Cross talk between beta(1) and alpha(V) integrins: beta(1) affects beta(3) mRNA stability.

This is the author's manuscript

Original Citation:
Cross talk between beta(1) and alpha(V) integrins: beta(1) affects beta(3) mRNA stability. / RETTA SF; CASSARA’ G; D’AMATO M; ALESSANDRO R; PELLEGRINO M; DEGANI S; DE LEO G; SILENGO L; TARONE G. - In: MOLECULAR BIOLOGY OF THE CELL. - ISSN 1059-1524. - 12(2001), pp. 3126-3138.

Availability:
This version is available http://hdl.handle.net/2318/41177 since

Terms of use:
Open Access
Anyone can freely access the full text of works made available as "Open Access". Works made available under a Creative Commons license can be used according to the terms and conditions of said license. Use of all other works requires consent of the right holder (author or publisher) if not exempted from copyright protection by the applicable law.

(Article begins on next page)
Cross Talk between $\beta_1$ and $\alpha_v$ Integrins: $\beta_1$ Affects $\beta_3$ mRNA Stability

Saverio Francesco Retta,*† Georgia Cassara,‡ Monica D’Amato,‡ Riccardo Alessandro,‡ Maurizio Pellegrino,* Simona Degani,* Giacomo De Leo,‡ Lorenzo Silengo,* and Guido Tarone*

*Department of Genetics, Biology, and Biochemistry, University of Torino, 10126 Torino, Italy; and ‡Department of Biopatologia e Metodologie Biomediche, University of Palermo, 90133 Palermo, Italy

Submitted March 20, 2001; Revised June 27, 2001; Accepted July 11, 2001

Monitoring Editor: Richard Hynes

There is increasing evidence that a fine-tuned integrin cross talk can generate a high degree of specificity in cell adhesion, suggesting that spatially and temporally coordinated expression and activation of integrins are more important for regulated cell adhesive functions than the intrinsic specificity of individual receptors. However, little is known concerning the molecular mechanisms of integrin cross talk. With the use of GD25 cells ectopically expressing the $\beta_1 A$ integrin subunit, we provide evidence for the existence of a cross talk between $\beta_1$ and $\alpha_v$ integrins that affects the ratio of $\alpha_v \beta_3$ and $\alpha_v \beta_5$ integrin cell surface levels. In particular, we demonstrate that a down-regulation of $\alpha_v \beta_3$ and an up-regulation of $\alpha_v \beta_5$ occur as a consequence of $\beta_1 A$ expression. Moreover, with the use of GD25 cells expressing the integrin isoforms $\beta_1 B$ and $\beta_1 D$, as well as two $\beta_1$ cytoplasmic domain deletion mutants lacking either the entire cytoplasmic domain ($\beta_1 TR$) or only its “variable” region ($\beta_1 COM$), we show that the effects of $\beta_1$ over $\alpha_v$ integrins take place irrespective of the type of $\beta_1$ isoform, but require the presence of the “common” region of the $\beta_1$ cytoplasmic domain. In an attempt to establish the regulatory mechanism(s) whereby $\beta_1$ integrins exert their trans-acting functions, we have found that the down-regulation of $\alpha_v \beta_3$ is due to a decreased $\beta_3$ subunit mRNA stability, whereas the up-regulation of $\alpha_v \beta_5$ is mainly due to translational or posttranslational events. These findings provide the first evidence for an integrin cross talk based on the regulation of mRNA stability.

INTRODUCTION

Integrins form one family of cell adhesion receptors that play a prominent role in the adhesive interactions between cells and their surrounding extracellular matrix (ECM) (Hynes, 1992). All integrins are heterodimers composed of noncovalently linked $\alpha$ and $\beta$ subunit transmembrane glycoproteins containing large extracellular domains, short transmembrane domains, and carboxyl-terminal cytoplasmic domains of variable length (Hynes, 1992). These adhesive receptors are endowed with both structural and regulatory functions, linking extracellular matrix to the actin cytoskeleton at focal adhesion sites and providing bidirectional transmission of signals across the plasma membrane (Schoenwaelder and Burridge, 1999; Critchley, 2000). The cytoplasmic domain of the $\beta$ subunit has been shown to play a critical role in focal adhesion and actin stress fiber organization and both outside-in and inside-out integrin signaling (Liu et al., 2000).

Through their molecular interactions integrins regulate a number of critical cellular processes, including proliferation, differentiation, survival, migration, and gene expression (Giancotti, 1997; Giancotti and Ruoslahti, 1999). It is now clear that altered, modulated, or regulated adhesive interactions can change the way cells interact with their environment with dramatic consequences for both normal and pathological conditions. Cells can vary their adhesive properties by selectively expressing different integrins and by modulating their integrin specificity and affinity for ligands (Hynes, 1996). However, cells often display multiple integrins capable of interacting with a particular ECM protein and, conversely, individual integrins can recognize several extracellular matrix molecules (Hynes, 1992). Thus, integrin expression and ligand specificity are often apparently redundant, at least in terms of simple adhesion. The biological significance of this phenomenon is not clear yet; nevertheless, there is increasing evidence that individual integrin

† Corresponding author. E-mail address: francesco.redda@unito.it.
Abbreviations used: ECM, extracellular matrix; FN, fibronectin.
receptors mediate distinct functions and can convey unique information (Giancotti, 2000).

Most integrins belong to one of two major subfamilies defined by the β1 and αv subunits. The β1 subunit pairs with at least 12 different α subunits (α1-α12, αv) to comprise receptors for a variety of ECM proteins, including collagen, laminin, fibronectin, and vitronectin (Hynes, 1992). A large body of literature (Brakebusch et al., 1997; Giancotti, 1997; reviewed in Schoenwaelder and Burridge, 1999) has addressed the role of β1 integrins in mediating important cell adhesion and signal transduction events. Four different β1 isoforms have been identified (β1A, β1B, β1C, and β1D), which differ in their cytoplasmic domains and differentially affect many integrin functions (Belkin et al., 1997; Fornaro and Languino, 1997; Belkin and Retta, 1998; Pfaff et al., 1998; Retta et al., 1998).

