) may derive from cholesterol oxidation by Aβ and amyloid protein precursor (APP). Aβ production is increased by HMG-CoA reductase and ACAT activity and by cholesterol which regulates β- and γ-secretase. LDL receptor-related protein (LRP) mediates the internalization of astrocyte-produced ApoE-cholesterol complexes into neurons. The intracellular domain of APP down-regulates cellular cholesterol uptake by acting as a transcriptional suppressor of LRP1 gene. 24-OH and 27-OH upregulate ATP-binding cassette transporter A1 (ABCA1), involved in cholesterol efflux, which modulates Aβ levels in neurons; these oxysterols increase ApoE levels and induce Aβ aggregation which is also promoted by free ApoE and cholesterol.
Fig. 1
Fig. 2
Fig. 3

A

LDH release (%)

control  Aβ42  27-OH  27-OH+AB42  7β-OH  28-OH+AB42  24-OH  24-OH+AB42

B

control  27-OH  7β-OH  24-OH  Aβ42  27-OH  Aβ42  7β-OH  Aβ42  24-OH  Aβ42

C

control  27-OH  7β-OH  24-OH  Aβ42  27-OH  Aβ42  7β-OH  Aβ42  24-OH  Aβ42
Fig. 4

NEURON

Acetyl-coA

HMG-CoA reductase

Aβ

APP

ApoE

LRP

ApoE/cholesterol complex

Cholesterol

ACAT

cholesteryl esters

non enzymatic oxidation

7β-OH

24-OH

27-OH

ABCA1

Aβ monomers

Aβ aggregates

Amyloid fibrils plaques

--- dashed line: promotion

--- solid line: inhibition