The αv subunit is known to associate with at least five different β subunits (β1, β3, β5, β6, and β8). Among these αv integrins, αvβ3 and αvβ5 have been extensively studied. The αvβ3 integrin, in particular, has a relatively limited cellular and tissue distribution (Yamada et al., 1995), but its expression and activity are tightly regulated during a variety of biological processes, including cell proliferation and survival (Montgomery et al., 1994), wound healing (Clark et al., 1996a), angiogenesis (Brooks et al., 1994), bone remodeling (McHugh et al., 2000), tumor progression (Albelda et al., 1996), and metastasis (Yun et al., 1996). This integrin can bind to a variety of ECM proteins, including vitronectin, fibronectin, fibrinogen, thrombospondin, von Willebrand factor, and denatured collagen (Kühn and Eble, 1994), and it is able to recruit cytoskeletal and signaling proteins to focal adhesion sites (Lewis et al., 1996). In addition, αvβ3 is one of the integrins that promotes the assembly of fibronectin matrix (Wennerberg et al., 1996; Wu et al., 1996; Retta et al., 1998).

In contrast to αvβ3, αvβ5 is among the most widely expressed integrins. This receptor can specifically and efficiently bind its ligand vitronectin but remains randomly distributed over the surface of the cells and does not trigger the assembly of focal adhesion structures (Wayner et al., 1991; Leavels et al., 1992). Moreover, αvβ5 integrin has different requirements than αvβ3 for mediating adhesive events, such as cell spreading and migration, to the common ligand vitronectin (Klemen et al., 1994; Lewis et al., 1996), and it can induce differential biological responses (Friedlander et al., 1995).

Perturbation experiments with antibodies, blocking peptides, and antisense oligonucleotides demonstrated that both β1 and αv integrins play a primary role in important physiological and pathological processes (reviewed in Varner and Chereson, 1996; Brakebusch et al., 1997; Bader et al., 1998). However, recent genetic analyses have clearly increased questions as to the primacy of these integrins, and instead have pointed to a cross talk model where spatiotemporal regulation, combinatorial expression, and activation of several integrin receptors generate a high degree of specificity in cell adhesion (Fassler et al., 1996; Hynes, 1996, 1999; Brakebusch et al., 1997; Bader et al., 1998; Hodivala-Dilke et al., 1999; McHugh et al., 2000). Several observations indicate the existence of a cross talk between β1 and αv integrins, which usually takes the form of one integrin influencing the functional behavior of another integrin expressed on the same cell (Yang and Hynes, 1996; Belkin et al., 1997; Retta et al., 1998; Bluestone et al., 1999; Corbett and Schwarzbaumer, 1999). However, in most cases, the mechanistic basis of this receptor cross talk is not completely understood, and it is unknown whether and how the integrin cross talk can regulate the ratio of integrin cell-surface expression levels.

GD25 cells, derived from β1-null mouse embryonic stem cells (Wennerberg et al., 1996), are a valuable model for examination of integrin cross talk. In fact, these cells express αvβ3, αvβ5, and αvβ6 as major integrin complexes but do not express integrins of the β1 subfamily, thus permitting a variety of genetic experiments exploring the basis of integrin cross talk. We have previously transfected GD25 cells with cDNAs encoding for the isoform A, B, or D of the human β1 integrin subunit or two β1 mutants lacking either the entire cytoplasmic domain (β1 TR) or only the cytoplasmic domain “variable” region that characterizes each isoform (β1COM) (Retta et al., 1998). With the use of these cells, we investigated the specific functional properties of the isoform B and D of the human β1 integrin subunit, showing the existence of a functional cross talk between these two β1 isoforms and the endogenous αv integrins. In particular, both β1B and β1D expression prevented different fibronectin (FN)-dependent αv integrin functions, including its ability to mediate cell adhesion, to localize to focal adhesions, and to assemble an FN matrix (Belkin et al., 1997; Retta et al., 1998).

In the present study, we show that the cross talk between β1 and αv integrins is mainly based on the regulation of β3 and β5 integrin subunit expression exerted by β1 integrins. In fact, the ectopic expression of either β1A, β1B, or β1D in GD25 cells induces a drastic down-regulation of β3 and an up-regulation of β5 integrin cell surface levels. Moreover, analysis of GD25 cells expressing β3 integrins lacking either the entire β3 cytoplasmic domain (β3TR) or only its variable region (β3COM) demonstrate that the “common” region of the β3 cytoplasmic domain is required for these effects. We further demonstrate that β3 exerts its control over αvβ3 expression level by modulating the β3 mRNA stability, whereas the up-regulation of αvβ5 is mainly due to translational or posttranslational events leading to an increased recruitment of the β5 subunit at the cell surface.

MATERIALS AND METHODS

Antibodies and Reagents

The mouse anti-human β1 monoclonal antibody (mAb) TS2/16 was obtained from American Type Culture Collection (Manassas, VA). The rat anti-mouse αv mAb GoH3 was a gift from A. Sonnenberg (The Netherlands Cancer Institute, Amsterdam, The Netherlands). The rabbit polyclonal antisera to αv, αv, and αv integrin cytoplasmic domains, produced in our laboratory, were previously described (Retta et al., 1998). The polyclonal antisera to β3 and β5 were produced with the use of a previously described protocol (Delliprino et al., 1995). Briefly, rabbits were immunized against a GST-β3 fusion protein containing the cytoplasmic domain of the mouse β3 integrin subunit and against a synthetic peptide reproducing an amino acid sequence from the carboxy terminus of mouse β3 integrin subunit, respectively. The β3 peptide EKAQLKPKATSDLA was synthesized by solid phase methods with the use of an LKB Biolynx synthesizer (Amersham Pharma Biotech AB, Uppsala, Sweden) and coupled to keyhole limpet hemocyanin with the use of glutaraldehyde. The mouse anti-paxillin mAb was purchased from Transduction Laboratories (Nottingham, United Kingdom). The affinity-purified rhodamine-labeled goat anti-mouse and goat anti-rabbit IgG were from Sigma (St. Louis, MO). Poly-l-lysine and monensin were from...
Cells and Culture Conditions

The mouse GD25 fibroblast line, which lacks expression of \(\beta_1\) integrin heterodimers because of disruption of the \(\beta_1\) gene by homologous recombination, was established after differentiation of \(\beta_1\)-null embryonic stem cells and immortalization with simian virus 40 large T antigen (Wennerberg et al., 1996). GD25 cells expressing the human \(\beta_1A, \beta_2B, \text{ or } \beta_6D\) integrin isoforms or the \(\beta_1\)TR and \(\beta_1\)COM human \(\beta_1\) mutants, lacking the entire cytoplasmic domain and the cytoplasmic domain variable region, respectively, were obtained as previously described (Belkin et al., 1997; Retta et al., 1998). To avoid selection for anomalous functional traits, no efforts were made to establish clonal cell lines; instead, bulk cell populations expressing \(\beta_1\) integrins were selected. Cells were cultured in DMEM (Invitrogen Carlsbad, CA) supplemented with 10% fetal bovine serum (FBS) (Invitrogen), 2 mM L-glutamine, 100 U/ml penicillin, and 100 \(\mu\)g/ml streptomycin. The \(\beta_1\)-expressing GD25 cells were cultured in the same medium plus 300 \(\mu\)g/ml hygromycin B (Roche Molecular Biochemicals, Mannheim, Germany). Cell populations expressing high levels of the \(\beta_1\) forms used were selected by the panning method and monitored by flow cytometry as described previously (Retta et al., 1998).

Biotinylation of Cell Surface Proteins

Adherent cells, grown to 80–90% confluence in 90-mm tissue culture dishes, were washed twice with ice-cold buffer A (1.3 mM CaCl\(_2\), 0.4 mM MgSO\(_4\), 5 mM KCl, 138 mM NaCl, 5.6 mM glucose, 25 mM HEPES, pH 7.4) and incubated with 0.5 mg/ml membrane-impermeable biotinylation reagent Sulfo-NHS-Biotin (Sigma) in buffer A at 4°C for 30 min. The reaction was quenched with DMEM containing 0.6% bovine serum albumin (BSA) and 25 mM HEPES, pH 7.4. The cells were then washed four times with ice-cold buffer A and lysed on ice in Tris-buffered saline (150 mM NaCl, 20 mM Tris-HCl, pH 7.4) containing 0.5% Triton X-100 and the protease inhibitors aprotinin (10 \(\mu\)g/ml), leupeptin (10 \(\mu\)g/ml), phenylmethylsulfonyl fluoride (1 mM), and benzamidine (1 mM) (all from Sigma). Cell lysates were centrifuged at 12,000 \(\times\) g for 30 min at 4°C, and total protein concentration in the supernatants was determined with the use of a bicinchoninic acid protein assay (Pierce, Rockford, IL). Supernatants containing equal amounts of proteins were precleared with a mixture of protein A-Sepharose and protein G-Sepharose and used in immunoprecipitation experiments.

Immunoprecipitation and Analysis of Integrins

Integrins were immunoprecipitated from precleared cell lysate supernatants by incubation with appropriate dilutions of specific antibodies and a mixture of protein A-Sepharose and protein G-Sepharose beads for 1 h at 4°C. Complexes were washed four times with the lysis buffer then the proteins were eluted with Laemmli’s sample buffer (62.5 mM Tris-HCl, pH 6.8, 20% glycerol, 2% SDS) and subjected to SDS-polyacrylamide (7.5%) gel electrophoresis under nonreducing conditions. To visualize the biotinylated proteins, the gel was electroblotted onto Hybond-C transfer membrane (Amersham Pharmacia Biotech AB). The blot was then blocked with 5% BSA in phosphate-buffered saline (PBS) for 1 h at 42°C, incubated with streptavidin-peroxidase (Sigma) (1:10,000 in PBS/1% BSA) for 1 h at room temperature, and further processed by the Western blotting enhanced chemiluminescence (ECL) detection system (Amersham Pharmacia Biotech AB).

Reverse Transcription-Polymerase Chain Reaction (RT-PCR) and Northern Blot Analysis

Total RNA was isolated from 1 \(\times\) 10\(^6\) cultured cells with the use of the RNAsesy Mini kit (Qiagen, Hilden, Germany) according to the manufacturer’s instructions. Where indicated, before RNA isolation confluent cells were detached by treatment with 5 mM EDTA in PBS, washed twice with serum-free DMEM, resuspended in the same medium containing 1 \(\mu\)M monensin, and plated for 2 h on tissue culture dishes that had been coated with 10 \(\mu\)g/ml polylysine, fibronectin, or vitronectin as previously described (Retta et al., 1998). A multiplex semiquantitative RT-PCR was used to detect the relative levels of \(\beta_3\) and \(\beta_6\) or \(\beta_1\) integrin mRNAs. cDNA was synthesized from 5 \(\mu\)g of cytoplasmic RNA with the use of the 1st Strand cDNA Synthesis kit (Roche Molecular Biochemicals), and subjected to 28 (\(\beta_3/\beta_6\)) or 32 (\(\beta_6/\beta_1\) or \(\beta_1/\beta_6\)) PCR cycles. The reaction conditions and oligonucleotide PCR primers used were optimized so that the amplification products fell within the range of PCR amplification linearity. PCR was performed with each reaction mixture containing 5 \(\mu\)l of cDNA, 1 \(\times\) reaction buffer (Amersham Pharmacia Biotech AB), 1.5 mM Mg\(_2\)Cl\(_2\), 200 \(\mu\)M dNTP, 0.5 \(\mu\)M of each primer, and 2 \(\mu\)l of Taq DNA polymerase (Amersham Pharmacia Biotech AB) in a total volume of 50 \(\mu\)l. The following stages were used for each PCR cycle: 94°C for 30 s, 55°C for 30 s, and 72°C for 30 s, with a prolonged extension stage of 72°C for 5 min after the final cycle. The primers were derived from nonhomologous regions of the mouse \(\beta_6\) and \(\beta_1\) and the human \(\beta_1\) cDNA sequences, and led to 715-, 570-, and 857-bp PCR products, respectively. PCR products were electrophoresed on 2% agarose gels and stained with ethidium bromide. Gels were photographed under UV light and intensities of the amplified cDNA fragments were quantified with the use of a densitometric software (Molecular Analyst; Bio-Rad, Hemel Hempstead, United Kingdom). Molecular size standards (123-bp DNA ladder) were from Sigma.

For Northern blot hybridization, equal amounts of the purified total RNA (25 \(\mu\)g/lane) were separated by electrophoresis on a 1.2% agarose gel containing 1.8% formaldehyde and 1 \(\times\) FA Gel buffer [20 mM 3-(N-morpholino)propanesulfonic acid, 5 mM NaAc, 1 mM EDTA, pH 7.0], transferred to a Nytran SuPerCharge transfer membrane (Schleicher & Schuell, Dassel, Germany) with the use of the TurboBlotter blotting device according to manufacturer’s instructions (Schleicher & Schuell), and UV cross-linked to the membrane. The membrane was prehybridized by incubation in Church’s buffer (0.5 M Na-phosphate buffer, 10 mg/ml BSA, 7% SDS, 1 mM EDTA, 0.1 mg/ml salmon sperm DNA, pH 7.4) for 8 h at 65°C and hybridized with \(32^P\)-labeled probes overnight at 65°C in Church’s buffer. After hybridization, the membrane was washed once in 2\(\times\) SSC + 0.1% SDS, once in 1\(\times\) SSC + 0.1% SDS, once in 0.2\(\times\) SSC + 0.1% SDS, and once in 0.1\(\times\) SSC + 0.1% SDS for 15 min each at 65°C. The membrane was then exposed to x-ray film for 24–72 h at -80°C with an intensifying screen. Probes were synthesized by random priming with cDNA fragments of mouse \(\beta_6\) and \(\beta_1\) integrin subunits amplified by PCR and cloned in our laboratory. The same blots were rehybridized with a probe of the housekeeping gene \(\beta\)-actin to ensure equal loading.

Measurement of mRNA Stability

The measurement of mRNA stability was performed as described by Xu and Clark (1996). Briefly, cells were divided into three plates and cultured in 10% FBS/DMEM for 24 h before the addition of 60 \(\mu\)M 5,6-dichloro-1-\(\beta\)-d-ribofuranosyl-benzimidazole (DRB; Sigma), an inhibitor of transcription initiation. After addition of DRB, the cells were collected at 0, 4, 8, 12, and 24 h for RNA analysis. Total RNA isolation and Northern analysis were performed as described above.

Immunofluorescence Microscopy

Immunofluorescence studies were performed as described previously (Retta et al., 1996). Briefly, cells were seeded onto fibronectin-
coated glass coverslips and allowed to spread for 3 h in complete culture medium. Cells were then washed with cold PBS, fixed for 10 min with 3.7% paraformaldehyde in PBS, permeabilized with ice-cold 0.5% Triton X-100, 3.7% paraformaldehyde in PBS for 5 min, and incubated with 1% BSA in PBS for 30 min. To localize αV and β1 integrins, the cells were stained with the rabbit antisera to αV (rabbit anti-αV) and β1 (mAb TS2/16) integrin subunits. The αV and β1 antibody–antigen complexes were then detected with rhodamine-conjugated anti-rabbit and anti-mouse secondary antibodies, respectively. Representative fields were photographed with the use of an Olympus BM11 microscope fitted with epifluorescence. Notice that β1 integrins displace αV integrins from focal adhesions. Bar, 15 μm.

RESULTS

Expression of β1 Integrins Affects Subcellular Localization of αV Integrins

To determine the distribution of αV and β1 integrin heterodimers on GD25 and GD25-β1A cells attached to fibronectin, indirect immunofluorescence experiments with specific antibodies were performed. GD25 cells, which do not express β1 integrin heterodimers (Figure 1b), formed αV-containing prominent focal adhesions when allowed to attach and spread on coverslips coated with fibronectin (Figure 1a), consistent with the reported ability of αVβ3 to localize to focal adhesions in these cells (Wennerberg et al., 1996; Retta et al., 1998). In contrast, the amount of αV-containing focal adhesions was consistently reduced on GD25-β1A cells attached to fibronectin (Figure 1c), whereas β1A-containing focal adhesions were abundant (Figure 1d).

Thus, β1A, by localizing to focal adhesions, displaces the αV-containing heterodimers from these structures. Interestingly, we have previously shown that the expression of two other human β1 isoforms, namely, β1B, that does not localize to focal adhesions, and β1D, that is efficiently targeted to focal adhesions, also causes the delocalization of αV heterodimers on the cell surface (Belkin et al., 1997; Retta et al., 1998). Taken together, these data indicate that in GD25 cells cultured on fibronectin αVβ1 takes over the function of β1 integrins in mediating focal adhesion assembly; however, when expressed, β1 integrins behave as trans-dominant molecules with respect to αV integrins.
Expression of the $\beta_1$ Integrin Subunit in GD25 Cells
Induces Drastic Reduction of Surface Level of $\alpha_v\beta_3$ and an Up-Regulation of $\alpha_v\beta_5$

Previous results showed that transfection of GD25 cells with cDNA constructs of human $\beta_1$, integrin led to surface expression of the $\beta_1$ integrin subunit associated with the endogenous $\alpha_v$, $\alpha_5$, and $\alpha_v$ subunits, but not with the $\alpha_v$ subunit (Retta et al., 1998). In addition, no obvious differences in $\alpha_v$, integrin expression were seen by immunoprecipitation from $^{125}$I-surface-labeled GD25 and GD25-$\beta_1$A cells with an anti-$\alpha_v$ antisemur (Retta et al., 1998). To understand the cellular mechanism(s) controlling the effect of $\beta_1$ over $\alpha_v$, integrins, we analyzed more in detail GD25 and GD25-$\beta_1$A cells for the expression levels of their $\alpha_v$, integrin heterodimers, namely, $\alpha_v\beta_3$ and $\alpha_v\beta_5$. Untransfected or $\beta_1$A-transfected GD25 cells were surface-labeled with Sulfo-NHS-Biotin then $\alpha_v\beta_3$ and $\alpha_v\beta_5$ integrins were immunoprecipitated from nonionic detergent cell extracts and analyzed by Western blot. As expected, a polyclonal serum to the $\alpha_v$, integrin subunit coinmunoprecipitated $\alpha_v$, together with its associated $\beta_3$ and $\beta_5$ subunits (Figure 2A). The biotinylated $\alpha_v\beta_3$ and $\alpha_v\beta_5$ proteins resulted as distinct bands in Western blots and, surprisingly, we noticed that, whereas expression of the $\alpha_v$ subunit did not change significantly, the relative amounts of $\beta_3$ and $\beta_5$ proteins in $\beta_1$A-expressing GD25 cells were clearly different from those of untransfected GD25 cells (Figure 2A). With the use of antibodies specific for $\beta_3$ and $\beta_5$, subunits, we confirmed this evidence: $\beta_3$ protein levels were much lower in GD25-$\beta_3$,A than in GD25 cells, whereas the opposite was true for $\beta_5$ protein levels (Figure 2B and C). Thus, although the expression of the human $\beta_1$A integrin isoform in GD25 cells did not modify the surface expression level of the $\alpha_v$, integrin subunit, it led to a down-regulation and an up-regulation of the levels of its associated $\beta_3$ and $\beta_5$ subunits, respectively. These data suggest that the trans-dominant effect of $\beta_1$ integrin isoforms over the subcellular localization of $\alpha_v$, integrins in GD25-$\beta_1$ cells is due to a switching of the relative amounts of the cell-surface expression levels of $\alpha_v\beta_3$ and $\alpha_v\beta_5$ integrins.

Expression of $\beta_1$ Integrins Differentially Regulates mRNA Steady-State Levels of $\beta_3$ and $\beta_5$ Integrin Subunits

To address at what level the control of $\beta_3$ and $\beta_5$ protein expression in GD25-$\beta_1$A cells was exercised, we first compared mRNA steady-state levels of these two integrin subunits in GD25-$\beta_1$A with those of untransfected GD25 cells, with the use of both RT-PCR and Northern blot procedures.

A duplex RT-PCR assay with two sets of primers was developed for the simultaneous detection of the relative levels of $\beta_3$ and $\beta_5$ integrin subunit mRNAs. As shown in Figure 3A, a great difference in $\beta_5$, mRNA steady-state levels was observed between GD25 and GD25-$\beta_1$A cells. Interestingly, the lower mRNA levels of $\beta_5$, in GD25-$\beta_1$,A compared with those in GD25 cells reflected what we observed at the level of protein cell-surface expression (compare with Figure 2, A and B). On the contrary, there was little difference in $\beta_3$, mRNA levels among untransfected and $\beta_1$,A-transfected GD25 cells, with a small elevation observed in GD25-$\beta_1$A cells (Figure 3A). RT-PCR for $\beta_3$, mRNA was performed as control (Figure 3B). Thus, the presence of $\beta_1$, integrins in GD25 cells differently modulates $\beta_3$ and $\beta_5$ mRNA expression.

Northern blot analysis demonstrated that our cDNA probes to $\beta_3$ and $\beta_5$, specifically recognized mRNAs of ~6.6 and 3.5 kb, respectively, consistent with what has been previously described (Yamada et al., 1995). As resulted from this analysis, the mRNA steady-state level of $\beta_3$, was much lower in GD25-$\beta_3$,A than in GD25 cells (Figure 3C, $\beta_3$), thus reflecting the difference observed by RT-PCR and protein analysis (see above). In addition, the Northern blot analysis confirmed that the difference in $\beta_5$, mRNA steady-state levels between GD25 and GD25-$\beta_1$A cells (Figure 3C, $\beta_5$) did not fully correlate with the difference in $\beta_5$, cell-surface expression level (compare with Figure 2, A and C). Thus, in GD25-$\beta_1$A cells the down-regulation of $\alpha_v\beta_3$, cell-surface expression strictly correlates with the down-regulation of $\beta_3$, mRNA steady-state level, whereas the up-regulation of $\alpha_v\beta_5$, is mainly due to translational or posttranslational events leading to an increase of $\beta_5$, subunit cell-surface recruitment.
equal amounts of total RNA (25 μg/lane) were probed sequentially by 32P-labeled mouse integrin α5 mRNA expression levels. Notice that the presence of β1 integrins causes a marked down-regulation of β3 and a little increase of β5 mRNA expression levels.

**Figure 3.** β1 integrins differentially regulate mRNA steady-state levels of β3 and β5 integrin subunits. Total RNA was isolated from 1 × 10^7 cultured GD25 (−/−) and GD25-β1A cells, and β5 and β5 mRNAs were evaluated by RT-PCR and Northern blot analyses as described in MATERIALS AND METHODS. (A) Duplex RT-PCR assay for the simultaneous detection of β5 and β5 mRNAs. (B) RT-PCR for β3 mRNA performed as control. Molecular size standards (123-bp DNA ladder) are shown on the left. (C) Northern blot: equal amounts of total RNA (25 μg/lane) were probed sequentially by 32P-labeled mouse integrin β1 and β5 cDNA fragments, and by a 32P-labeled β-actin probe as a control for RNA loading. Notice that the presence of β1 integrins causes a marked down-regulation of β3 and a little up-regulation of β5 mRNA expression levels.

**β1 Effect over α5 Integrins Occurs Irrespective of the Type of β1 Isoform and Is Dependent on the Presence of the β1 Cytoplasmic Domain Common Region**

We have previously characterized some of the functional properties of β1B and β1D integrin isoforms, comparing these properties with those of the common β1A isoform. In particular, we have shown that the unique cytoplasmic sequences of β1B and β1D endow these molecules with distinctive functional properties with respect to a number of cellular functions (Balzac et al., 1994; Belkin et al., 1997; Belkin and Retta, 1998; Calli et al., 1998; Retta et al., 1998).

To analyze more in detail the effects of β1 over β3 and β5 integrins, we tested GD25 cells expressing β1B and β1D isoforms as well as two β1 deletion mutants lacking almost the entire cytoplasmic domain (β1A) or the cytoplasmic domain variable region (β1-D, or β1-D, or β1-A, or β1-A). We found that β1A cells, whereas in GD25-β1TR, we found that in GD25-B1A and GD25-β1D cells it was as low as in GD25-β1-A cells, whereas in GD25-β1-B, it was also reduced but to a lower extent (Figure 5, A–C). On the contrary, the β1 mRNA level was higher and similar to that of β1-deficient GD25 cells (Figure 5, A–C). On the other hand, although a little increase of β5 mRNA steady-state level was observed in GD25 cells expressing β1B, β1D, or β1-A, it was not fully reflected in the high increase observed at the β5 protein level in the same cells.

These results indicate that the β1-dependent modulation of β3 and β5 integrin subunit expression was not confined to GD25-β1A cells, but it was also present in GD25 cells expressing two other β1 isoforms. In addition, the fact that a down-regulation of β3 and an up-regulation of β5 was also observed in GD25-B1TR, but not in GD25-β1B cells, strongly suggests that the control of the expression level of β3 and β5 integrin subunits was dependent on the presence of the β1 cytoplasmic domain common region.

**Cell Adhesion to ECM Proteins Is Not Required for β1 Effect on β1 mRNA Steady-State Level**

To determine whether cell adhesion to ECM proteins was required for β1 effect over β3 expression levels, we performed Northern blot analysis of β3 mRNA steady-state level in cells plated on tissue culture dishes coated with either polyelectrolyte or two ECM proteins, namely, fibronectin and vitronectin. GD25-β1A, TR, and GD25-β1-B, COM cells were cultured to confluence in complete culture medium and then resuspended in serum-free medium, containing 0.1 μM mimosine, and allowed to attach and spread on collagen-, fibronectin, and vitronectin-coated dishes for 2 h at 37°C before RNA isolation for Northern blot analysis. The results, shown in Figure 6, indicate that the β1 mRNA steady-state level was constitutively low in GD25-β1A cells compared well with our subsequent mRNA analysis. In fact, when we analyzed by RT-PCR and Northern blot the mRNA steady-state level of the β3 subunit in GD25 cells expressing either β1B, β1D, β1-A, or β1-B, TR, we found that in BD25-β1A and BD25-β1D cells it was as low as in BD25-β1-A cells, whereas in BD25-β1-B, COM cells it was also reduced but to a lower extent (Figure 5, A–C). On the contrary, the β1 mRNA level was higher and similar to that of β1-deficient BD25 cells (Figure 5, A–C). On the other hand, although a little increase of β5 mRNA steady-state level was observed in BD25 cells expressing β1B, β1D, or β1-A, it was not fully reflected in the high increase observed at the β5 protein level in the same cells.

Because human β1 expression in BD25 cells leads to the assembly and cell-surface recruitment of integrin complex
Expression of $\beta_1$ integrins induces a marked decrease in $\beta_1$ mRNA stability

Because modulation of mRNA stability is a potential regulatory mechanism for integrin expression (Sachs, 1993; Feng et al., 1999), we next asked whether the changes in mRNA steady-state levels were due to changes in integrin mRNA stability. The rate of turnover of $\beta_1$ and $\beta_3$ mRNAs was determined by inhibition of RNA synthesis with 60 $\mu$M DRB followed by quantitative blot hybridization analysis of $\beta_3$ and $\beta_5$ mRNA as a function of time. In GD25-$\beta_1$A cells grown on tissue culture dishes a clear decrease of $\beta_3$ mRNA stability was detected compared with GD25 cells (Figure 7). In contrast, the stability of $\beta_5$ mRNA was much higher than that of $\beta_3$, and no significant difference was observed when GD25 and GD25-$\beta_1$A cells were compared. Therefore, the effects of $\beta_1$ expression over $\beta_3$ and $\beta_5$ mRNA levels clearly involve a regulation of $\beta_3$, but not $\beta_5$, mRNA stability.

DISCUSSION

There is increasing evidence that a coordinated cross talk between integrin receptors is crucial for an integrated and functional response of a single cell to the extracellular environment (Porter and Hogg, 1997; Blystone et al., 1999; Hynes, 1999). However, the molecular mechanisms of integrin cross talk remain mostly undetermined.

Previously, we showed that the expression of either $\beta_1$ or $\beta_5$ integrin isomers in $\beta_1$-null GD25 cells prevented different FN-dependent functions of endogenous $\alpha_\gamma$ integrins, including their ability to mediate cell adhesion, to localize to focal adhesions, and to assemble an FN matrix, thus indicating the existence of a functional cross talk between these two $\beta_1$ isomers and $\alpha_\gamma$ integrins (Belkin et al., 1997; Retta et al., 1998). The present study was undertaken to examine this integrin cross talk and establish the regulatory mechanism(s) whereby $\beta_1$ integrins exert their trans-acting functions. The main findings are that 1) de novo expression of the $\beta_1$ integrin subunit in $\beta_1$-null GD25 cells induces a

with endogenous $\alpha_\gamma$, $\alpha_5$, and $\alpha_6$ subunits (Retta et al., 1996), it was possible that these $\beta_1$-associated $\alpha$ subunits could play a direct role in the down-regulation of $\alpha_\gamma$ integrin. To exclude this possibility we took advantage of GD25-$\beta_1$TR cells by extending our observations with this cell line. The expression of the $\beta_1$ TR mutant at the surface of GD25-$\beta_1$TR cells was comparable with that of the $\beta_1$A isoform in GD25-$\beta_1$A cells, as previously determined by flow cytometry analysis (Retta et al., 1998). In addition, by immunoprecipitation experiments we did not see any detectable change in the pattern of $\alpha$ subunits associated with $\beta_1$TR in GD25 cells compared with GD25-$\beta_1$A cells (Figure 7). Nevertheless, the presence of $\beta_1$TR integrin heterodimers did not lead to any detectable effect over $\alpha_\gamma$ or $\alpha_5$ integrins (Figures 4, A–C, and 5, A–D). Taken together, these data strongly suggest that the down-regulation of $\beta_3$ is not directly due to de novo surface expression of the $\beta_1$-associated $\alpha$ subunits and confirm that for the effect of $\beta_1$ integrins over $\alpha_\gamma$/$\alpha_5$ integrin ratio a $\beta_1$ subunit carrying, at least, the common region of the cytoplasmic domain is required.
drastic down-regulation of \( \alpha_v \beta_3 \) and an up-regulation of \( \alpha_v \beta_5 \) integrin cell surface levels; 2) this \( \beta_1 \) effect occurs irrespective of the type of \( \beta_1 \) isoform but is dependent on the presence of the common region of the \( \beta_1 \) cytoplasmic domain; and 3) the down-regulation of \( \alpha_v \beta_3 \) is due to a decreased mRNA stability of the \( \beta_3 \) subunit, whereas the up-regulation of \( \alpha_v \beta_5 \) is mainly due to translational or posttranslational events. These findings provide the first evidence of a cross talk between \( \beta_1 \) and \( \alpha_v \) integrins based on mechanisms of control of mRNA and protein levels.

Figure 5. Comparative analysis of \( \beta_3 \) and \( \beta_5 \) mRNA steady-state levels in GD25 cells expressing different integrin \( \beta_1 \) forms. Total RNA was isolated from \( 1 \times 10^7 \) cultured cells as described in MATERIALS AND METHODS. (A) Multiplex RT-PCR assay for the simultaneous detection of \( \beta_1^A \), \( \beta_1^B \), and \( \beta_1^D \) mRNAs in GD25 cells expressing either the \( \beta_1^A \), \( \beta_1^B \), or \( \beta_1^D \) isoforms or two \( \beta_1 \) deletion mutants lacking the entire cytoplasmic domain (\( \beta_1^{TR} \)) or the cytoplasmic domain variable region (\( \beta_1^{COM} \)). (B) Northern blot: equal amounts of total RNA (25 \( \mu \)g/lane) were probed sequentially by \( ^32 \)P-labeled mouse integrin \( \beta_3 \) and \( \beta_5 \) cDNA fragments and by a \( ^32 \)P-labeled \( \beta \)-actin probe as a control for RNA loading. The positions of 28s and 18s rRNAs are indicated as markers for RNA sizes. (C and D) Scanning densitometry analysis of \( \beta_3 \) and \( \beta_5 \) mRNA levels as detected by Northern blot. Northern signals were normalized to \( \beta \)-actin and displayed as percentage of the control (GD25). Data are representative of three independent experiments. Notice that the down-regulation of \( \beta_3 \) mRNA steady-state level occurs irrespective of the type of the \( \beta_1 \) isoform and is dependent on the presence of the \( \beta_1 \) cytoplasmic domain common region.

Vol. 12, October 2001 3133
Figure 6. Cell adhesion to ECM proteins is not required for β3 effect on β3 mRNA steady-state level. GD25-β3TR and GD25-β3COM cells were cultured to confluence in complete culture medium. Cells were then resuspended in serum-free medium containing 1 μM monensin and allowed to attach and spread on polylysine- (PL), FN-, or vitronectin (VN)-coated tissue culture dishes for 2 h at 37°C before lysis. Total RNA was isolated as described in MATERIALS AND METHODS, and equal amounts (25 μg/lane) were analyzed for β3 mRNA steady-state level by Northern blot hybridization with the use of 32P-labeled mouse integrin β3 cDNA fragments as probe. Equal loading was confirmed by hybridization of the same blot with a 32P-labeled probe for β-actin. The positions of 28s and 18s RNAs are indicated as markers for RNA sizes. Notice that the β3 mRNA steady-state level is constitutively low in GD25-β3COM compared with that of GD25-β3TR cells.

Expression of the β3 Integrin Subunit in GD25 Cells Induces a Drastic Reduction of Surface Level of α,Vβ3 and an Up-Regulation of α,Vβ5

Despite the apparent high degree of integrin-ligand binding redundancy (Hynes, 1992), the localization of distinct integrins to focal adhesions is usually very restricted (Fath et al., 1989). In GD25 cells two integrins are believed to be able to localize to focal adhesions on fibronectin, namely, α,Vβ3 and, upon β3 ectopic expression, α,Vβ1 (Wennerberg et al., 1996; Belkin et al., 1997; Retta et al., 1998). However, whereas α,V integrins can take over some FN-dependent functions in the absence of β3 integrins (Wennerberg et al., 1996; Retta et al., 1998), immunofluorescence analyses of β3A-transfected and untransfected GD25 cells plated on fibronectin show that β3, integrins clearly dominate upon α,V integrins in localizing to focal adhesions. This phenomenon is mainly due to a cross talk between β3 and α,V integrins that occurs at the level of expression on the cell surface. In fact, although the presence of β3 integrins in GD25 cells does not affect the total amount of α,V integrins, it causes a clear rearrangement of the relative cell surface levels of β3 and β5 subunits, leading to a marked down-regulation of α,Vβ3 and a correspondent up-regulation of α,Vβ5.

A great deal of experimental work has shown that integrin expression is highly dynamic during development (reviewed in Darribere et al., 2000; Tarone et al., 2000). In particular, it has been suggested that the presence and functions of the α,V integrins are developmentally controlled by differential temporal and spatial regulation of its β subunits (Yamada et al., 1995), whereas there are reports showing that a balanced ratio of integrin receptors is crucial for the maintenance of the differentiation state of a particular cell (Carroll et al., 1985; Sastry et al., 1996). However, genetic ablation experiments have shown that the absence of some widely expressed integrins that were believed to be key regulators of development and differentiation has resulted into mild or late phenotypes. In particular, it has come out that processes such as myogenesis, vasculogenesis, and angiogenesis, which through antibody or peptide perturbation experiments were shown to be dependent on specific β3 or α,V integrins, can actually proceed without these integrins (Bader et al., 1998; Hirsch et al., 1998), suggesting that there might be some overlapping or compensatory functions between different integrins. A very striking example of this point comes from a thorough study of compound mutations showing that gene knockouts of α,V and α,β3 integrin subunits have synergistic effects when combined pairwise and result in a phenotype similar to that of FN-null mutation, suggesting that α,Vβ3 and α,V integrins normally overlap or can compensate each other in mesodermal development (Yang et al., 1999). Other examples derive from in vitro experiments showing that in cultured cells α,V integrins are able to compensate for the loss of the α,Vβ3 fibronectin receptors (Wennerberg et al., 1996; Yang and Hynes, 1996; Retta et al., 1998). On the other hand, α,Vβ3 has previously been shown to negatively regulate α,Vβ3-mediated cell migration (Bilato et al., 1997; Simon et al., 1997; Blystone et al., 1999) and phagocytosis (Blystone et al., 1994, 1999). Notably, the above-cited reports are all examples of functional cross talk between α,V and β3 integrins, apparently without quantitative up-regulation of integrin levels. Our present report now demonstrates that, besides the existence of a functional compen-
Cross Talk between $\beta_1$ and $\alpha_\nu$ Integrins

The expression of integrins can be modulated by a variety of agents, including proinflammatory cytokines, growth factors, hormones, extracellular matrix components, and pharmacological agents (Delcommenne and Streuli, 1995; Kim and Yamada, 1997). In particular, the integrin $\alpha_\nu\beta_1$ has been shown to be up-regulated by transforming growth factor-$\beta_1$, platelet-derived growth factor-BB (Janat et al., 1992), basic fibroblast growth factor (Sepp et al., 1994), vitamin D (Medhora et al., 1993), fibronectin (Feng et al., 1999), and phorbol esters (Swerlick et al., 1992), and down-regulated by tumor necrosis factor-$\alpha$, interferon-$\gamma$ (Dellipheri et al., 1991), and collagen (Feng et al., 1999). Mechanisms regulating integrin expression include regulation of protein levels by transcriptional or posttranscriptional events, alternative splicing of mRNA, and mobilization of preexisting intracellular stores (Xu and Clark, 1996; Brakebusch et al., 1997; Bader et al., 1998; Hirsch et al., 1998; McHugh et al., 2000) could be explained by taking into account that a compensatory up-regulation both in level and function between integrins can occur.

Figure 8. The presence of the $\beta_1$ cytoplasmic domain doubles the decay rate of the $\beta_3$ mRNA in GD25 cells. GD25-$\beta_1$A and GD25-$\beta_1$TR cells grown to confluence were divided into three 10-cm Petri dishes and cultured in DMEM containing 10% FBS for 24 h before the addition of 60 $\mu$M DRB, an inhibitor of transcription initiation. After addition of DRB, the cells were collected at 0, 4, and 8 h for RNA analysis. Total RNA isolation and Northern analysis were performed as described in MATERIALS AND METHODS. (A) Northern blot: equal amounts of total RNA (25 $\mu$g/lane) were probed sequentially by $^32$P-labeled mouse integrin $\beta_1$ and $\beta$-actin cDNA fragments. (B) Scanning densitometry analysis of $\beta_3$ mRNA levels as detected by Northern blot. $\beta_1$ Northern signals were normalized to $\beta$-actin and displayed as percentage of the baseline (time 0). Data presented are the mean values $\pm$ SE of three independent experiments. Notice the lower stability of $\beta_1$, integrin subunit mRNA in GD25-$\beta_1$A than in GD25-$\beta_1$TR cells.

Expression of $\beta_1$ Integrins Differentially Regulates mRNA Steady-State Levels of $\beta_3$ and $\beta_5$ Integrin Subunits

The expression of integrins can be modulated by a variety of agents, including proinflammatory cytokines, growth factors, hormones, extracellular matrix components, and pharmacological agents (Delcommenne and Streuli, 1995; Kim and Yamada, 1997). In particular, the integrin $\alpha_\nu\beta_1$ has been shown to be up-regulated by transforming growth factor-$\beta_1$, platelet-derived growth factor-BB (Janat et al., 1992), basic fibroblast growth factor (Sepp et al., 1994), vitamin D (Medhora et al., 1993), fibronectin (Feng et al., 1999), and phorbol esters (Swerlick et al., 1992), and down-regulated by tumor necrosis factor-$\alpha$, interferon-$\gamma$ (Dellipheri et al., 1991), and collagen (Feng et al., 1999). Mechanisms regulating integrin expression include regulation of protein levels by transcriptional or posttranscriptional events, alternative splicing of mRNA, and mobilization of preexisting intracellular stores (Xu and Clark, 1996; Brakebusch et al., 1997; Bader et al., 1998; Hirsch et al., 1998; McHugh et al., 2000) could be explained by taking into account that a compensatory up-regulation both in level and function between integrins can occur.
availability of the α subunit as a rate-limiting step in integrin complex assembly and cell-surface expression (Swerlick et al., 1992).

The regulation of mRNA stability is a very important mechanism of posttranscriptional regulation of gene expression, and evidence exists for both a wide range of half-lives for different mRNAs in the same cells and different half-lives for the same mRNA in the same cell under different circumstances (Sachs, 1993). Interestingly, it has been previously suggested that coordinate signals from ECM molecules and growth factors can modulate the mRNA decay rate of specific integrins (Xu and Clark, 1996). In addition, a recent report shows that the β3 mRNA stability can be increased by cell interaction with fibrin but not with collagen (Feng et al., 1999). Because most extracellular matrix proteins signal through integrins, which have also been shown to physically associate and act synergistically with growth factor receptors (Giancotti and Ruoslahti, 1999), it is possible to hypothesize a scenario where the expression of a specific integrin can influence the expression of another integrin by affecting its mRNA stability, either directly or with the cooperation of an associated growth factor receptor. This could be a way for a rapid change of integrin ratios in response to a variation of the extracellular environment, as it occurs during tissue formation or repair. Interestingly, αvβ3 expression has been shown to increase focally and transiently during cutaneous wound repair (Feng et al., 1999). On the other hand, the existence of this regulatory integrin cross talk could explain the compensatory up-regulation of αvβ3 in the absence of β1 integrins, highlighting the usefulness of stabilizing specific integrin mRNA only when it is needed.

**β1 Effect over β3 Expression Is Dependent on the Presence of the β1 Cytoplasmic Domain Common Region**

Our results clearly demonstrate that the control of β1/β3 ratio is β1-dependent, and neither confined to a particular cell population nor restricted to a specific β1 isofrom. Instead, it requires the presence of the common region of the β1 cytoplasmic domain. In addition, the fact that the β1-COM mutant does not contribute to cell adhesion (Retta et al., 1998), together with the observation that the β1 mRNA steady-state level is constitutively low in GD25-β1COM cells compared with GD25-β1TR cells, indicates that the binding of extracellular ligands is not required for β1 to regulate β3 expression. On the other hand, it is noteworthy that, although in long-term adherent cells the β1-COM mutant was less effective than β1A in inducing an effect over the β3 mRNA steady-state level, no significant difference was observed when we compared β3 mRNA levels in GD25-β1COM and GD25-β1A cells either plated on polylysine or kept in suspension for 2 h (unpublished). These results could be explained by taking into account that, in contrast with the expression of β1A, the expression of β1-COM in GD25 cells does not entirely prevent the localization of αv integrins to FN-dependent focal adhesions (Retta et al., 1998). Thus, a likely possibility is that the common region of the β1 cytoplasmic domain is able to constitutively induce a down-regulation of β3 mRNA steady-state level; however, the αvβ3 integrin, due to its ability to localize to focal adhesions in GD25-β1COM cells, counteracts this β3 constitutive action leading to a mild effect on β3 expression level. Interestingly, in accordance with a recent report (Feng et al., 1999), the higher β3 mRNA level in cells cultured on fibronectin or vitronectin than on polylysine (Figure 6) suggests that ECM proteins that are ligands for αvβ3 can sustain β3 mRNA steady-state level.

In conclusion, our results indicate a novel mechanism of integrin cross talk where one integrin can regulate the expression of another by modulating the decay rate of its mRNA. The biological implications of this integrin cross talk are potentially of high functional significance as a fine-tuned mechanism for selective and transient integrin expression in different extracellular contexts. Our attempt now will be to uncover β3-dependent events regulating the β3 mRNA steady-state levels.

**ACKNOWLEDGMENTS**

We thank Reinhard Fässler and Arnoud Sonnenberg for generously providing GD25 cells and mAb GoH3, respectively. The students Tiziana Spatola and Federica Logrand are gratefully acknowledged for helping in some experiments. We also thank Fiorefia Balzac and Emilio Hirsch for helpful discussions. This work was supported by grants from the Italian Association for Cancer Research to G.L., from the University of Torino (ex 60%) to SFR and GT and from the University of Palermo (ex 60%) to GDL.

**REFERENCES**


Blystone, S.D., Slater, S.E., Williams, M.P., Crow, M.T., and Brown, E.J. (1999). Molecular mechanism of integrin crosstalk: αvβ3 sup-


